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(54) Title: METHOD OF DIRECTING BIOSYNTHESIS OF SPECIFIC POLYKETIDES

(57) Abstract

A method to produce novel polyketide structures by designing and introducing specified changes in the DNA governing the synthesis of the polyketide is disclosed. The biosynthesis of specific polyketide analogs is accomplished by genetic manipulation of a polyketide-producing microorganism by isolating a polyketide biosynthetic gene-containing DNA sequence, identifying enzymatic activities associated within the DNA sequence, introducing one or more specified changes into the DNA sequence which codes for one of the enzymatic activities which results in an altered DNA sequence, introducing the altered DNA sequence into the polyketide-producing microorganism to replace the original sequence, growing a culture of the altered microorganism under conditions suitable for the formation of the specific polyketide analog, and isolating the specific polyketide analog from the culture. The method is most useful when the segment of the chromosome modified is involved in an enzymatic activity associated with polyketide biosynthesis, particularly for manipulating polyketide synthase genes from *Saccharapolyspora* or *Streptomyces*.

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METHOD OF DIRECTING BIOSYNTHESIS OF SPECIFIC POLYKETIDES

Field of the Invention.

The present invention relates to a method for directing the biosynthesis of specific polyketide analogs by genetic manipulation. In particuar, polyketide biosynthetic genes are manipulated to produce precise, novel polyketides of predicted structure.

Background of the Invention

Polyketides are a large class of natural products that includes many important antibiotics and immunosuppressants such as erythromycins, tetracyclines, and rapamycins. Their synthesis proceeds by an ordered condensation of acyl esters to generate carbon chains of varying length and substitution pattern that are later converted to mature polyketides. This process has long been recognized as resembling fatty acid biosynthesis, but with important differences. Unlike a fatty acid synthase, a typical polyketide synthase is programmed to make many choices during carbon chain assembly: For example, the choice of "starter" and "extender" units, which are often selected from acetate, propionate or butyrate residues in a defined sequence. The choice of using a full cycle of reduction-dehydration-reduction after some condensation steps, omitting it completely, or using one of two incomplete cycles (reduction alone or reduction followed by dehydration), which determines the pattern of keto or hydroxyl groups and the degree of saturation at different points in the chain is additionally programed. Finally the choice of stereochemistry for the substituents at many of the carbon atoms is programmed by the polyketide synthase.

Because of the commercial significance of *Streptomyces*, a great amount of effort has been expended in the study of *Streptomyces* genetics. Consequently much is known about *Streptomyces* and several cloning vectors exist for transformations of the organism.

Although many polyketides have been identified, there remains the need to obtain novel polyketide structures with enhanced properties. Current methods of obtaining such molecules include screening of natural isolates and chemical modification of existing polyketides, both of which are costly and time consuming. Current screening methods are based on gross properties of the molecule, i.e. antibacterial, antifungal

activity, etc., and both *a priori* knowledge of the structure of the molecules obtained or predetermination of enhanced properties are virtually impossible. Chemical modification of preexisting structures has been successfully employed, but it still suffers from practical limitations to the type of compounds obtainable, largely connected to the poor yield of multistep syntheses and available chemistry to effect modifications. The following modifications are extremely difficult or inefficient at the present time: change of the stereochemistry of the side chains in the completed polyketide; change of the length of the polyketide by removal or addition of carbon units from the interior of the acyl chain; and dehydroxylation at unique positions in the acyl chain. Accordingly, there exists the need to obtain molecules wherein such changes can be specified and performed and would represent an improvement in the technology to produce altered polyketide molecules with predicted structure.

Summary of the Invention

The present invention provides a method to produce novel structures from designing and introducing specified changes in the DNA governing the synthesis of the polyketide. According to the method of the present invention, the biosynthesis of specific polyketide analogs is accomplished by genetic manipulation of a polyketide-producing microorganism comprising the steps of:

(1) isolating a polyketide biosynthetic gene-containing DNA sequence;

(2) identifying enzymatic activities associated within said DNA sequence;

(3) introducing one or more specified changes into said DNA sequence which codes for one of said enzymatic activities which results in an altered DNA sequence;

(4) introducing said altered DNA sequence into the polyketide-producing microorganism to replace the original sequence;

(5) growing a culture of the altered microorganism under conditions suitable for the formation of the specific polyketide analog; and

(6) isolating said specific polyketide analog from the culture.

The present method is most useful when the segment of the chromosome modified is involved in an enzymatic activity associated with polyketide biosynthesis. The present invention is especially useful

in manipulating polyketide biosynthetic genes from *Streptomyces*, an organism which provides over one-half of the clinically useful antibiotics.

Brief Description of the Drawings

FIG. 1 illustrates the organization of gene encoding polyketide synthase and designated *eryA* as follows: (a) Map coordinates of the DNA; (b) DOTPLOT of the output of COMPARE (window = 50, stringency = 32) program (Sequence Analysis Software Package of the Genetics Computer Group, University of Wisconsin, Biotechnology Center, 1710 University Avenue, Madison, Wisconsin 53705 of *eryA* segment (x-axis) vs. subsegment of *eryA* comprises between 23 - 27.5 sequence coordinates (y-axis) [see Fig. 2]; (c) Open reading frame organization of *eryA* and enzymatic activities encoded. PT = propionyltransferase; ACP = acyl carrier protein; KS=β-ketoacyl ACP synthase; RmT = (2R) methylmalonyl CoA transferase; KR =β-ketoreductase; SmT = (2S) methylmalonyl CoA transferase; DH = dehydratase; ER = enoylreductase; TE = thioesterase; and (d) Schematic diagram showing the extent of each of the six modules in *eryA*.

FIG. 2. illustrates the nucleotide sequence of *eryA* with corresponding translation of the three open reading frames. Standard one letter codes for the amino acids appear beneath their respective nucleic acid codons. The standard one letter codes for the amino acid sequences are as follows:

A	-alanine
R	-arginine
N	-asparagine
D	-aspartic acid
C	-cysteine
Q	-glutamine
E	-glutamic acid
G	-glycine
H	-histidine
I	-isoleucine
L	-leucine
K	-lysine
M	-methionine (start)
F	-phenylalanine

P	-proline
S	-serine
T	-threonine
W	-tryptophan
Y	-tyrosine
V	-valine

FIG. 3. is a schematic representation of Type I, Type II and Type III changes in *eryA* and structures of corresponding novel polyketides produced. Δ69 (Type I) and Δ33 (Type II) represent in-frame deletions of the base pairs in the DNA segments corresponding to the KR of module 2 and the β–ketoacyl ACP synthase of module 2, respectively. Insertion of a complete copy of module 4 within module 1 is also shown. Production of 11-epifluoro-15-norerythromycin in strain that carries Δ33 occurs when substrate analog (2S,3S,4S,5S)2,4-dimethyl-3-fluoro-5-hydroxyhexanoic acid-ethyl thioester is fed.

FIG. 4 illustrates the restriction site coordinates of cosmid pR1 5' to the sequence of *eryA* (Fig 2).

Detailed Description of the Invention

For the purposes of the present invention as disclosed and claimed herein, the following terms are defined.

The term "polyketide" as used refers to a large and diverse class of natural products, including antibiotics, pigments, and immunosuppressants. Antibiotics include, but are not limited to anthracyclines, tetracyclines, polyethers, ansamycins, macrolides of different types (polyenes and avermectins as well as classical macrolides such as erythromycins).

The term "polyketide-producing microorganism" as used herein includes any *Actinomycetales* which can produced a polyketide. Examples of Actinomycetes that produce polyketides include but are not limited to *Micromonospora rosaria, Micromonospora megalomicea, Sacharapolyspora erythraea, Streptomyces antibioticus, Streptomyces albireticuli, Streptomyces ambofasciens, Streptomyces avermitilis, Streptomyces fradiae, Streptomyces hygroscopicus, Streptomyces tsukubaensis, Streptomyces griseus, Streptomyces mycarofasciens, Streptomyces platensis, Streptomyces venezuelae, Streptomyces*

violaceoniger, and various *Actinomadura*, *Dactylosporangium* and *Nocardia* strains that produce polyether type of polyketides.

The term "polyketide synthase" as used herein refers to the complex of enzymatic activities responsible for the biosynthesis of polyketides which include but are not limited to β-ketoreductase, dehydratase, acyl carrier protein, enoylreductase, β-ketoacyl ACP synthase, and acyltransferase.

The term "extender" as used herein refers to a coenzyme A thioester of a dicarboxylate which is incorporated into a polyketide by a polyketide synthase.

The term "starter" as used herein refers to a coenzyme A thioester of a carboxylic acid which is used by the polyketide synthase as the first building block of the polyketide.

The term "*eryA*" as used herein refers to the genes involved in the formation of the polyketide moiety of erythromycin.

The term "condensation" as used herein refers to the addition of an extender unit out to the nascent polyketide chain and requires the action of β-ketoacyl ACP synthase, acyltransferase, and acyl carrier protein.

The term "β-carbonyl processing" as used herein refers to changes effecting the carbonyl group of the growing polyketide via β-ketoreductase, dehydratase, and enoylreductase.

The term "module" as used herein refers to the genetic element encoding one condensation step, as defined above, and one β-carbonyl processing step, as defined herein.

The term "Type I change" as used herein refers to changes in DNA sequence which will result in the production of polyketide rings of length identical to that of 6-deoxyerythronolide A, but with altered functional groups at specific ring positions.

The term "Type II change" as used herein refers to alterations which will result in the production of macrolide rings only when fed exogenously with substrate analogs, e.g.thioesters of appropriate acyl compounds of various length. Thus Type II mutants are erythromycin non-producing (Ery^-) mutants. The structure of the resulting macrolides will depend on the substrate employed.

The term "Type III change" as used herein refers to alterations which will result in the biosynthesis of macrolide rings of length reduced (deletion) or increased (insertion) by two carbon units, or macrolide rings altered in specific portions of the chain (replacement).

In its broadest sense, the present invention entails a general procedure for producing novel polyketide structures *in vivo* by selectively altering the genetic information of the organism that naturally produces a related polyketide. A set of examples described herein are a series of novel polyketides that make use of the genetic information for the biosynthesis of the polyketide portion of the macrolide antibiotic erythromycin. The organization of the segment of the *Saccharapolyspora erythraea* chromosome, designated *eryA*, and the corresponding polypeptides which it encodes that determine the biosynthesis of the polyketide segment of erythromycin, are shown in FIG. 1. It is seen that *eryA* is organized in modules, as shown, and that each module takes care of one condensation step, through the action of the β–ketoacyl ACP synthase specified within, wherein an extender unit, methylmalonyl CoA, is added first to the starter unit, propionyl CoA, and then to the successively growing acyl chain. The precise succession of elongation steps is dictated by the genetic order of the six modules: module 1 determines the first condensation; module 2, the second; module 3, the third, and so on until the sixth condensation step has occurred. Furthermore, the processing of the growing chain after each condensation is also determined by the information within each module. Thus β-ketoreduction of the β-carbonyl takes place after each step except for step 3, as determined by the presence of a functional β-ketoreductase in all modules except module 3, whereas dehydration and enoylreduction only take place after the fourth extender unit is added to the growing acyl chain, as determined by the presence of dehydratase and enoylreductase in module 4. Furthermore, the choice of the correct enantiomer (2R or 2S) of methylmalonyl-CoA as the extender unit employed at each condensation is specified by the acyltransferase function determined by each module (FIG. 1C).

In the present invention, novel polyketide molecules of desired structure are produced by the introduction of specific genetic alterations of the *eryA* sequence into the *Sac. erythraea* chromosome. The complete nucleotide sequence of the *eryA* segment of the *Sac. erythraea* chromosome and the sequence of the corresponding polypeptides are shown in FIG. 2. Three types of alterations to the *eryA* DNA sequence are described: (i) those inactivating a single function in a module which does not arrest acyl chain growth (β-ketoreductase, dehydratase, or enoylreductase); (ii) those inactivating a single function in a module

which does arrest chain growth (β–ketoacyl ACP synthase, acyltransferase or acyl carrier protein); and (iii) those affecting an entire module (deletion, insertion, or replacement). The novel polyketides produced by strains carrying these types of mutations can be classified accordingly. Type I changes will result in the production of polyketide rings of length identical to that of 6-deoxyerythronolide A, but with altered functional groups at specific ring positions. Strains carrying type II alterations will result in the production of macrolide rings only when fed exogenously with substrate analogs, e.g.thioesters of appropriate acyl compounds of various length. Thus Type II mutants are erythromycin non-producing (Ery^-) mutants. The structure of the resulting macrolides will depend on the substrate employed. Type III changes will result in the biosynthesis of macrolide rings of length reduced (deletion) or increased (insertion) by two carbon units, or macrolide rings altered in specific portions of the chain (replacement). A schematic representation of some examples of Type I, Type II and Type III alterations in *eryA* and the corresponding novel polyketides produced in hosts that carry such alterations is shown in FIG. 3.

In the examples described herein, specific mutations in the *eryA* region of the *Sac. erythraea* chromosome are introduced by a simple two-step approach: 1) introduction of a specified change in a cloned DNA segment; 2) exchange of the wild type allele with the mutated one. Step 1 requires standard recombinant DNA manipulations employing *E. coli* as the host. Step 2 requires one or more plasmids out of the several *E. coli-Sac. erythraea* shuttle vectors available and a simple screening procedure for the presence of the colony carrying the altered gene. Two methods are used to introduce the altered allele into the chromosome to replace the wild type allele. The first employs gene replacement, described in Examples 7, 11, 15, 19 and 24, wherein the gene to be altered, along with adjacent upstream and downstream DNA, is mutated and cloned into a *Sac. erythraea* non-replicating vector. The plasmid carrying the altered allele is then introduced into the host strain by transformation of protoplasts employing selection for a plasmid marker. Since the plasmid does not replicate, regenerated cells that carry the marker have undergone a single homologous recombination between one of the two segments flanking the mutation on the plasmid and its homologous counterpart in the chromosome. Some of the colonies that have subsequently lost the marker will have undergone a second recombination between the other

plasmid borne adjacent DNA segment and its homologous chromosomal counterpart resulting in the retention of the mutation in the chromosome, replacing the normal allele with the mutant one. The second method to introduce an altered allele into the chromosome employs gene conversion, described in Examples 37 and 43. In this method, an Ery⁻ *Sac. erythraea* strain carrying a deletion of a specified region of the *eryA* segment of the chromosome is used as a host. Into a *Sac. erythraea* multicopy plasmid that carries a selectable marker is cloned the wild type counterpart (segment 1) of the *eryA* segment mutant in the host. Subsequently, the desired homologous or heterologous DNA segment to be introduced (segment 2) is cloned within the portion of segment 1 which is deleted in the mutant strain. The resulting plasmid is then introduced into the host employing selection for the marker. Among the transformants will be a population that have integrated segments 1 and 2 from the plasmid by the process of gene conversion which can be verified by examination of the DNA among the colonies that have recovered the ability to produce erythromycin.

Two examples each of Types I, II and III alterations to the *eryA* DNA sequence and the resultant novel polyketides produced are described in the examples described herein. Examples 1 through 8, 9 through 12 and 13 through 16 describe the construction and effect of three Type I mutants. Examples 17 through 22 and 23 through 27 describe the construction of two Type II mutants and the effects of feeding two different synthetic substrates to the mutant strains. Examples 28 through 38 and 39 through 44 outline the steps in constructing Type III changes and their respective effects on the structure of the novel polyketides produced. In Examples 1 through 7 a plasmid that contains a substantial deletion of the segment of the gene corresponding to the b-ketoreductase of module 5 is created, the altered gene is inserted into the *Sac. erythraea* chromosome to replace the wild type allele and the new strain carrying the altered gene is identified and isolated. In Example 8, the new strain is fermented and the novel polyketide 5-oxo-5,6-dideoxy-3α-mycarosyl erythronolide B that results from the introduction of the mutant allele is isolated. In Examples 9 through 11, a mutation is introduced into the β–ketoreductase of module 2 and the mutated allele is then used to replace the wild type allele in the chromosome. In Example 12, the strain carrying the altered allele is fermented and the novel compound 11-oxo-11-deoxyerythromycin A is isolated. Similarly, in Examples 13 through 16 a mutation is introduced

into the dehydratase of module 4 and the mutated allele is then used to replace the wild type allele in the chromosome. The strain carrying this altered allele is then fermented and the novel products 7-hydroxyerythromycin A and 6-deoxy-7-hydroxyerythromycin A are isolated. In Examples 17 through 21, a mutation is made in the DNA corresponding to the β-ketoacyl-ACP synthase of module 1 and introduced into the chromosome to replace the wild type allele. This mutation has the effect of arresting the synthesis of the polyketide chain and results in the Ery⁻ phenotype. The synthetic substrate (2S,3R,4S,5S)3,5-dihydroxy-2,4-dimethylhexanoic acid-ethyl ester is then made and fed to the mutant resulting in the production of the novel compound (14S,15S)14(1-hydroxyethyl)erythromycin. Similarly, in Examples 22 through 24, a mutation is created in the β-ketoacyl-ACP synthase of module 2 and introduced into the chromosome to replace the wild type allele. In Example 25 and 26, the synthetic substrate (2S,3S,4S,5S)2,4-dimethyl-3-fluoro-5-hydroxyhexanoic acid-ethyl thioester is made and fed to the module 2 β-ketoacyl-ACP synthase mutant and the resulting novel compound 11-epifluoro-15-norerythromycin is isolated. In Examples 27 through 38, a copy of the DNA sequence corresponding to module 4 is introduced into the deleted segment of the β-ketoacyl-ACP synthase of module 1 resulting in the production of the novel compound 14(1-propyl)erythromycin. In Examples 40 through 44, a copy of the DNA sequence corresponding to module 5 is introduced into the deleted segment of the β-ketoacyl ACP synthase of module 1 resulting in the production of the novel compound 14[1(1-hydroxypropyl)]erythromycin.

GENERAL METHODS

Materials, Plasmids and Bacterial Strains

Restriction endonucleases, T4 DNA ligase, nick-translation kit, competent E. coli DH5α cells , X-gal, IPTG, and plasmids pUC19 and pUC12 are purchased from Bethesda Research Laboratories (BRL), Gaithersburg, MD. [α-^{32}P]dCTP and Hybond N are from Amersham Corp., Chicago, IL. Seakem LE agarose and Seaplaque low gelling temperature agarose are from FMC Bioproducts, Rockland, ME. E. coli K12 strains carrying the E. coli-Sac. shuttle plasmids pWHM3 or pWHM4 (Vara et al., J. Bacteriol., 171: 5872 (1989)) or the cosmids pS1 (Tuan et al., Gene, 90: 21 (1990)) and

Sac. erythraea strain NRRL2338 have been deposited in the culture collection of the Agricultural Research Laboratories, Peoria, IL and are available under the accession numbers NRRL XXXX, respectively. Staphylococcus aureus Th^R (thiostrepton resistant) is obtained by plating 10^8 cells of S. aureus on agar medium containing 10 mg/ml thiostrepton and picking a survivor after 48 hr growth at 37°C. Thiostrepton is obtained from Squibb-Bristol Myers, New Brunswick, NJ. All other chemical and reagents are from standard commercial sources unless specified otherwise.

DNA Manipulations

Standard conditions (Maniatis et al., Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y., 1982) are employed for restriction endonuclease digestion, agarose gel-electrophoresis, nick translation of DNA to make ^{32}P-labeled probes, DNA ligation, and transformation of E. coli employing selection for ampicillin resistance (Ap^R) on LB agar plates. Plasmid DNA is isolated from minipreps of E. coli transformants by the boiling method (Maniatis et al., 1982, supra). DNA fragments are recovered from low melting agarose gels using the method of Langridge et al., 1980. Total DNA from Sac. erythraea strains is prepared according to described procedures (Hopwood et al., Genetic Manipulation of Streptomyces, A Laboratory Manual, John Innes Foundation, Norwich, U.K., 1985). DNA is transferred from agarose gels onto Hybond N following the manufacturer's instructions. Hybridizations are performed in sealed bags containing 10-20 ml of [1xNET (20xNET = 3 M NaCl, 0.3 M TrisHCl, 20 mM Na_2EDTA, pH 8.0), 5XDenhardt's solution (Maniatis et al., 1982, supra), 0.2 mg/ml denatured calf thymus DNA, 0.2% SDS, and 0.5-2x10^7 cpm of the nick-translated probe] for 16-20 hr at 65 °C. Filters are washed three times in 1xNET/0.1% SDS for 20 min each at room temperature, and once in 0.05xNET/0.1% SDS for 20 min at 70 °C. Filters are reused as described (Donadio et al., 1990).

Amplification of DNA fragments

Synthetic deoxyoligonucleotides are synthesized on an ABI Model 380A synthesizer (Applied Biosystems, Foster City, CA) following the manufacturer's recommendations. Amplification of DNA fragments is performed by the polymerase chain reaction (PCR) employing a Coy

thermocycler. Reactions contain 100 pmol e ch primer, 1 μg of template DNA (cosmid pS1 carrying the *eryA* segmen rom Sac. erythraea strain NRRL 2338), and 2.5 units of Thermus aquaticus DNA polymerase in a 100 ml volume of PCR buffer [50 mM KCl, 10 mM TrisHCl (pH 8.0) 2 mM $MgCl_2$, 0.01% gelatin) containing 200 mM of the 4 dNTPs. The above reagents are from Perkin Elmer Cetus, Norwalk, CT. The reaction mixture is overlaid with a drop of paraffin oil and subjected to 30-50 cycles. Each cycle consists of one 94 °C, one 55 °C and one 72 °C period, each of the duration of 3 min. The progress of the amplification is monitored by agarose gel-electrophoresis. The PCR primers described in the examples below are derived from the nucleotide sequence of *eryA* of FIG. 2.

Gene replacement and gene conversion

Protoplasts of *Sac. erythraea* strains are prepared and transformed with miniprep DNA isolated from E. coli according to published procedures (Yamamoto et al., 1986). Integrative transformants, in the case of pWHM3 derivatives, are selected after one round of non-selective growth of the primary Th^R transformants as described by Weber et. al, Gene, 68: 173 (1988). Loss of the Th^R phenotype is monitored by plating serial dilutions of a Th^R integrant on non-selective medium, followed by replica-plating on thiostrepton-containing medium. Th^S (thiostrepton-sensitive) colonies arise at a frequency of 10^{-2} (Donadio et al., 1990). The retention of the mutant allele is established by Southern hybridization of a few Th^S colonies.

A few hundred Th^R colonies obtained by transformation of an *eryA* strain with pWHM4 derivatives are screened for antibiotic production by the agar-plug assay employing *Staphylococcus aureus* as Th^R organism as described (Tuan et al.,Gene, 90: 21 (1990)). The frequency of gene conversion between a 5 kb segment of homologous sequence and a strain carrying a small deletion is >25% (Tuan et al.,Gene, 90: 21 (1990)). Colonies found to produce antibiotic activity are inoculated in SGGP (Yamamoto et al., 1986), protoplasts are prepared, and the regenerated protoplasts are scored for loss of the plasmid by replica-plating on non-selective medium. Th^S colonies are then rechecked for antibiotic production, and six producers are analyzed on Southern blots.

Fermentation

Sac. erythraea cells are inoculated into 100 ml SCM medium (1.5% soluble starch, 2.0% Soytone [Difco], 0.15% Yeast Extract [Difco], 0.01% $CaCl_2$) and allowed to grow at 32°C for 3 to 6 days. The entire culture is then inoculated into 10 liters of fresh SCM medium. The fermenter is operated for a period of 7 days at 32°C maintaining constant aeration and pH at 7.0. After fermentation is complete, the cells are removed by centrifugation at 4°C and the fermentation beer is kept in the cold until further use.

The present invention will now be illustrated, but is not intended to be limited, by the following examples:

Example 1
Construction of plasmid pABX9

The 9.6 kb BamHI-XhoI segment comprised between sequence coordinates 21.96 and 31.52 was isolated from cosmid pS1 and ligated to SalI-digested pUC19 DNA. The resulting mixture contained the desired plasmid pABX9.

Example 2
Construction of E. coli K12 DH5α/pABX9

Approximately 10 ng of plasmid pABX9, prepared as described in Example 1, were transformed into E. coli K12 DH5α and a few of the resulting white Ap^R colonies that appeared on the LB-agar plates containing X-gal and ampicillin were analyzed for their plasmid content. One colony was found to carry pABX9, as verified by the observation of fragments of 3.93, 3.39, 2.01, 1.56, 0.87, and 0.48 kb in size upon agarose gel electrophoresis after SmaI digestion of the plasmid.

Example 3
Construction of plasmid pABX9DN

Plasmid pABX9, isolated from E. coli K12 DH5α/pABX9, was digested with NcoI and then treated with T4 DNA ligase. The resulting mixture contained the desired plasmid pABX9DN.

Example 4

Construction of E. coli K12 DH5a/pABX9DN

Approximately 10 ng of plasmid pABX9DN, prepared as described in Example 3, were transformed into E. coli K12 DH5α and a few of the resulting white Ap^R colonies that appeared on the LB-agar plates containing X-gal and ampicillin were analyzed for their plasmid content. Colonies carrying pABX9DN exhibited a single NcoI fragment of 11.5 kb visible by agarose gel electrophoresis, confirming that the 813 bp NcoI - NcoI fragment from pABX9 has been deleted in pABX9DN.

Example 5

Construction of plasmid pABX95DN

Plasmid pABX95DN was digested with EcoRI and HindIII and ligated to pWHM3 digested with the same two enzymes. The resulting mixture contained the desired plasmid pABX95DN.

Example 6

Construction of E. coli K12 DH5α/pABX95DN

Approximately 10 ng of plasmid pABX95DN, prepared as described in Example 5, were transformed into E. coli K12 DH5a and a few of the resulting white Ap^R colonies that appeared on the LB-agar plates containing X-gal and ampicillin were analyzed for their plasmid content. Colonies carrying pABX95DN exhibited fragments of 8.8 and 7.2 kb visible in agarose gels after EcoRI and HindIII digestion.

Example 7

Construction of Sac. erythraea AKR5 carrying the eryAKR5 allele by gene replacement

Approximately 1 mg of plasmid pABX95DN, isolated from E. coli K12 DH5α/pABX95DN, was transformed into *Sac. erythraea* NRRL 2338 and stable Th^R colonies were isolated. Serial dilutions of one of these colonies were screened for the loss of the antibiotic resistance marker and total DNA from 5 Th^S colonies as well as from untransformed *Sac. erythraea* NRRL 2338 was digested with SstI and analyzed by Southern

hybridization employing the 0.8 kb SalI fragment between sequence coordinates 24.26 and 25.06 (from pABX9) as probe. Whereas NRRL 2338 showed one SstI band of 3.7 kb that hybridized to the probe, samples from four of the Th^S strains exhibited a SstI-hybridizing band of 6.1 kb indicating the presence of the mutant allele. One of these colonies was kept and designated strain AKR5. It carries a deletion of 813 bp in the KR5 segment of *eryA* and is designated the eryAKR5 allele.

Example 8

Isolation, purification and properties of 5-oxo-5,6-dideoxy-3-a-mycarosyl erythronolide B from Sac. erythraea AKR5

A 10-liter fermentation of *Sac. erythrea* AKR5 carrying the eryAKR5 allele in a Biolafitte fermentor using SNC Media. The fermentor was inoculated with 100 ml of a 3 day old seed. The pO_2 was initially 80 ppm and the temperature was maintained at 32°C. The pH was controlled to 7.0 ± 0.2 by addition of propionic acid or potassium hydroxide as needed. At harvest (3 days), the whole broth was extracted three times with 4-liter portions of ethylacetate. The combined extracts were concentrated under reduced pressure and the residue was chromatographed on a column (50 x 5 cm) of silica gel packed and loaded in toluene and eluted with a stepwise gradient of increasing concentration of isopropanol in toluene. Fractions were analyzed by TLC and spots were detected by spraying with anisaldehyde sulfuric acid spray reagent and heating. A major component giving blue colored spots eluted with approximately 7% isopropanol. Fractions containing this material were combined and concentrated to a residue (800 mg). This was further chromatographed on a column (100 x 3 cm) of Sephadex LH-20 in chloroform-heptane-ethanol, 10:10:1, v/v/v. Fractions were analyzed as above, early fractions (9-13) yielded 5,6-dideoxy-3-a-mycarosyl-5-oxoerythronolide B (45 mg) which was crystallized from heptane/ethylacetate mixture to mp 163-164 °C.

CMR spectrum in $CDCl_3$ (ppm downfield from TMS)

8.6	37.9	70.0
9.9	38.7	76.2
9.9	40.4	76.4
10.4	40.7	80.4
14.5	43.3	100.4
15.2	45.8	175.8
17.1	46.8	210.8
17.7	48.9	217.7
25.3	66.5	
25.5	69.4	

Structure was determined by single crystal X-ray diffraction.

Later fractions (15-17) yielded 5,6-dideoxy-5-oxoerythronolide B (10 mg) and still later fractions yielded 5,6-dideoxy-6,6a-epoxy-5-oxoerythronolide B (2.8 mg).

Example 9
Construction of plasmid pALeryAKR2

The 1.3 kb DNA segment comprised between coordinates 8.63-9.93 (fragment 1) is amplified by PCR employing two oligodeoxynucleotides, 1a (5'-GGGAGCATGCTCTCGGTGCGCGGCGGCCGC-3') and 1b (5'-GCCCTGCAGCGCGTACTCCGAGGTGGCGGT-3'). Similarly, the 1.3 kb DNA segment between coordinates 9.99-11.26 (fragment 2) is PCR-amplified employing primers 2a (5'-TGGTCTGCAGGCGAGGCCGGACACCGAGG-3') and 2b (5'-GGAAGAAGTCAAAGTTCCTCGGTCCCTTCT-3'). After digestion with SphI + PstI (fragment 1) and PstI + EcoRI (fragment 2), the two fragments are ligated to EcoRI + SphI-digested pWHM3. The resultant mixture contains the desired plasmid pALeryAKR2.

Example 10
Construction of E. coli K12 DH5a/pALeryAKR2

Approximately 10 ng of plasmid pALeryAKR2, prepared as described in Example 9, are transformed into E. coli K12 DH5α, and a few of the resulting white Ap^R colonies that appear on the LB-agar plates containing X-gal and ampicillin are analyzed for their plasmid content. The identity of plasmid pALeryA2KR2, 9.8 kb in size, and carrying a 2.6 kb EcoRI-SphI insert with an internal PstI site, is verified by SalI digestion (fragments at 2.91, 2.21, 1.61, 1.42, 1.08, 0.29, 0.12 and 0.10 kb are released, visible by agarose gel electrophoresis). pALeryAKR2 contains an in-frame deletion of 102 base pairs of the corresponding segment of the wild type *eryA* chromosomal DNA. The cloned segment in pALeryAKR2 is designated the eryAKR2 allele.

Example 11
Construction of Sac. erythraea AKR2 carrying the eryAKR2 allele by gene replacement

Approximately 1 mg of plasmid pALeryAKR2, isolated from E. coli K12 DH5α/pALeryAKR2, is transformed into *Sac. erythraea* protoplasts and stable Th^R colonies are isolated. Serial dilutions of one of these colonies are screened for loss of the antibiotic resistance marker, and six Th^S colonies are analyzed for their genotype by Southern hybridization. Total DNA from the six Th^S colonies and from untransformed Sac. erythraea NRRL2338 is digested with PstI and with SalI and is then examined by Southern hybridization using the 2.6 kb EcoRI-SphI insert from pALeryAKR2 as probe. Whereas NRRL2338 contains a 39 kb PstI hybridizing band, colonies in which the mutation in KR2 has been introduced (strain AKR2) exhibit two bands of approximately equal intensity, one at 27 kb and the other at 12 kb. The SalI digest, with bands at 1.04, 0.75, 0.29, 0.12 and 0.10 kb common to NRRL2338 and AKR2, but with the 1.16 kb band in NRRL2338 replaced by the 1.06 kb band in AKR2, confirms that the only change introduced into strain AKR2 is the deletion of the 102 bp segment from KR2, resulting in a strain carrying the eryAKR2 allele.

Example 12

Isolation and purification of 11-deoxy-11-oxoerythromycin A

The fermentation beer of strain AKR2, cooled to 4°C is adjusted to pH 8.0 and is extracted sequentially with three equal volumes of methylene chloride. The combined methylene extracts are concentrated to an oily residue and partitioned between heptane and methanol. The methanol layer is removed, washed once with heptane and concentrated to a residue. The residue is digested in methylene chloride and washed once with potassium phosphate buffer pH 7.8 and once with water. The methylene chloride layer is concentrated to a residue and digested in the lower phase (1:1:1, v/v/v) of a carbon tetrachloride; methanol; aqueous phosphate buffer (0.05 M, pH 7.0) system and chromatographed on an Ito Coil Planet Centrifuge in the same system. Fractions containing the desired 11-oxo-11-deoxyerythromycin A were combined, concentrated, digested in methylene chloride, washed well with water and concentrated on rotary evaporator under reduced pressure to yield 11-deoxy-11-oxoerythromycin A as an off-white solid froth. Its identity is confirmed by comparison with antibiotic L53-18A. 11-Deoxy-11-oxoerythromycin A is dissolved in tetrahydrofuran and the solution is diluted with an equal volume of water. This is then acidified to pH 4.0 and allowed to stand at room temperature for 4 hours. The pH is adjusted to 9.0 and the solution is diluted with an equal volume of water and extracted with two volumes of methylene chloride. The combined methylene chloride extracts were evaporated to dryness under reduced pressure to yield antibiotic L53-18A as a white solid.

Example 13

Construction of plasmid pALeryADH4

Primers 3a (GCGCGAGCTCGACGACCAGGGCGGCATGGT) and 3b (GGTGGCATGCTGCGACCACTGCGCGTCGGC) are used to PCR-amplify the 1.05 kb *eryA* segment of the Sac. erythraea chromosome between sequence coordinates 18.47-20.07 (fragment 3), and primers 4a (AGCTGCATGCTCTGGACTGGGGACGGCTAG) and 4b (CGCGGGATCCCAGCTCCCACGCCGATACCG) are used to amplify the 1.35 kb segment between sequence coordinates 20.58-21.96 (fragment 4) as described in Example 1. Fragment 3 and 4, after digestion with SstI + SphI

and with SphI + BamHI, respectively, are ligated to SstI -, BamHI-digested pWHM3. The resulting ligation mixture contains the desired plasmid pALeryADH4.

Example 14
Construction of E. coli K12 DH5a/pALeryADH4

Approximately 10 ng of pALeryADH4, prepared as described in Example 13, are transformed transformed into E. coli K12 DH5α, and a few of the resulting white Ap^R colonies that appear on the LB-agar plates containing X-gal and ampicillin are analyzed for their plasmid content. The identity of plasmid pALeryADH4, 9.6 kb in size, is verified by SphI + EcoRI digestion (fragments at 7.2, 1.35 and 1.05 kb are released). pALeryADH4 carries a 498 base pair in-frame deletion of the corresponding segment of the wild type *eryA* DNA. The cloned segment in pALeryADH4 is designated the eryADH4 allele.

Example 15
Construction of Sac. erythraea ADH4 carrying the eryADH4 allele by gene replacement

Approximately 1 mg of plasmid pALeryADH4, isolated from E. coli K12 DH5α/pALeryADH4, is used for transformation into *Sac. erythraea* protoplasts and stable Th^R colonies are isolated. Serial dilutions of one of these colonies are screened for loss of the antibiotic resistance marker, and six Th^S colonies are analyzed for their genotype by Southern hybridization. Total DNA from the six Th^S colonies and from untransformed *Sac. erythraea* NRRL2338 is digested with SphI and with SstI and examined by Southern hybridization using the 2.4 kb SstI-BamHI insert from pALeryADH4 as probe. Strains in which the wild type allele has been replaced by the mutated copy show two SphI bands, one at 13.5 kb and the other at 12.4 kb, whereas the wild type strain exhibits a single band at 26 kb. The SstI pattern, with the 2.9 kb band from NRRL2338 being replaced in ADH4 by a 2.5 kb band, confirms that the 487 bp deletion created in plasmid pALeryADH4 has been transferred into the chromosome of ADH4. Strains that carry the eryADH4 allele in place of the wild type sequence are designated Sac. erythraea ADH4.

Example 16

Isolation and characterization of 7-hydroxyerythromycin A and 6-deoxy-7-hydroxyerythromycin A

The fermentation beer of strain ADH4 is cooled to 4°C and the pH is adjusted to 5.0. The mixture is extracted once with an equal volume of methylene chloride. The pH of the aqueous layer is readjusted to 9.0 and two further methylene chloride extracts are carried out. These two extracts are combined, washed with water and concentrated to a residue. This is digested in 10 ml of the upper phase of a (3:7:5, v/v/v) mixture of hexane, ethylacetate, aqueous phosphate buffer (0.05 M, pH 7.5) and chromatographed on an Ito Coil Planet Centrifuge in the same system. Fractions containing the desired 7-hydroxyerythromycin were combined, concentrated, and partitioned between methylene chloride and dilute (pH 9.5) ammonium hydroxide solution. Fractions containing the desired 6-deoxy-7-hydroxyerythromycin were combined, concentrated, and partitioned between methylene chloride and dilute (pH 9.5) ammonium hydroxide solution. The methylene chloride layers are washed with water and then concentrated to yield the desired 7-hydroxyerythromycin A and 6-deoxy-7-hydroxyerythromycin A as white foams.

Example 17

Construction of plasmid pALeryAKS1

The 1.4 kb segment of *eryA* , between sequence coordinates 1.11-2.54 (fragment 5) and the 1.5 kb segment between sequence coordinates 2.88-4.37 (fragment 6) are PCR-amplified using primers 5a (TGCAGAATTCGCTGGCCGCGCTCTGGCGCT) and 5b (GAGAGCTGCAGCATGAGCCGCTGCTGCGGG), and 6a (CATGCTGCAGGACTTCAGCCGGATGAACTC) and 6b (GAGGAAGCTTCCAGCCGGTCCAGTTCGTCC), respectively, as described in Example 9. After digestion with EcoRI + PstI (fragment 5) and PstI + HindIII (fragment 6), the two fragments are ligated to EcoRI + HindIII-digested pWHM3. The resulting mixture contains the desired plasmid pALeryAKS1.

Example 18
Construction of E. coli K12 DH5a/pALeryAKS1

Approximately 10 ng of pALeryAKS1, prepared as described in Example 17, are transformed into E. coli K12 DH5α, and a few of the resulting white Ap^R colonies that appear on the LB-agar plates containing X-gal and ampicillin are analyzed for their plasmid content. The identity of plasmid pALeryAKS1, 10.1 kb in size, is verified by digestion with PstI + HindIII (fragments of 8.6 and 1.5 kb are observed by agarose gel electrophoresis) and with SalI (fragments of 2.93, 2.21, 1.42, 1.37, 0.86, 0.54, 0.27, 0.14, 0.13, and 0.10 kb are observed). pALeryAKS1 carries an in-frame deletion of 282 base pairs of the corresponding wild type *eryA* DNA. The cloned insert in plasmid pALeryAKS1 is designated the eryAKS1 allele.

Example 19
Construction of Sac. erythraea AKS1 carrying the eryAKS1 allele by gene replacement

Approximately 1 mg of plasmid pALeryAKS1, isolated from E. coli K12 DH5α/pALeryAKS1, is used for transformation into *Sac. erythraea* protoplasts and stable Th^R colonies are isolated. Serial dilutions of one of these colonies are screened for loss of the antibiotic resistance marker, and six Th^S colonies are analyzed for their genotype by Southern hybridization. Total DNA from the six Th^S colonies and from untransformed *Sac. erythraea* NRRL2338 is digested with PstI and with SmaI and examined in Southern hybridization employing the 2.9 kb EcoRI-HindII insert from pALeryAKS1 as probe. Colonies in which the wild type allele has been replaced by the mutated copy (strain AKS1) show two PstI bands, one at 34.5 and the other at 4.4 kb, whereas the wild type strain exhibits a single band at 39 kb. The SmaI pattern, with the 2.9 kb band from NRRL2338 being replaced in AKS1 by a 2.6 kb band, confirms that the 282 bp created in plasmid pALeryAKS1 has been transferred into strain AKS1. Strains that carry the eryAKS1 allele are designated Sac. erythraea AKS1.

Example 20

Synthesis of (2S,3R,4S,5S)3,5-dihydroxy-2,4-dimethylhexanoic acid n-butyl thioester

A convenient source of this compound in chiral purity is the antibiotic oleandomycin. Oleandomycin (5 g) is dissolved in an aprotic solvent such as toluene and treated with diazabicyclo[5.4.0]undecene-5 (1 g) and heated for one hour. The resulting solution is poured into iced water, agitated well and the organic layer is drawn off and concentrated to a residue. The residue is digested in methylene chloride and treated exhaustively with a solution of ozone. The resulting ozonide is oxidatively decomposed with dilute hydrogen peroxide in sufficient aqueous ethanol to yield a monophasic mixture. This is further diluted with water and made 0.1 N with sodium hydroxide. The mixture is warmed for one hour at 70°C and then cooled before being acidified to pH 2.5 with dilute sulfuric acid. The mixture is then exhaustively extracted with methylene chloride. The combined extracts are concentrated to an oily residue and the desired lactone is recovered by chromatography on silica gel eluted with a gradient of toluene-isopropanol.

The δ-lactone is converted to the butyl thioester before feeding to *Sac. erythrea* AKS1 by refluxing with n-butylthiol in the presence of a catalytic amount of triethylamine.

Example 21

Isolation of (14S,15S)14(1-hydroxyethyl)erythromycin A

The fermentation broth of AKS1 is cooled to 4°C and adjusted to pH 4.0 and extracted once with methylene chloride. The aqueous layer is readjusted to pH 9.0 and extracted twice with methylene chloride and the combined extracts are concentrated to a solid residue. This is digested in methanol and chromatographed over a column of Sephadex LH-20 in methanol. Fractions are tested for bioactivity against a sensitive organism, such as Staphylococcus aureus Th^R, and active fractions are combined. The combined fractions are concentrated and the residue is digested in 10 ml of the upper phase of a solvent system consisting of n-heptane, benzene, acetone, isopropanol, 0.05 M, pH 7.0 aqueous phosphate buffer (5:10:3:2:5, v/v/v/v/v), and chromatographed on an Ito Coil Planet

Centrifuge in the same system. Active fractions are combined, concentrated and partitioned between methylene chloride and dilute ammonium hydroxide (pH 9.0). The methylene chloride layer is separated and concentrated to yield the desired product as a white foam.

Example 22
Construction of plasmid pALeryAKS2

Primers 7a (CGCCCGAATTCGAGGCGCTGGGCGCCCGGAC) and 7b (CCACCTGCAGCGCGGGACCTTCCAGCCCC), and primers 8a (GTGGGTCGCTGCAGACGGTGACTGCGG) and 8b (GGTCAAGCTTCGTCGGCGAGCAGCTTCTC) are used to PCR-amplify the 1.45 kb *eryA* segment between sequence coordinates 5.71-7.16 (fragment 7) and the 1.5 kb *eryA* segment between sequence coordinates 7.22-8.70 (fragment 8), respectively. After digestion with EcoRI + PstI (fragment 7) and with PstI + HindIII (fragment 8), the two fragments are ligated to pWHM3 cut with EcoRI + HindIII. The resulting mixture contains the desired plasmid pALeryAKS2.

Example 23
Construction of E. coli K12 DH5a/pALeryAKS2

Approximately 10 ng of pALeryAKS2, prepared as described in Example 22, are transformed into E. coli K12 DH5α_r and a few of the resulting white Ap^R colonies that appear on the LB-agar plates containing X-gal and ampicillin are analyzed for their plasmid content. The identity of plasmid pALeryAKS2, 10.1 kb in size, is verified by digestion with PstI + HindIII (fragments of 8.6 and 1.5 kb are observed by agarose gel electrophoresis) and with SstII (fragments of 4.0, 2.3, 2.0, 0.72, 0.43, 0.40, 0.20, 0.18, 0.13 and 0.11 kb observed). Plasmid pALeryAKS2 carries an in-frame deletion of 60 base pairs of the corresponding wild type *eryA* DNA. This deletion removes the active site cysteine from KS2. The cloned insert in plasmid pALeryAKS2 is designated the eryAKS2 allele.

Example 24

Construction of Sac. erythraea AKS2 carrying the eryAKS2 allele by gene replacement

Approximately 1 mg of plasmid pALeryAKS2, isolated from E. coli K12 DH5α/pALeryAKS2, is used for transformation into Sac. erythraea protoplasts and stable Th^R colonies are isolated. Serial dilutions of one of these colonies are screened for loss of the antibiotic resistance marker, and six Th^S colonies are analyzed for their genotype by Southern hybridization. Total DNA from the six Th^S colonies and from untransformed Sac. erythraea NRRL2338 is digested with PstI and with SstII and examined in Southern hybridization employing the 2.9 kb EcoRI-HindII insert from pALeryAKS2 as probe. Colonies in which the wild type allele has been replaced by the mutated copy (strain AKS2) show two PstI bands, one at 34.5 and the other at 4.4 kb, whereas the wild type strain exhibits a single band at 39 kb. The SstII pattern, with the 0.78 kb band from NRRL2338 being replaced in AKS2 by a 0.72 kb band, confirms that the 60 bp created in plasmid pALeryAKS2 has been transferred into strain AKS2. Strains that carry the eryAKS2 allele are designated Sac. erythraea AKS2.

Example 25

Synthesis of (2R,3R,4S,5R)2,4-dimethyl-3-fluoro-5-hydroxyhexanoic acid n-butyl thioester

(2R,3S,4S,5R)3,5-Dihydroxy-2,4-dimethylhexanoic acid-δ-lactone (1 g) from Example 20 is digested in 10 ml of pyridine and treated with p-toluenesulfonyl chloride (1.3 g) and allowed to stand at room temperature overnight. The mixture is poured into iced water and extracted with methylene chloride and the methylene chloride is concentrated to the crude sulfonate ester. This is digested in acetonitrile (100 ml) and heated under reflux after the addition of tetrabutylammonium fluoride (1.75 g). After 6 hours the mixture is cooled, poured over iced water (300 ml) and extracted three times with 200 ml portions of methylene chloride. The combined methylene chloride extracts were concentrated and the residue was chromatographed on a column of silica gel eluted with a stepwise gradient of isopropanol (0 to 50%) in toluene. Fractions containing (2R,3R,4S,5R)2,4-dimethyl-3-fluoro-5-hydroxyhexanoic acid d-lactone were

combined and concentrated to a white solid. The lactone is then converted to the n-butyl thiolester by refluxing in n-butyl thiol with a catalytic amount of triethylamine. Solvent is removed and the residue is digested in DMSO before feeding to fermentations of Sac. erythraea AKS2.

Example 26

Isolation and purification of 11-epifluoro-15-norerythromycin A

The fermentation broth of strain AKS2 is cooled to 4°C and adjusted to pH 4.0 and extracted once with ethylacetate. The aqueous layer is adjusted to pH 9.0 and extracted twice with methylene chloride and the combined extracts are concentrated to a white solid. This is chromatographed over a column of Sephadex LH-20 in a mixture of heptane, chloroform, ethanol (10:10:1, v/v/v) and fractions containing the desired product are combined and concentrated to a solid residue. This is further purified by countercurrent chromatography on an Ito Coil Planet Centrifuge on a system composed of carbon tetrachloride; methanol; 0.05 M; pH 7.0 aqueous potassium phosphate buffer (1:1:1, v/v/v). Fractions containing the desired 11-epifluoro-15-norerythromycin were combined, and concentrated to a residue. This was digested in methylene chloride and dilute (pH 9.5) ammonium hydroxide and the methylene chloride layer was separated, washed with water and concentrated to yield the desired 11-epifluoro-15-norerythromycin A as white solid.

Example 27

Construction of plasmid pALeryAM4.1

Primers 9a (GCGCCGAATTCTCGAGACGGCGTGGGAGGCA) and 9b (TTGCGGTACCAGTAGGAGGCGTCCATCGCG) are employed to PCR-amplify the 2.0 kb *eryA* segment between sequence coordinates 17.35-19.38 (fragment 9). After digestion with EcoRI + KpnI, fragment 9 is ligated to pUC19 cut with the same two enzymes The resulting mixture contains the desired plasmid pALeryAM4.1.

Example 28

Construction of E. coli K12 DH5a/pALeryAM4.1

Approximately 10 ng of pALeryAM4.1, prepared as described in Example 27, are transformed into E. coli K12 DH5a, and a few of the resulting white Ap^R colonies that appear on the LB-agar plates containing X-gal and ampicillin are analyzed for their plasmid content. The identity of plasmid pALeryAM4.1, 4.7 kb in size, is verified by digestion with SalI (fragments of 2.8, 0.85, 0.53, 0.27 and 0.22 kb are observed by agarose gel electrophoresis).

Example 29

Construction of plasmid pALeryAM4.2

Primers 10a (GCTGGGATCCCGCGGCGCGGGTTGCAGCAC) and 10b (CGGAACTCGGTGAGCATGCCGGGACTGCTC) are used to PCR-amplify the 2.1 kb *eryA* segment between sequence coordinates 21.94-24.00 (fragment 10). The 2.6 kb fragment KpnI(96)-BamHI(102) from cosmid clone pR1, and fragment 10 cut with BamHI + SphI, are ligated to pALeryAM4.1 cut with KpnI + SphI. The resulting mixture contains the desired plasmid pALeryAM4.2.

Example 30

Construction of E. coli K12 DH5a/pALeryAM4.2

Approximately 10 ng of pALeryAM4.2, prepared as described in Example 29, are transformed into E. coli K12 DH5a, and a few of the resulting white Ap^R colonies that appear on the LB-agar plates containing X-gal and ampicillin are analyzed for their plasmid content. The identity of plasmid pALeryAM4.2, 9.3 kb in size, is verified by digestion with XhoI + SphI (to ensure that the entire 6.65 kb insert is released) and with SalI, with fragments of 2.8, 1.82, 1.09, 0.94, 0.85, 0.75, 0.45, 0.27, 0.22 and 0.13 kb are observed by agarose gel electrophoresis).

Example 31

Construction of plasmid pALeryAM1

The 2.9 kb SmaI(4)-SmaI(20) fragment from cosmid clone pR1 is ligated to pUC12 cut with SmaI. The resulting mixture contains plasmid pALeryAM1.

Example 32

Construction of E. coli K12 DH5αa/pALeryAM1

Approximately 10 ng of pALeryAM1, prepared as described in Example 31, are transformed into E. coli K12 DH5α, and a few of the resulting white Ap^R colonies that appear on the LB-agar plates containing X-gal and ampicillin are analyzed for their plasmid content. The identity of plasmid pALeryAM1, 5.6 kb in size, is verified by digestion with SmaI (the 2.9 kb insert is realeased) and with SphI, with release of one 4.4 and one 1.07 kb bands. Both orientations of the insert in plasmid pALeryAM1 are useful.

Example 33

Construction of plasmid pALeryAM4.3

Plasmid pALeryAM1 is cut with XhoI to completion, partially with SphI, and the resulting 5.25 kb band, isolated from an agarose gel, is ligated to the 6.65 kb insert released from pALeryAM4.2 by XhoI + SphI digestion The resulting mixture contains the desired plasmid pALeryAM4.3.

Example 34

Construction of E. coli K12 DH5a/pALeryAM4.3

Approximately 10 ng of pALeryAM4.3, prepared as described in Example 33, are transformed into E. coli K12 DH5α, and a few of the resulting white Ap^R colonies that appear on the LB-agar plates containing X-gal and ampicillin are analyzed for their plasmid content. The identity of plasmid pALeryAM4.1, 11.9 kb in size, is verified by XhoI + SphI digestion (fragments of 6.65 and 5.25 kb are visible by agarose gel-electrophoresis). Plasmid pALeryAM4.3 carries the entire *eryA* module 4

inserted into the KS region of module 1. The cloned insert in pALeryAM4.3 is degnated the eryAM412 allele.

Example 35
Construction of plasmid pALeryAM4.4

Plasmid pALeryAM4.3 is cut with EcoRI + HindIII, and the resulting 9.2 kb band, recovered from an agarose gel, is ligated to pWHM4 cut with the same two enzymes. The resulting mixture contains the desired plasmid pALeryAM4.4.

Example 36
Construction of E. coli K12 DH5α/pALeryAM4.4

Approximately 10 ng of pALeryAM4.4, prepared as described in Example 35, are transformed into E. coli K12 DH5α, and a few of the resulting white Ap^R colonies that appear on the LB-agar plates containing X-gal and ampicillin are analyzed for their plasmid content. The identity of plasmid pALeryAM4.1, 16.5 kb in size, is verified by EcoRI + HindIII digestion, with fragments of 9.2 and 7.3 kb released. Plasmid pALeryAM4.4 carries the eryAM412 allele on the Sac. erythraea multicopy vector pWHM4.

Example 37
Construction of Sac. erythraea AM412 carrying the eryAM412 allele by gene conversion

Approximately 1 mg of plasmid pALeryAM4.4, isolated from E. coli K12 DH5α/pALeryAM4.4, is used for transformation into Sac. erythraea strain AKS1 protoplasts. A few hundred transformants are screened for antibiotic production by the agar-plug assay, and one of the colonies found to produce antimicrobial activity is cured of pALeryAM4.4 by protoplast formation and regeneration as described in General Methods. Total DNA from six antibiotic-producing, Th^S colonies (strain AM412)and from strain AKS1 is digested with SphI and with XhoI and the resulting Southern blot is hybridized first to the 2.9 kb insert from pALeryAM1, and then to the 2.9 kb SstI(95)-SstI(101) fragment from plasmid pALeryAM4.2. With the first probe, the SphI band at 0.8 kb in strain AKS1 is seen to be replaced by a 7.5

kb band in strain AM412, whereas the other two bands at 2.4 kb and 5.2 kb are unaffected. In the XhoI digest, the AKS1 band at 2.9 kb is replaced by a 9.6 kb band in AM412, with the other band at 5.2 kb conserved in both strains. Using the SstI(95)-SstI(101) fragment as probe, strain AKS1 exhibits one band at 25.5 kb and one at 17.9 kb in the SphI and XhoI digests, respectively, whereas, in addition to these bands, strain AM412 shows one SphI band at 7.5 kb and one XhoI band at 9.6 kb. In this way, it is established that the eryAKS1 allele has been converted into the eryAM412 allele in strain AM412.

Example 38
Isolation and purification of 14-(1-propyl)erythromycin A

At harvest the fermentation is adjusted to pH 9.5 and extracted twice with equal volumes of methylene chloride. The combined extracts are washed once with water and concentrated to an oily residue. This is partitioned in a heptane methanol water (5:5:1, v/v/v) system and the lower layer is washed once with heptane and then concentrated to a semisolid residue. This is digested in methanol and chromatographed over a column of Sephadex LH-20 in methanol. Fractions are tested for bioactivity in an agar diffusion assay on plates seeded with the macrolide-sensitive strain Staphylococcus aureus Th^R. Active fractions are combined and further purified by chromatography over silica gel a chloroform:methanol gradient containing 0.1% triethylamine. Fractions containing the desired 14-(1-propyl)erythromycin A are combined and concentrated to yield the product as a white solid.

Example 39
Construction of plasmid pALeryAM5.1

The 4.7 kb *eryA* fragment between sequence coordinates 23.65-28.36 (fragment 11) is PCR-amplified employing primers 11a (ATGCTCGAGATCTCGTGGGAGGCGCTGGA) and 11b (AGAACTCGGTGAGCATGCCCGGGCCCGCCA). Fragment 11, after digestion with XhoI + SphI, is ligated to the 5.25 kb fragment resulting from complete XhoI and partial SphI digestion of pALeryAM1, as in Example 33. The resulting mixture contains the desired plasmid pALeryAM5.1.

Example 40

Construction of E. coli K12 DH5α/pALeryAM5.1

Approximately 10 ng of pALeryAM5.1, prepared as described in Example 39, are transformed into E. coli K12 DH5α, and a few of the resulting white Ap^R colonies that appear on the LB-agar plates containing X-gal and ampicillin are analyzed for their plasmid content. The identity of plasmid pALeryAM5.1, 9.95 kb in size, is verified by SphI + XhoI digestion, with fragments of 5.25 and 4.7 kb released, and by SmaI digestion where fragments of 3.39, 2.68 and 1.94 (doublet) kb are observed. Plasmid pALeryAM5.1 carries the entire *eryA* module 5 inserted into the β–ketoacyl ACP synthase region of module1. The cloned insert in plasmid pALeryAM5.1 is designated the eryA512 allele.

Example 41

Construction of plasmid pALeryAM5.2

Plasmid pALeryAM5.1 is cut with EcoRI + HindIII and the resulting 6.3 kb fragment, recovered from an agarose gel, is ligated to pWHM4 cut with the same two enzymes. The resulting mixture contains the desired plasmid pALeryAM5.2.

Example 42

Construction of E. coli K12 DH5α/pALeryAM5.2

Approximately 10 ng of pALeryAM5.2, prepared as described in Example 41, are transformed into E. coli K12 DH5α, and a few of the resulting white Ap^R colonies that appear on the LB-agar plates containing X-gal and ampicillin are analyzed for their plasmid content. The identity of plasmid pALeryAM5.2, 13.6 kb in size, is verified by digestion with EcoRI + HindIII, with fragments of 7.3 and 6.3 kb released. Plasmid pALeryAM5.2 contains the eryAM512 allele on the Sac. erythraea multicopy vector pWHM4.

Example 43

Construction of Sac. erythraea AM512 carrying the eryAM512 allele by gene conversion

Approximately 1 mg of plasmid pALeryAM5.2, isolated from E. coli K12 DH5α/pALeryAM5.2, is used for transformation into Sac. erythraea strain AKS1 protoplasts. A few hundred transformants are screened for antibiotic production by the agar-plug assay, and one of the colonies found to produce antimicrobial activity is cured of pALeryAM5.2 by protoplast formation and regeneration as described in General Methods. Total DNA from six antibiotic-producing, Th^S colonies (strain AM512)and from strain AKS1 is digested with SphI and with XhoI and the resulting Southern blot is hybridized first to the 2.9 kb insert from pALeryAM1, and then to the 0.8 kb NcoI(119)-NcoI(123) fragment from plasmid pALeryAM5.1. With the first probe, the SphI band at 0.8 kb in strain AKS1 is replaced by a 5.5 kb band in strain AM512, whereas the other two bands at 2.4 kb and 5.2 kb are unaffected. In the XhoI digest, the AKS1 band at 2.9 kb is replaced by a 7.6 kb band in AM512, with the other band at 5.2 kb conserved in both strains. Using the NcoI(119)-NcoI(123) fragment as probe, strain AKS1 exhibits one band at 25.5 kb and one at 17.9 kb in the SphI and XhoI digests, respectively, whereas, in addition to these bands, strain AM512 shows one SphI band at 5.5 kb and one XhoI band at 7.6 kb. In this way, it is established that the eryAKS1 allele has been converted into the eryAM512 allele in strain AM512.

Example 44

Isolation and purification of 14[1(1-hydroxypropyl)]erythromycin A

At harvest the pH of the fermentation of AM512 is adjusted to 9.5 and the mixture is extracted twice with equal volumes of ethylacetate. The combined ethylacetate extracts are washed with water, dried and partitioned in a heptane, methanol, water (5:5:1, v/v/v) system. The lower (methanolic phase) is washed with an equal volume of heptane and is concentrated to a residue. This is chromatographed on a Sephadex LH-20 column in methanol and fractions containing the desired 14[1(1-hydroxypropyl)]erythromycin A are concentrated and further purified by chromatography on an Ito Coil Planet Centrifuge in a system consisting of n-heptane, benzene, acetone, isopropanol, 0.65 M, pH 7.0 aqueous

potassium phosphate buffer (5:10:2:3:5, v/v/v/v/v). Fractions containing the desired product are concentrated to a solid residue and partitioned between methylene chloride and dilute (pH 9.5) ammonium hydroxide. The organic layer is washed with water and concentrated to yield 14[1(1-hydroxypropyl)]erythromycin A as a white solid.

Although the present invention is described in the Examples listed above in terms of preferred embodiments, they are not to be regarded as limiting the scope of the invention. The above descriptions serve to illustrate the principles and methodologies involved in creating the three types of mutations that can be introduced into the *eryA* segment of the *Sac. erythraea* chromosome that result in the synthesis of novel polyketide products. Although single Type I alterations, leading to the production of 5-oxo-5,6,-dideoxy-3α-mycorosyl erythronolide B, 11-oxo-11-deoxyerythromycin A, 7-hydroxyerythromycin A, 7-oxo-7deoxyerythromycin A, 5-desosaminyl-3-oxo-3-deoxyerythronolide A, and Δ-6,7-anhydro-6-deoxyerythromycin A are specified herein, it is obvious that other Type I changes can be introduced into the *eryA* segment leading to novel polyketide structures. Among the additional Type I alterations that can be obtained are those in which two or more modules are affected leading to the synthesis of novel polyketides. Examples of combinations of two Type I alterations leading to useful compounds include but are not limited to: mutants of the the β-ketoreductase of module 2 (KR2) and the β-ketoreductase of module 4 (KR4) leading to the formation of 7,11-dioxo-7,11-dideoxyerythromycin A; mutants of KR2 and the β-ketoreductase of module 6 (KR6) leading to the formation of 3,11-dioxo-3,11-dideoxy-5-desosaminylerythronolide A; mutants of KR2 and the dehydratase of module 4 (DH4) leading to the synthesis of 7-hydroxy-11-oxo-11-deoxyerythromycin A; mutants of KR2 and the enoylreductase of module 4 (ER4) leading to the synthesis of Δ-6,7-anhydro-11-oxo-11-deoxyerythromycin A; mutants of KR4 and KR6 leading to the synthesis of 3,7-dioxo-3,7-dideoxy-5-desosaminylerythronolide A; mutants of KR6 and DH4 leading to the synthesis of 3-oxo-3-deoxy-5-desosaminyl-7-hydroxyerythronolide A; mutants of KR6 and ER4 leading to the synthesis of 3-oxo-3-deoxy-5-desosaminyl-Δ-6,7-anhydroerythronolide A. Examples of combinations of three Type I alterations leading to the synthesis of novel polyketides include but are not limited to: mutants of KR2, KR4 and KR6 leading to the synthesis of 3,7,11-trioxo-3,7,11-trideoxy-5-

desosaminylerythronolide A; mutants of KR2, KR6 and DH4 leading to the synthesis of 3,11-dioxo-3,11-dideoxy-5-desosaminyl-7-hydroxyerythronolide A; mutants of KR2, KR6 and ER4 leading to the synthesis of 3,11-dioxo-3,11-dideoxy-5-desosaminyl-D-6,7-anhydroerythronolide A. All combinations of two or three Type I mutants, the Sac. erythraea strains that carry said combinations and the corresponding polyketides produced from said strains, therefore, are included within the scope of the present invention.

Although the Type II mutants specified herein have been constructed in the β–ketoacyl ACP synthase of module 1 (KS1) and the β–ketoacyl ACP synthase of module 2 (KS2), other Type II mutants can be constructed in other domains to result in the synthesis of novel polyketide structures upon feeding with appropriate substrate analogs. Other Type II mutants include but are not limited to: inactivation of the either of the acyltransferases or acyl carrier proteins of module 1, or the acyltransferase or acyl carrier protein of module 2, the β–ketoacyl ACP synthase, acyltransferase or acyl carrier protein of module 3, module 4 or module 5. Furthermore, compounds other than (2S,3R,4S,5S)3,5-dihydroxy-2,4-dimethylhexanoic acid-ethyl thioester and (2S,3S,4S,5S)2,4-dimethyl-3-fluoro-5-hydroxyhexanoic acid-ethyl thioester specified herein can be synthesized and fed to strains AKS1 or AKS2 specified herein or other strains that carry other Type II mutations to result in the creation of novel polyketides that are within the scope of the present invention.

Although two examples of Type III alterations are specified herein, it is apparent to those skilled in the art that many other examples of Type III changes are possible. Strains of *Sac. erythraea* carrying changes of this type offer the very high potential for the production of novel polyketides of specified structure, since they do not require synthetic substrates as do Type II mutants and they are not limited to the formation of derivatives of erythromycin, as in the case of Type I mutants. In the embodiments of Type III mutants specified herein, we have illustrated how a second copy of a complete module can be introduced at a desired position by gene conversion to result in the synthesis of 14-(1-propyl)erythromycin A or 14-[1(1-hydroxypropyl])erythromycin A. These alterations make use of the high conservation and simultaneous lack of specificity of the β–ketoacyl ACP synthases of modules 1 and 2, thereby making possible the construction of hybrid β–ketoacyl ACP synthase functions consisting of portions of proteins derived from different modules. Those skilled in the

art understand, therefore, that it is possible, as exemplified for KS1 and KS2, to delete a small portion of the β–ketoacyl ACP synthase of other modules and to construct strains carrying such alterations which can then be employed as hosts for introducing at the deleted β–ketoacyl ACP synthase location a second copy of any homologous module. Furthermore, as exemplified herein, it is also possible to delete any segment of *eryA* by ligation of two non-contiguous PCR-generated fragments and to subsequently construct strains, therefore, devoid of any or all portions of any module. Such strains deleted of a full module can be employed for reintroduction of either the same or a different module at a different location. It is possible, therefore, to determine the novel structures desired and then create a series of *Sac. erythraea* strains containing the corresponding arrangements of *eryA* modules that would produce said novel structures that are included within the scope of the present invention. Additional examples of novel compounds produced from the construction of Type III alterations include but are not limited to 11-deoxyerythromycin, resulting from the insertion of the *eryA* segment encoding DH4 and ER4 in module 2.

Moreover, it will also be apparent that two or more modules can be excised and introduced into various sites of the *Sac. erythraea* chromosome to produce novel polyketides of predicted structure such as the introduction of the *eryA* segment encoding DH4 and ER4 in both module 1 and module 2 to result in the production of 14(R)[1-hydroxypropyl]11-deoxyerythromycin A. All combinations, therefore, of Type III alterations and the strains of *Sac. erythraea* that carry said alterations as well as the polyketides produced from said strains are included within the scope of the present invention.

In addition, it is also possible to create combinations of Type I, Type II and Type III alterations and insert such alterations into *Sac. erythraea* to produce novel polyketides. Examples of such combinations include but are not limited to the following. The combination of a Type I alteration, such as an alteration in DH4 and a Type II alteration, such as a mutation in the KS1 to result in the formation of (14S,15S)14-[1-hydroxyethyl]-7-hydroxyerythromycin A when the strain of *Sac. erythraea* carrying such alterations is fed with the compound (2S,3R,4S,5S)3,5-dihydroxy-2,4-dimethylhexanoic acid ethyl ester. The combination of a Type I alteration, such as an alteration in DH4 and a Type III alteration, such as found in *Sac. erythraea* strain AM412, wherein a copy of the DNA segment of

module 4 is introduced in module 1, such that the *Sac. erythraea* strain so constructed produces the compound 7-hydroxy-14-propylerythromycin A. All combinations of two or more alterations of Type I, Type II and Type III alterations, the *Sac. erythraea* strains that carry such alterations, and the polyketides produced from such strains are included within the scope of the present invention. It will also occur to those skilled in the art that novel structures can be produced by altering the specificity of the acyltransferase functions in any module. Examples include: replacement of the acyltransferase domains of modules 1, 2, 3, 4, 5,or 6 in *eryA* with those of modules 4, 4, 2, 2, 2, and 4, respectively, to result in the production of 12-epierythromycin A,10-epierythromycin A, 8-epierythromycin A, 6-epierythromycin A, 4-epierythromycin A and 2-epierythromycin A, respectively, that are included within the scope of the present invention.

It should be emphasized that the introduction of an entire *eryA* module at a different location, as exemplified for the construction of *Sac. erythraea* strains AM412 and AM512 in Examples 29 and 35, respectively, does not rely on homologous recombination between the incoming *eryA* module and the host chromosome. Rather, gene conversion of the host allele with the *eryA* allele residing on the multicopy plasmid requires DNA sequences homologous to the host allele flanking the incoming module. Thus, any module carrying the desired specificities, either from homologous or heterologous sources, can be employed in gene conversion of the host allele, provided that is flanked by segments of homology. It will occur to those skilled in the art, therefore, that, given the large number of natural polyketide molecules existing, a wide variety of additional novel molecules of predicted structure can be produced in Type III mutants containing an additional module of desired specificities or where an endogenous module is replaced by an exogenous one. The length of the acyl chain can be easily controlled by suitably changing the number of modules involved in its synthesis. Similarly, the introduction of keto, hydroxy, enoyl, or methylene groups at specific points along the acyl chain can be easily achieved by introducing the proper b-carbon processing functions (β-ketoreductase, dehydratase and enoylreductase) in the required modules. Exogenous modules constitute the source of specificities for starter and extender units other than those employed by *Sac. erythraea* for erythromycin biosynthesis, making it thereby possible to employ, for example, malonylCoA or (2R)- or (2S)ethylmalonylCoA, etc.

as extender units, and acetyl CoA, butyryl CoA, etc. as the starter unit. The result will be the formation of erythromycin analogs containing the desired functional groups and side chains with the desired stereochemistry. As an extension of the examples reported with *eryA* , the construction of a *Sac. erythraea* strain carrying a heterologous module inserted into *eryA* requires: (i) cloning of the genes from any other Actinomyces producing a polyketide with desired structural features; (ii) mapping of the modular organization of the cloned genes by low stringency hybridization and restriction analysis; (iii) locating the module carrying the desired specificities by partial sequencing; (iv) precise excision of the desired genetic element and cloning into a vector suitable for gene conversion; (v) construction and transformation of a *Sac. erythraea* strain suitable for gene conversion and screening for the novel compound. Any module, or portion thereof, can thus be precisely excised from the genome of a polyketide-producing microorganism and introduced into suitable *Sac. erythraea* strains to create a novel polyketide of predicted structure. Thus, replacement of the acyltransferase segments of modules 1, 2, 3, 4, 5,or 6 in *eryA* with the acyltransferase segment specific for malonyl CoA, such as can be found in the polyketide synthase genes for the synthesis of pikromycin in Streptomyces venezuelae, to result in the synthesis of 12-norerythromycin A, 10-norerythromycin A, 8-norerythromycin A, 6-norerythromycin A, 4-norerythromycin A and 2-norerythromycin A, respectively, that are included within the scope of the present invention. In addition, replacement of the acyltransferase segments of modules 1, 2, 3, 4, 5,or 6 in *eryA* with an acyltransferase specific for (2R)-ethylmalonyl CoA, such as can be found in the polyketide synthase genes for the synthesis of spiramycin in Streptomyces ambofasciens, will result in the formation of 12-homoerythromycin A, 10-homoerythromycin A, 8-epihomoerythromycin A, 6-epihomoerythromycin A, 4-epihomoerythromycin A and 2-homoerythromycin A, respectively, all of which are included within the scope of the present invention. Similarly, introduction of acyltransferase segments carrying desired specificities for the starter or extender unit into *eryA* DNA that results in the synthesis of novel compounds are included within the scope of the present invention. The erythromycin analogs produced by the method of this invention are structurally similar to known antibacterial and prokinetic agents.

It will also occur to those skilled in the art that genetic manipulations described herein need not be limited to *Sac. erythraea*.

Suitable hosts are any other polyketide-producing Actinomyces where DNA can be precisely inserted into the chromosome. Hence, the choice of a convenient host is based solely on the relatedness of the novel polyketide to a natural counterpart so as to minimize the number of module rearrangements required for its biosynthesis. Therefore, Type I, Type II and Type III alterations can be constructed in other Actinomyces employing either endogenous or exogenous modules to produce novel polyketides employing strategies analogous to those described herein for *Sac. erythraea*. Thus all Type I, Type II or Type III mutations or various combinations thereof constructed in other actinomycetes according to the principles described herein, and the respective polyketides produced from such strains, are included within the scope of the present invention. Examples of polyketides that can be altered by creating Type I, Type II or Type III changes in the producing microorganisms include, but are not limited to macrolide antibiotics such as erythromycin, tylosin, spiramycin, etc.; ansamacrolides such as rifamycins, maytansines, etc.; polyketide antibiotics such as tetracycline; polyethers such as monesin, salinomycin, etc.; polyenes such as candicidin, amphothericins; immunosuppressants such as FK506, ascomycin, rapamycin, etc. and other complex polyketides such as avermectin.

Whereas the novel derivatives or modifications of erythromycin described herein have been specified as the A derivatives, such as 7-hydroxyerythromycin A, 11-oxo-11-deoxyerythromycin A, 14[1(1-hydroxypropyl)]erythromycin A, etc., those skilled in the art understand that the wild type strain of *Sac. erythraea* produces a family of erythromycin compounds, including erythromycin A, erythromycin B, erythromycin C and erythromycin D. Thus, modified strains of *Sac. erythraea*, such as strain AKR2, for example, would be expected to produce the corresponding members of the 11-oxo-11-deoxyerythromycin family, including 11-oxo-11-deoxyerythromycin A, 11-oxo-11-deoxyerythromycin B, 11-oxo-11-deoxyerythromycin C, and 11-oxo-11-deoxyerythromycin D. Similarly, strain AM412 would be expected to produce not only 14(1-propyl)erythromycin A but also the other members of the 14(1-propyl)erythromycin family including 14(1-propyl)erythromycin B, 14(1-propyl)erythromycin C and 14(1-propyl)erythromycin D. Similarly, all other modified strains of *Sac. erythraea* described herein that produce novel erythromycin derivatives would be expected to produce the A, B, C, and D forms of said derivatives. Therefore, all members of the family of

each of the novel polyketides described herein are included within the scope of the present invention.

Variations and modifications of the methods for obtaining the desired plasmids, hosts for cloning and choices of vectors and segments of *eryA* DNA to clone and modify, other than those described herein that result in substantially the same strains and same products as those described herein will occur to those skilled in the art. For example, although we have described the use of the plasmids pWH3 and pWHM4 as E. coli-Sac. erythraea shuttle vectors, other vectors can be employed wherein all or part of pWHM3 or pWHM4 is replaced by other DNA segments that function in a similar manner, such as replacing the pUC19 component of pWHM3 and pWHM4 with pBR322, available from BRL, employing different segments of the pIJ101 or pJV1 replicons in pWHM3 and pWHM4, respectively, or employing selectable markers other than thiostrepton- and ampicillin-resistance. These are just few of a long list of possible examples all of which are included within the scope of the present invention. Similarly, the segments of the *eryA* locus subcloned into pWHM3 for generating strains AKS1, AKS2, etc. specified herein can readily be substituted for other segments of different length encoding the same functions, either produced by PCR-amplification of genomic DNA or of an isolated clone, or by isolating suitable restriction fragments from *Sac. erythraea*. In the same way, it is possible to create *eryA* strains carrying mutations functionally equivalent to those described herein by deleting different portions of the corresponding genes, by creating insertions into them, or by site-directed mutagenesis of specific nucleotide residues. Moreover, *Sac. erythraea* strains with mutant alleles other than the β–ketoacyl ACP synthase portions of *eryA* can be employed as hosts for gene conversion; Type III mutants can be constructed by double reciprocal crossover as exemplified for Type I and Type II mutants rather than by the gene conversion method described herein. Additional modifications include changes in the restriction sites used for cloning or in the general methodologies described above. All such changes are included in the scope of the invention. It will also occur to those skilled in the art that different methods are available to ferment *Sac. erythraea*, to extract the novel polyketides specified herein, and to synthesize substrate analogs, and that all such methods are also included within the scope of the present invention.

It will be apparent that many modifications and variations of the invention as set forth herein are possible without departing from the spirit and scope thereof, and that, accordingly, such limitations are imposed only as indicated by the appended claims.

What is claimed is:

1. A method for directing the biosynthesis of specific polyketide analogs by genetic manipulation of a polyketide-producing microorganism, said method comprising the steps of:

(1) isolating a polyketide biosynthetic gene-containing DNA sequence;

(2) identifying enzymatic activities associated within said gene-containing DNA sequence;

(3) introducing one or more specified changes into said gene-containing DNA sequence which codes for one of said enzymatic activities resulting in an altered DNA sequence;

(4) introducing said altered DNA sequence into a polyketide-producing microorganism to replace the original sequence;

(5) growing a culture of the altered microorganism under conditions suitable for the formation of the specific polyketide analog; and

(6) isolating said specific polyketide analog from the culture.

2. The method of claim 1 wherein said polyketide biosynthetic gene-containing DNA sequence comprises genes which encode the enzymatic activities comprising a polyketide synthase.

3. The method of claim 2, wherein said polyketide synthase enzymatic activities comprise β-ketoreductase, dehydratase, acyl carrier protein, enoylreductase, β-ketoacyl ACP synthase, and acyltransferase.

4. The method of claim 1 wherein said alteration which occurs in the DNA sequence results in the inactivation of one or more enzymatic activities involved in the processing of the β-carbonyl of said polyketide.

5. The method of claim 4, wherein said inactivated enzymatic activities affecting the processing of the β-carbonyl of said polyketide comprise β-ketoreductase, dehydratase, and enoylreductase.

6. The method of claim 4 wherein said alteration in the DNA sequence results in the addition of one or more enzymatic activities involved in the β-carbonyl processing of said polyketide.

7. The method of claim 6 wherein said additional enzymatic activities are selected from the group consisting of β-ketoreductase, β-ketoreductase and dehydratase, and β-ketoreductase, dehydratase and enoylreductase.

8. The method of claim 1 wherein said alteration occurring in the DNA segment results in the inactivation of one or more enzymatic activities involved in the condensation of carbon units to the nascent polyketide structure.

9. The method of claim 8 wherein said enzymatic activities affecting the condensation of carbon units to the nascent polyketide structure comprise β-ketoacyl ACP synthase, acyl carrier protein and acyltransferase.

10. The method of claim 1 wherein said alteration in the DNA sequence results in the change of the length of the polyketide synthesized.

11. The method of claim 10 wherein said alteration results in the increase of the length of the polyketide.

12. The method of claim 11 wherein said alteration comprises the addition of DNA sequences encoding the enzymatic activities consisting of acyltransferase, acyl carrier protein and β-ketoacyl ACP synthase.

13. The method of claim 10 wherein said alteration results in the decrease of the length of the polyketide.

14. The method of claim 13 wherein said alteration consists of the deletion of a DNA segment between two sequences encoding corresponding enzymatic activities.

15. The method of claim 14 wherein said corresponding enzymatic activities are selected from the group consisting of β-ketoreductases, dehydratases, acyl carrier proteins, enoylreductases, β-ketoacyl ACP synthases, and acyltransferases.

16. The method of claim 1 wherein said alteration consists in the replacement of the DNA segment encoding an acyltransferase with a DNA segment encoding an acyltransferase of different specificity.

17. The method of claim 1 wherein said DNA sequence is isolated from a species from the *Actinomycetales* family.

18. The method of claim 17 wherein said DNA sequence is isolated from a genus selected from the group consisting of *Actinomyces, Dactylosporangium, Micromonospora, Nocardia, Sac., Streptoverticillium,* and *Streptomyces.*

19. The method of claim 17 wherein said genus is selected from the group consisting of *Saccharapolyspora* and *Streptomyces.*

20. The method of claim 19 wherein said genus is *Saccharapolyspora* and the species is *erythraea.*

21. The method of claim 19 wherein said genus is *Streptomyces* and the species is *hygroscopicus.*

22. The method of claim 1 wherein said polyketide is selected from the group consisting of macrolides, tetracyclines, polyethers, polyenes, ansaymcins and derivatives or analogs thereof.

23. The method of claim 22 wherein said polyketide is a macrolide.

24. The method of claim 23 wherein said macrolide is an erythromycin.

25. The method of claim 24 wherein said erythromycin analog is selected from the group consisting of 11-oxo-11-deoxyerythromycin A, 7-hydroxyerythromycin A, 6-deoxy-7-hydroxyerythromycin A, 7-oxoerythromycin A, 3-oxo-3-deoxy-5-desosaminylerythronolide A, Δ-6,7-anhydroerythromycin A, ((14S,15S)14(1-hydroxyethyl)erythromycin A, 11-epifluoro-15-norerythromycinA, 14-(1-propyl)erythromycin A, 14(1-propyl)erythromycin A, and 14[1(1-hydroxypropyl)]erythromycin A.

26. The method of claim 1 wherein said DNA sequence, designated *eryA,* encodes the enzymatic activities associated with the formation of 6-deoxyerythronolide B.

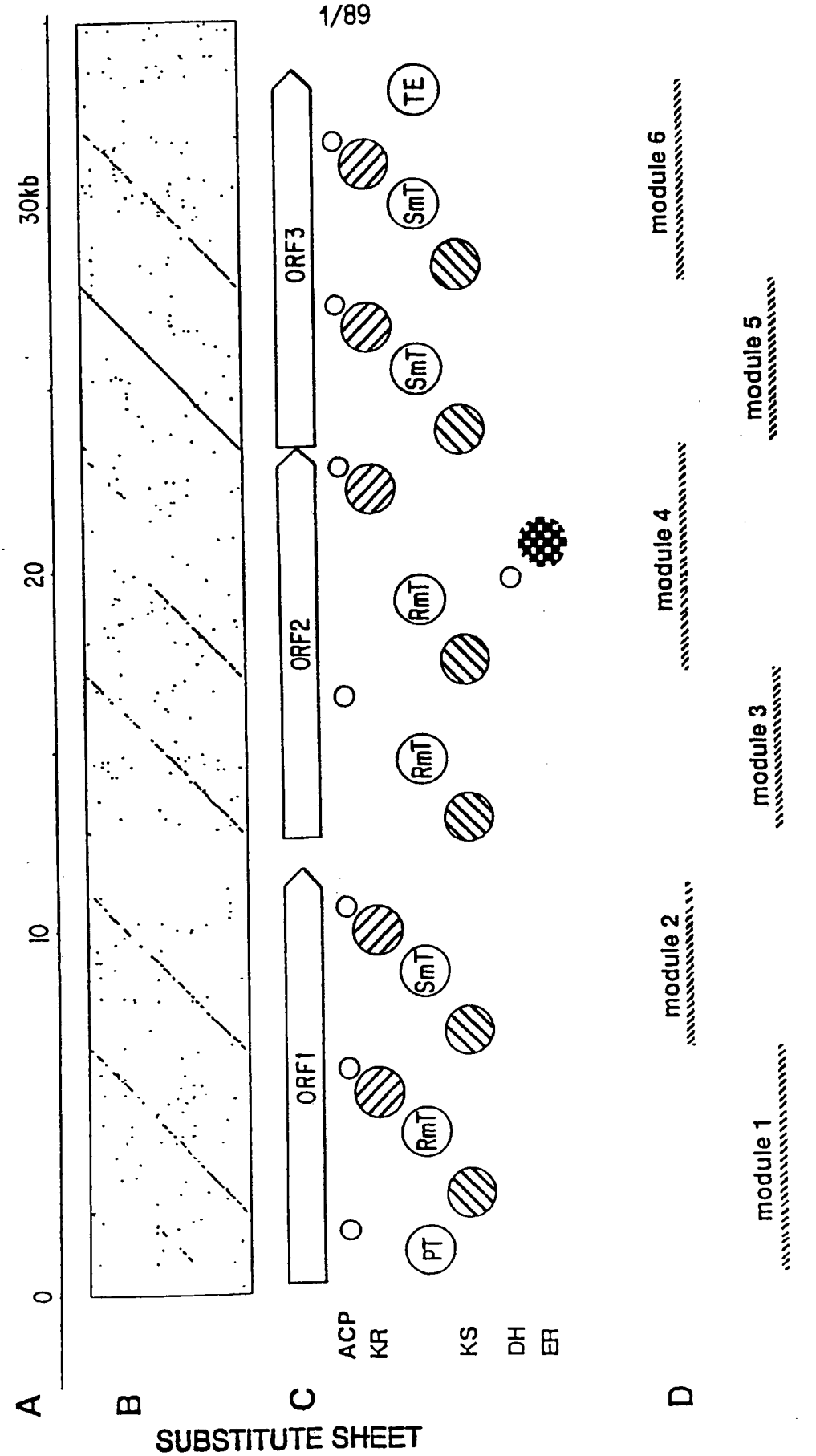
27. The method of claim 26 wherein said DNA sequence comprises: the DNA sequence of Figure 2.

28. The method of claim 1 wherein said gene-containing DNA sequence encodes one or more enzymatic activities in the rapamycin biosynthetic pathway.

29. The method of claim 23 wherein said macrolide is a rapamycin analog.

30. A compound selected from the group consisting of 7-hydroxyerythromycin A; 6-deoxy-7-hydroxyerythromycin A; 7-oxoerythromycin A, 3-oxo-3-deoxy-5-desosaminyl-erythronolide A; Δ-6,7-anhydroerythromycin A; ((14S,15S)14(1-hydroxyethyl)erythromycin A; 11-epifluoro-15-norerythromycin A; 14-(1-propyl)erythromycin A; 14(1-propyl)erythromycin A; and 14[1(1-hydroxypropyl)]erythromycin A.

FIG.1


A
B
C
D
0
10
20
30kb
ORF1
ORF2
ORF3
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RmT
SmT
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module 4
module 5
module 6

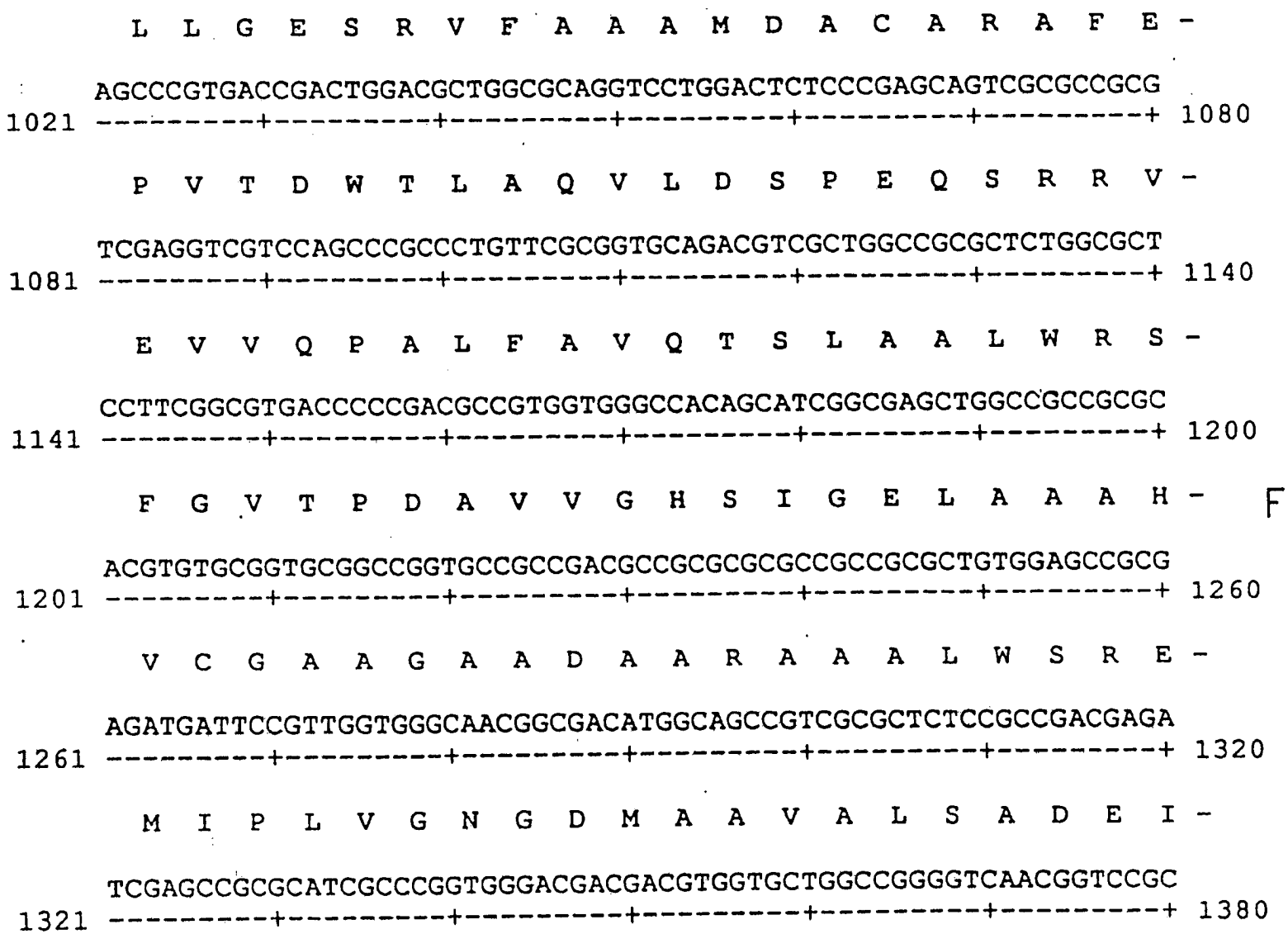

FIG.2-1

SUBSTITUTE SHEET

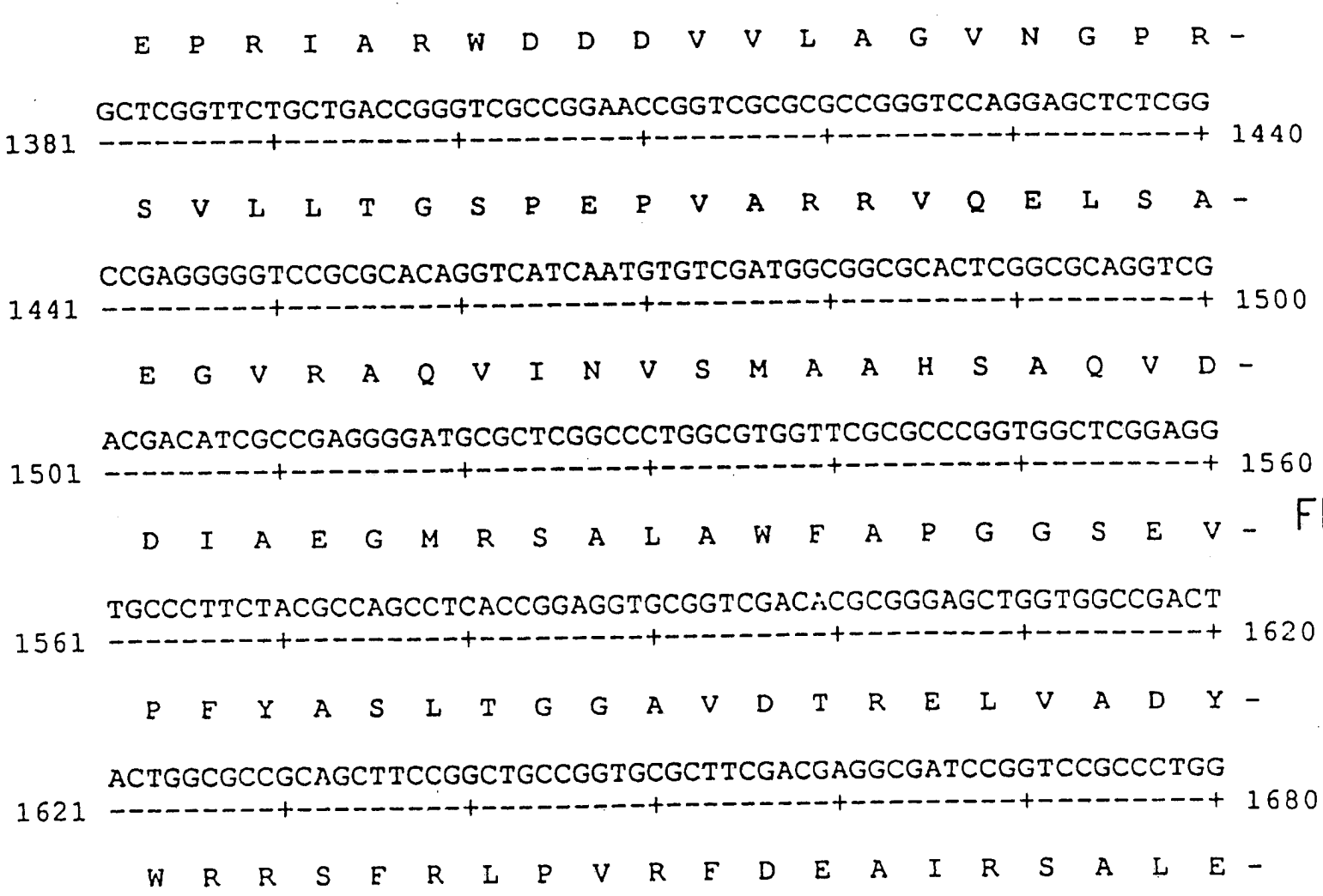

FIG.2-2

```
     AGGTCGGTCCCGGCACGTTCGTCGAAGCGAGCCCGCACCCGGTGCTGGCCGCCGCGCTCC
1681 ---------+---------+---------+---------+---------+---------+ 1740

       V  G  P  G  T  F  V  E  A  S  P  H  P  V  L  A  A  A  L  Q -

     AGCAGACGCTCGACGCCGAGGGCTCCTCGGCCGCGGTGGTCCCGACGCTGCAACGCGGGC
1741 ---------+---------+---------+---------+---------+---------+ 1800

       Q  T  L  D  A  E  G  S  S  A  A  V  V  P  T  L  Q  R  G  Q -

     AGGGCGGCATGCGGCGGTTCCTGCTGGCCGCGGCCCAGGCGTTCACCGGCGGCGTGGCCG
1801 ---------+---------+---------+---------+---------+---------+ 1860

       G  G  M  R  R  F  L  L  A  A  A  Q  A  F  T  G  G  V  A  V -

     TCGACTGGACCGCCGCCTACGACGACGTGGGGCCGAACCCGGCTCTCTGCCGGAGTTCGC
1861 ---------+---------+---------+---------+---------+---------+ 1920

       D  W  T  A  A  Y  D  D  V  G  P  N  P  A  L  C  R  S  S  R -

     GCCGGCCGAGGAGGAAGACGAGCCGGCCGAGTCCGGCGTCGACTGGAACGCGCCACCGCA
1921 ---------+---------+---------+---------+---------+---------+ 1980

       R  P  R  R  K  T  S  R  P  S  P  A  S  T  G  T  R  H  R  T -

     CGTGCTGCGAGCGGCTGCTCGCGGTCGTCAACGGCGAGACCGCCGCGTTGGCGGGCCGCG
1981 ---------+---------+---------+---------+---------+---------+ 2040

       C  C  E  R  L  L  A  V  V  N  G  E  T  A  A  L  A  G  R  E -
```

FIG.2-3

SUBSTITUTE SHEET

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                                                                    AAGCCGACGCCGAGGCCACGTTCCGCGAGCTGGGGCTGGACTCGGTGCTGGCCGCGCAGC
2041 ----------+---------+---------+---------+---------+---------+---------+---------+---------+---------+---------+---------+ 2100
                A  D  A  E  A  T  F  R  E  L  G  L  D  S  V  L  A  A  Q  L  -

                                                                    TGCGCGCCAAGGTGAGCGCCGCGATCGGGCGCGAGGTCAACATCGCCCTGCTCTACGACC
2101 ----------+---------+---------+---------+---------+---------+---------+---------+---------+---------+---------+---------+ 2160
                R  A  K  K  V  S  A  A  I  G  R  E  V  N  I  A  L  L  Y  D  H  -

                                                                    ACCCGACTCCGCGTGCGCTCGCGGAAGCACTCGCGGCGCGGGAACCGAGGTCGCACAACGGG
2161 ----------+---------+---------+---------+---------+---------+---------+---------+---------+---------+---------+---------+ 2220
                P  T  P  R  A  L  A  E  A  L  A  A  G  T  E  V  A  Q  R  E  -

                                                                    AAACCCGCGCGCGGACCAACGAAGCGGCGCCCGGCGAACCGGTCGCGGTCGTCGCGATGG
2221 ----------+---------+---------+---------+---------+---------+---------+---------+---------+---------+---------+---------+ 2280
                T  R  A  R  T  N  E  A  A  P  G  E  P  V  A  V  V  A  M  A  -

                                                                    CCTGCCCGGCTGCCCGGCGGTGTGAGCACCCCGGAGGAGTTCTGGGAGCTGCTGTCGGAGG
2281 ----------+---------+---------+---------+---------+---------+---------+---------+---------+---------+---------+---------+ 2340
                C  R  L  P  G  G  V  S  T  P  E  E  F  W  E  L  L  S  E  G  -

                                                                    GCCGCGACGCGGTCGCGGGACTGCCGACCGACCGCGGCTGGGACCTGGACTCGCTGTTCC
2341 ----------+---------+---------+---------+---------+---------+---------+---------+---------+---------+---------+---------+ 2400
```

FIG.2-4

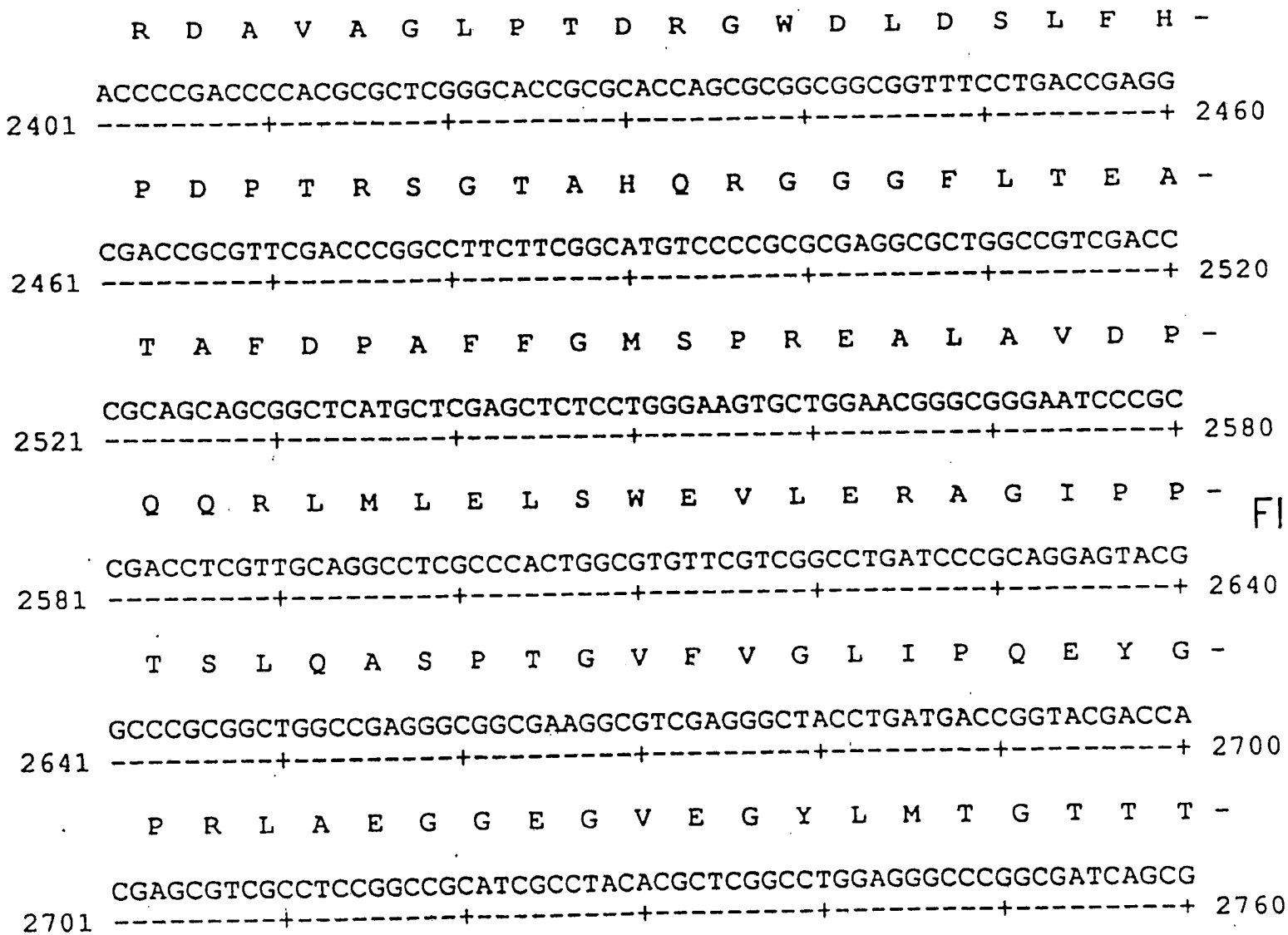

FIG.2—5

SUBSTITUTE SHEET

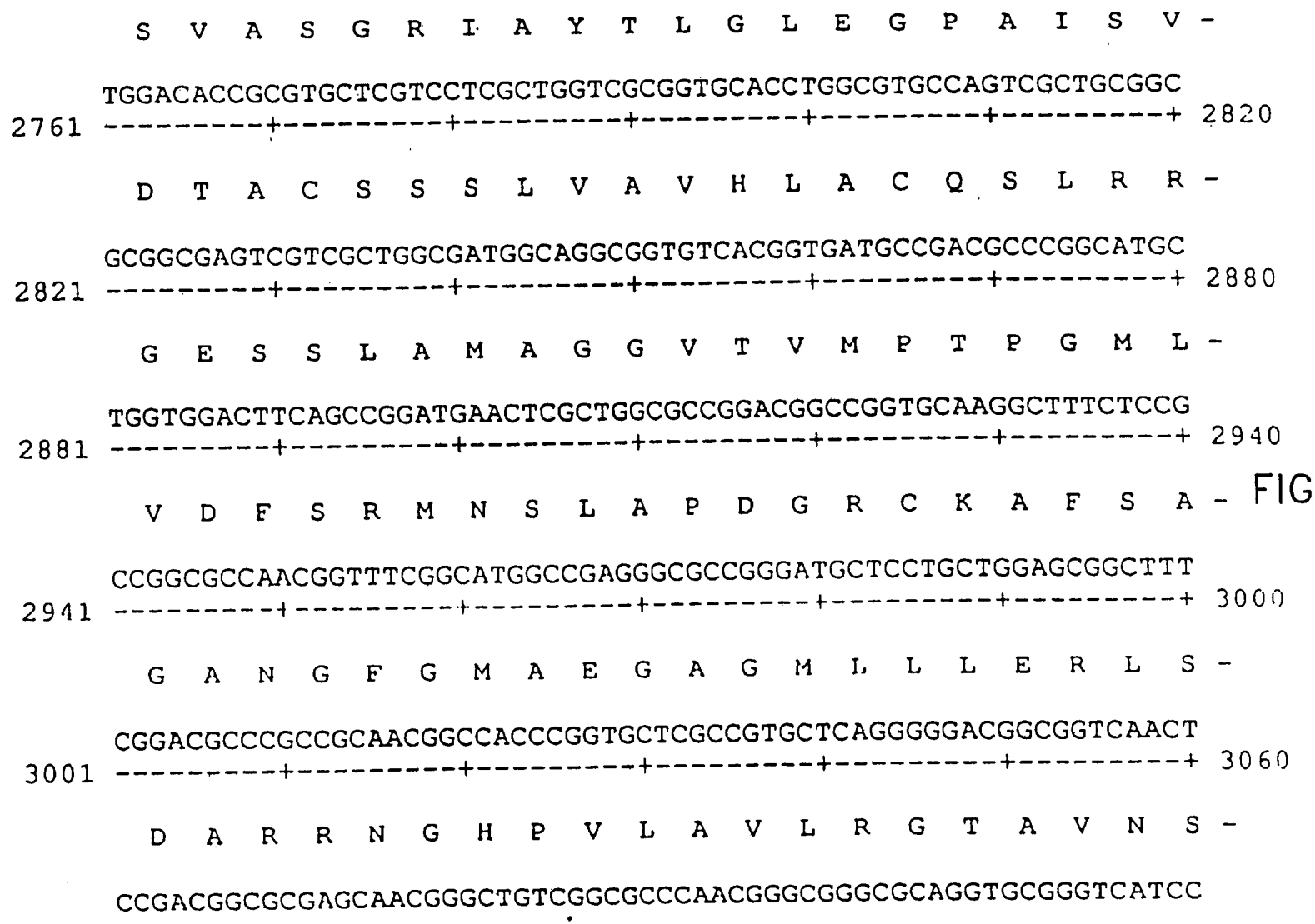
S V A S G R I A Y T L G L E G P A I S V -
TGGACACCGCGTGCTCGTCCTCGCTGGTCGCGGTGCACCTGGCGTGCCAGTCGCTGCGGC
2761 ---------+---------+---------+---------+---------+---------+ 2820
D T A C S S S L V A V H L A C Q S L R R -
GCGGCGAGTCGTCGCTGGCGATGGCAGGCGGTGTCACGGTGATGCCGACGCCCGGCATGC
2821 ---------+---------+---------+---------+---------+---------+ 2880
G E S S L A M A G G V T V M P T P G M L -
TGGTGGACTTCAGCCGGATGAACTCGCTGGCGCCGGACGGCCGGTGCAAGGCTTTCTCCG
2881 ---------+---------+---------+---------+---------+---------+ 2940
V D F S R M N S L A P D G R C K A F S A -
CCGGCGCCAACGGTTTCGGCATGGCCGAGGGCGCCGGGATGCTCCTGCTGGAGCGGCTTT
2941 ---------+---------+---------+---------+---------+---------+ 3000
G A N G F G M A E G A G M L L L E R L S -
CGGACGCCCGCCGCAACGGCCACCCGGTGCTCGCCGTGCTCAGGGGGACGGCGGTCAACT
3001 ---------+---------+---------+---------+---------+---------+ 3060
D A R R N G H P V L A V L R G T A V N S -
CCGACGGCGCGAGCAACGGGCTGTCGGCGCCCAACGGGCGGGCGCAGGTGCGGGTCATCC

FIG.2-6

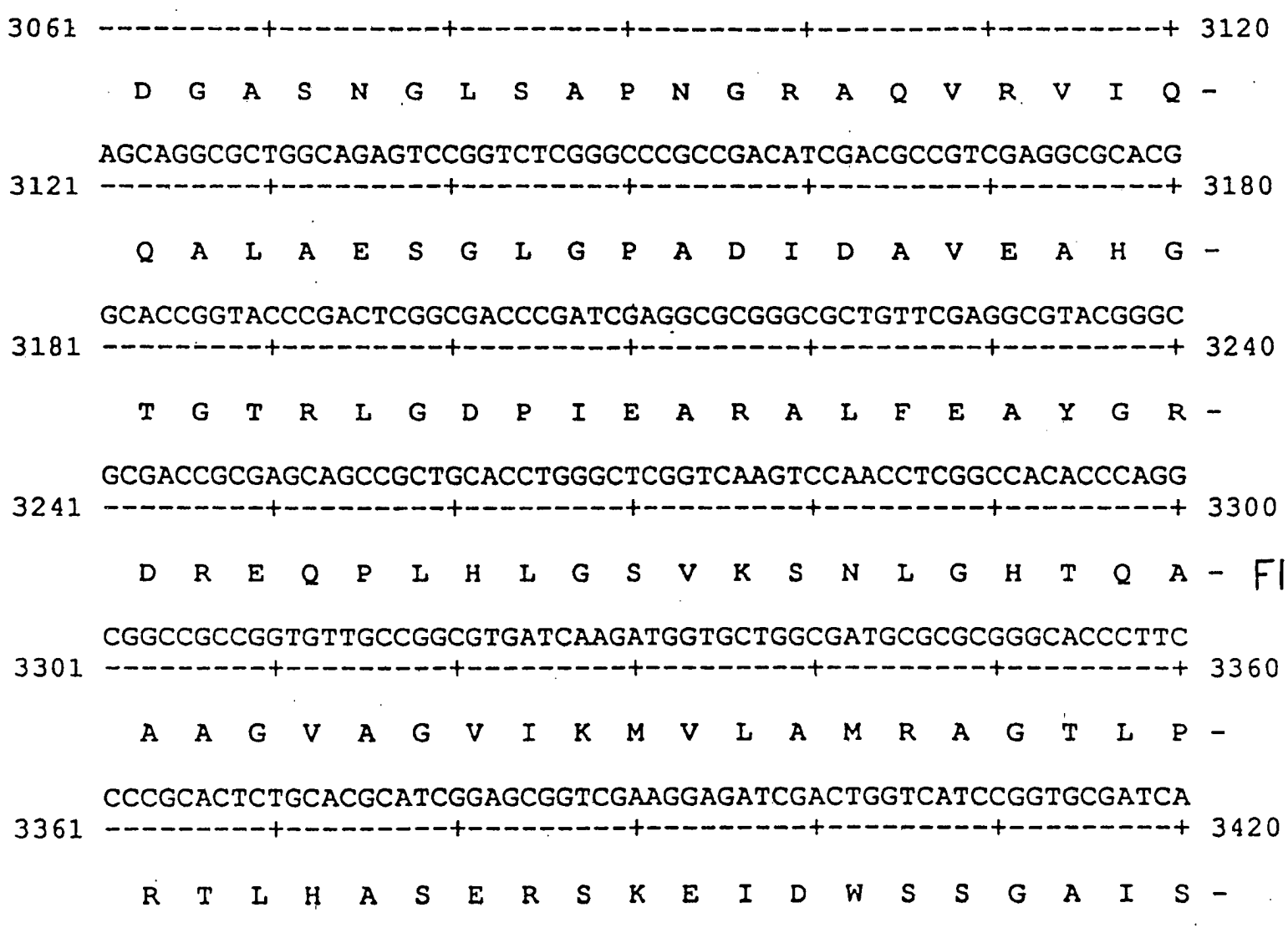

FIG.2-7

SUBSTITUTE SHEET

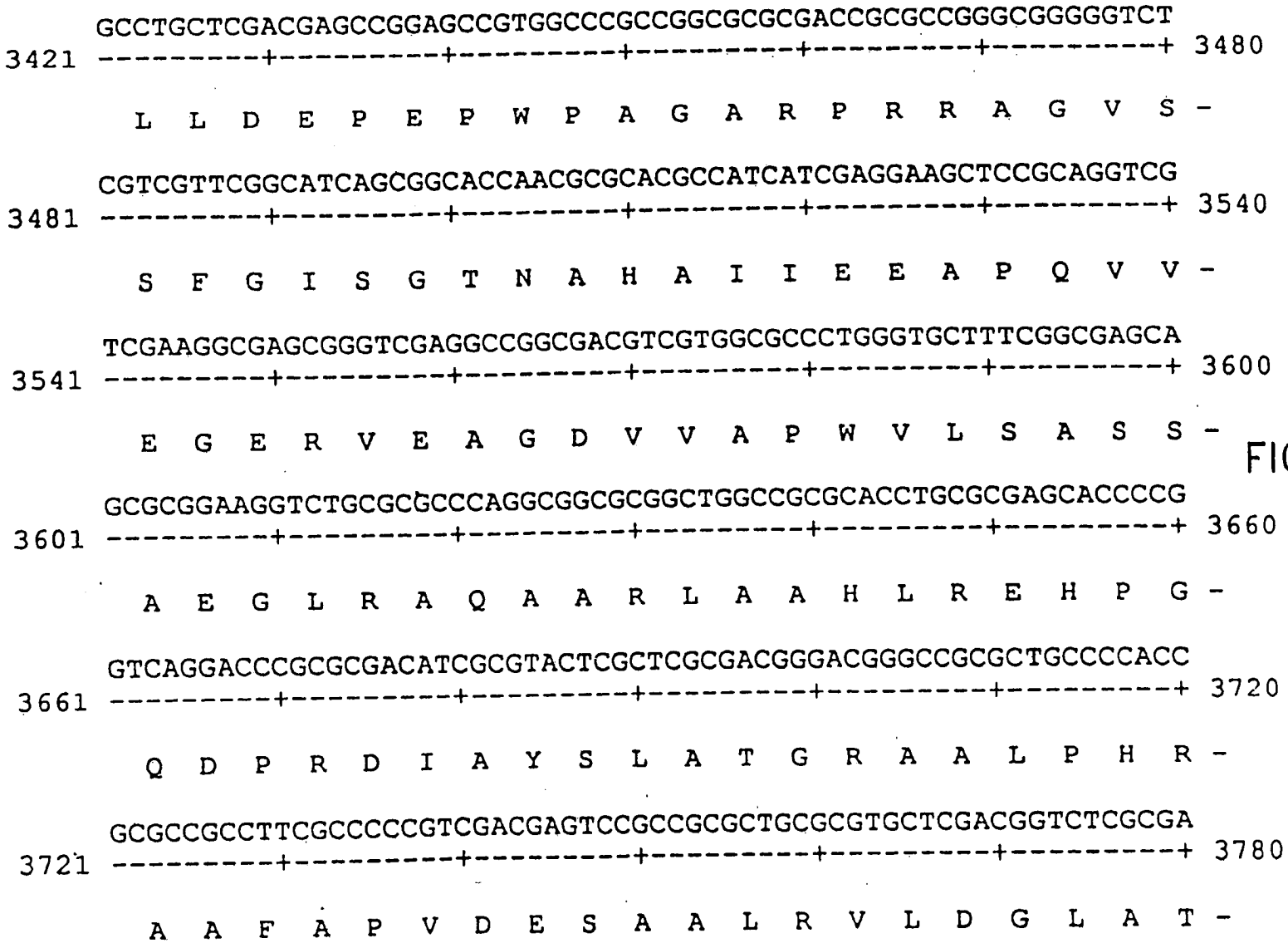
GCCTGCTCGACGAGCCGGAGCCGTGGCCCGCCGGCGCGCGACCGCGCCGGGCGGGGGTCT
3421 ---------+---------+---------+---------+---------+---------+ 3480
L L D E P E P W P A G A R P R R A G V S -
CGTCGTTCGGCATCAGCGGCACCAACGCGCACGCCATCATCGAGGAAGCTCCGCAGGTCG
3481 ---------+---------+---------+---------+---------+---------+ 3540
S F G I S G T N A H A I I E E A P Q V V -
TCGAAGGCGAGCGGGTCGAGGCCGGCGACGTCGTGGCGCCCTGGGTGCTTTCGGCGAGCA
3541 ---------+---------+---------+---------+---------+---------+ 3600
E G E R V E A G D V V A P W V L S A S S -
GCGCGGAAGGTCTGCGCGCCCAGGCGGCGCGGCTGGCCGCGCACCTGCGCGAGCACCCCG
3601 ---------+---------+---------+---------+---------+---------+ 3660
A E G L R A Q A A R L A A H L R E H P G -
GTCAGGACCCGCGCGACATCGCGTACTCGCTCGCGACGGGACGGGCCGCGCTGCCCCACC
3661 ---------+---------+---------+---------+---------+---------+ 3720
Q D P R D I A Y S L A T G R A A L P H R -
GCGCCGCCTTCGCCCCCGTCGACGAGTCCGCCGCGCTGCGCGTGCTCGACGGTCTCGCGA
3721 ---------+---------+---------+---------+---------+---------+ 3780
A A F A P V D E S A A L R V L D G L A T -

FIG.2-8

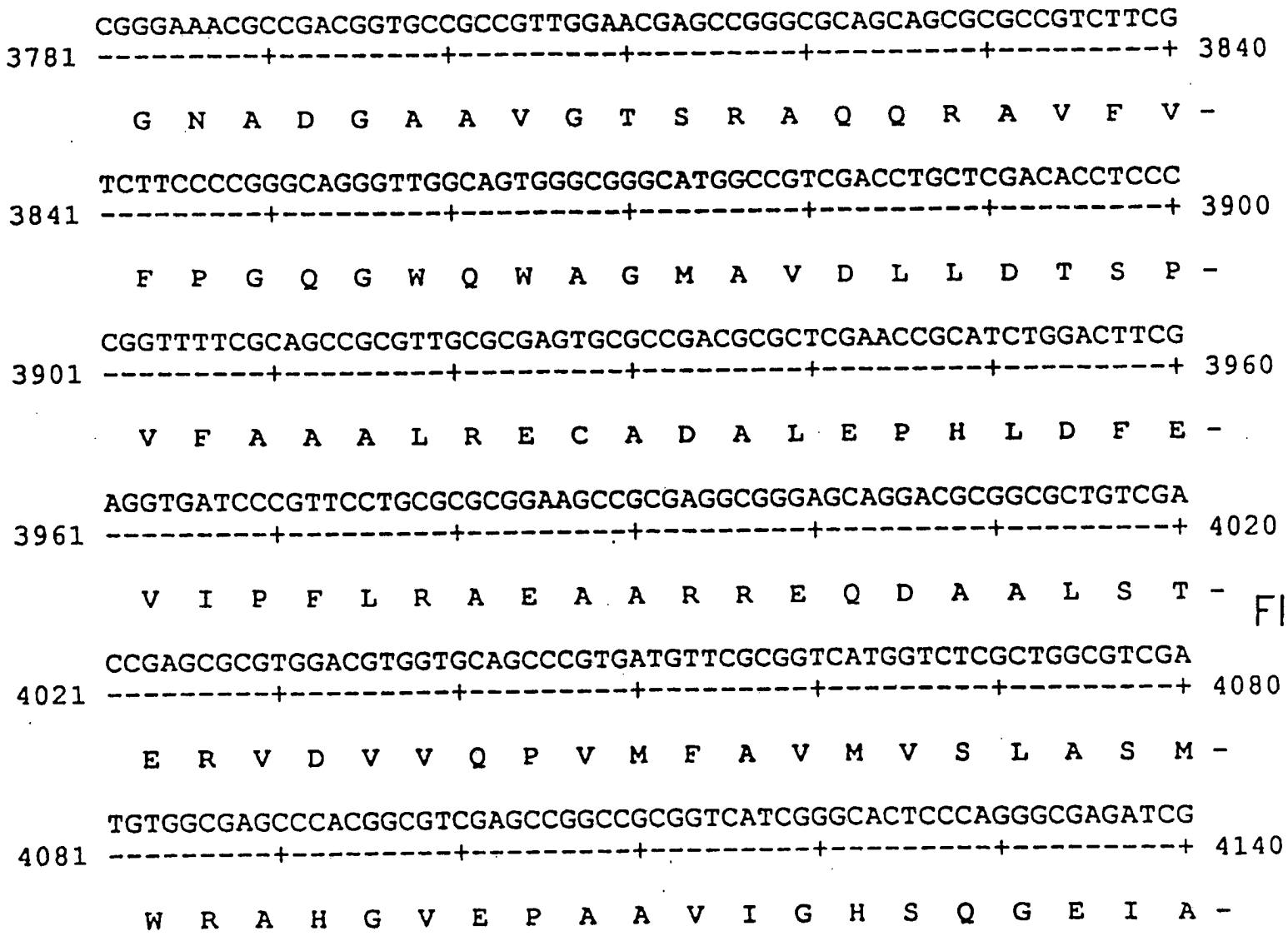

FIG.2-9

```
     CCGCCGCGTGCGTCGCGGGCGCGCTCTCGCTGGACGACGCCGCGCGCGTGGTCGCGCTGC
4141 ---------+---------+---------+---------+---------+---------+ 4200
       A  A  C  V  A  G  A  L  S  L  D  D  A  A  R  V  V  A  L  R -

     GCAGCCGCGTCATCGCCACCATGCCCGGGAACAAGGGCATGGCCTCGATCGCCGCTCCGG
4201 ---------+---------+---------+---------+---------+---------+ 4260
       S  R  V  I  A  T  M  P  G  N  K  G  M  A  S  I  A  A  P  A -

     CCGGCGAAGTCCGCGCGCGAATCGGTGACCGCGTCGAGATCGCCGCCGTCAACGGTCCGC
4261 ---------+---------+---------+---------+---------+---------+ 4320
       G  E  V  R  A  R  I  G  D  R  V  E  I  A  A  V  N  G  P  R -

     GCTCGGTGGTGGTCGCCGGCGACAGCGACGAACTGGACCGGCTGGTCGCTTCCTGCACCA
4321 ---------+---------+---------+---------+---------+---------+ 4380
       S  V  V  V  A  G  D  S  D  E  L  D  R  L  V  A  S  C  T  T -

     CCGAGTGCATCCGCGCCAAGCGGCTGGCCGTGGACTACGCGTCGCACTCCTCGCACGTCG
4381 ---------+---------+---------+---------+---------+---------+ 4440
       E  C  I  R  A  K  R  L  A  V  D  Y  A  S  H  S  S  H  V  E -

     AGACGATCCGAGACGCACTGCACGCCGAGCTGGGAGAGGACTTCCACCCGCTGCCGGGGT
4441 ---------+---------+---------+---------+---------+---------+ 4500
```

FIG.2-10

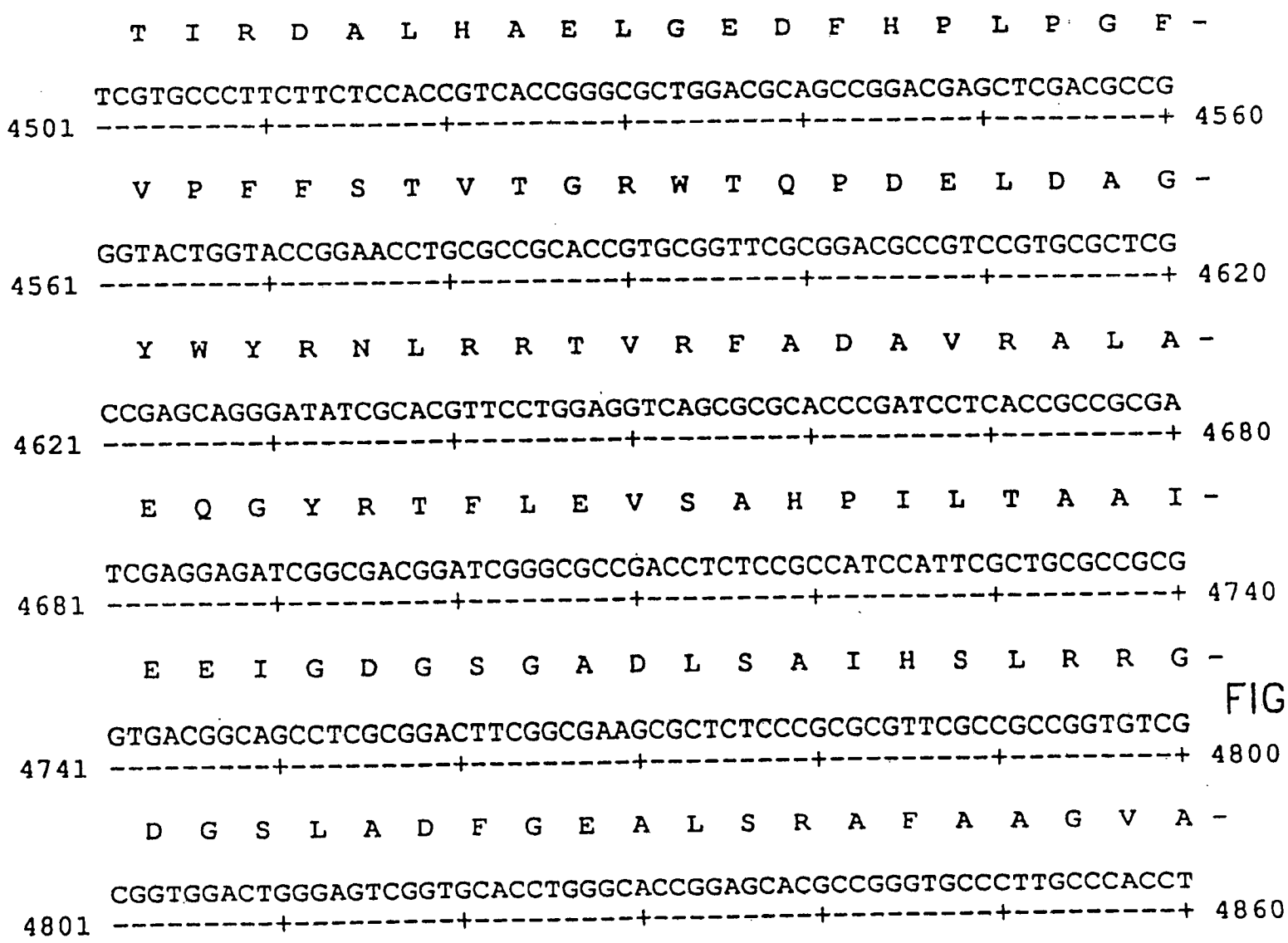

FIG.2-11

SUBSTITUTE SHEET

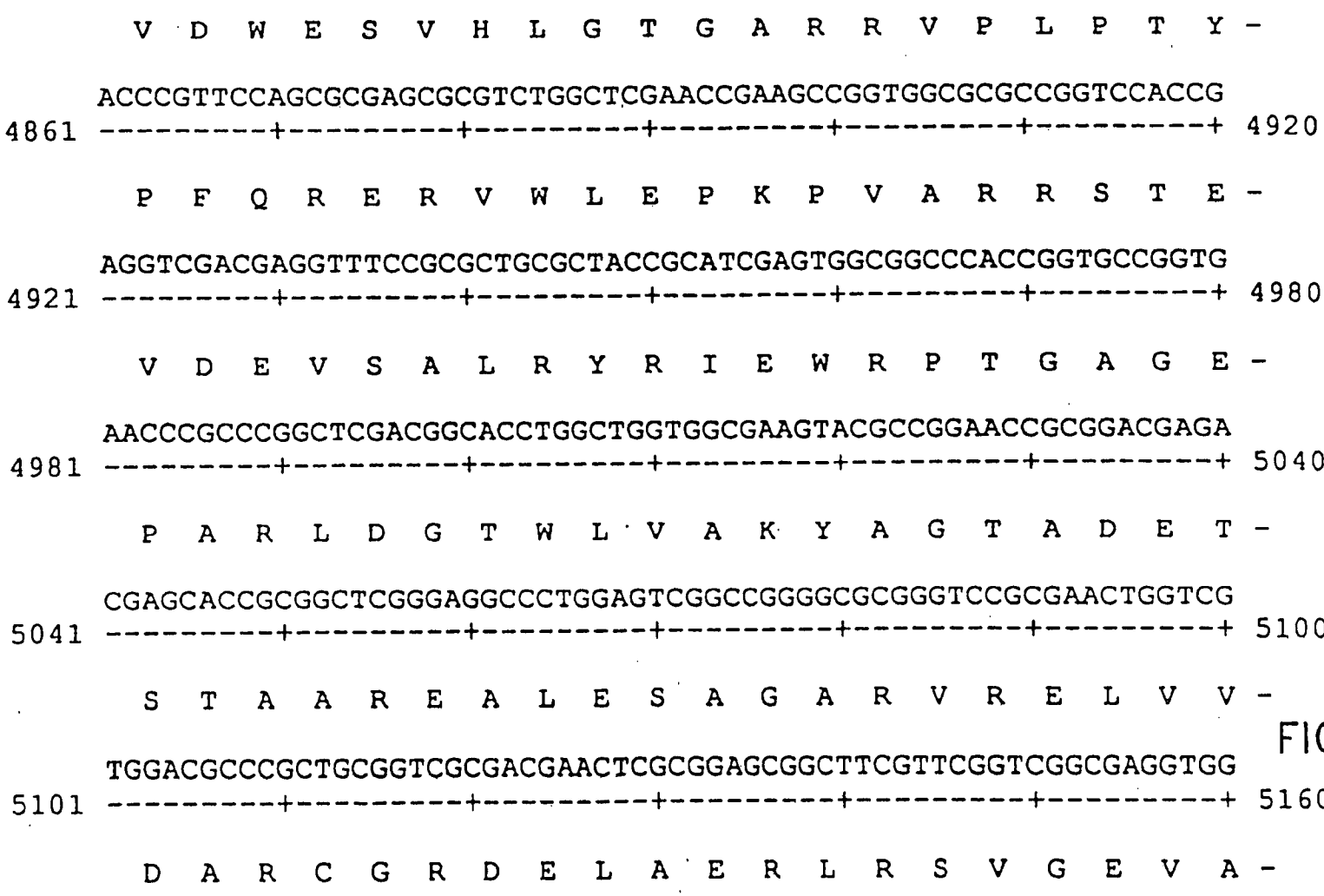

FIG.2—12

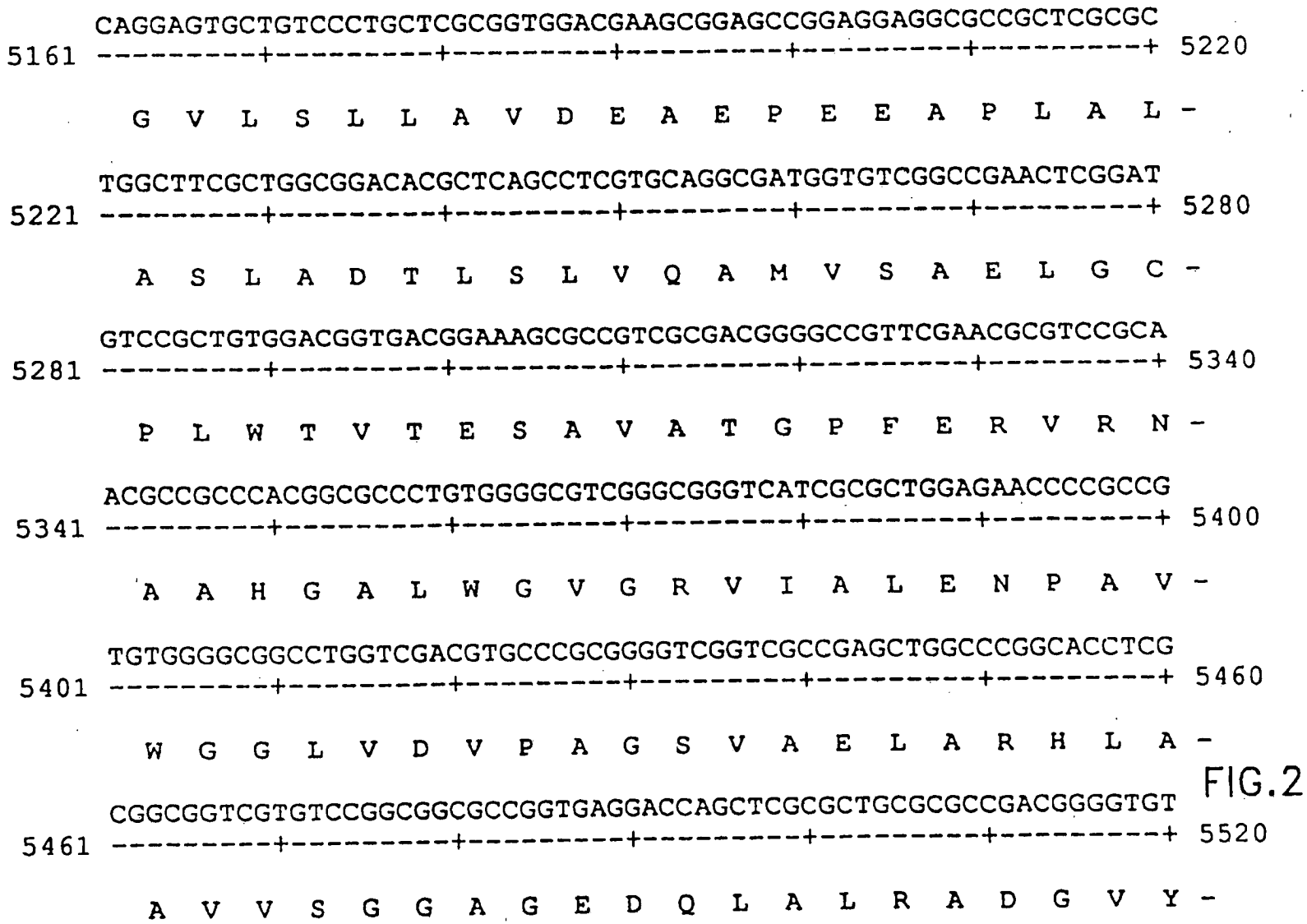

FIG.2-13

```
     ACGGACGCCGGTGGGTGCGCGCGGCGGCCCCGGCGACCGATGACGAGTGGAAACCCACCG
5521 ---------+---------+---------+---------+---------+---------+ 5580
       G  R  R  W  V  R  A  A  A  P  A  T  D  D  E  W  K  P  T  G -

     GAACCGTGCTGGTCACCGGTGGCACGGGCGGTGTCGGCGGGCAGATCGCGCGCTGGCTCG
5581 ---------+---------+---------+---------+---------+---------+ 5640
       T  V  L  V  T  G  G  T  G  G  V  G  G  Q  I  A  R  W  L  A -

     CCCGGCGGGGCGCGCCCCACCTGCTGCTGGTGAGCCGCAGCGGGCCGGACGCGGACGGCG
5641 ---------+---------+---------+---------+---------+---------+ 5700
       R  R  G  A  P  H  L  L  L  V  S  R  S  G  P  D  A  D  G  A -

     CCGGCGAACTGGTCGCCGAGCTCGAGGCGCTGGGCGCCCGGACGACCGTCGCGGCCTGCG
5701 ---------+---------+---------+---------+---------+---------+ 5760
       G  E  L  V  A  E  L  E  A  L  G  A  R  T  T  V  A  A  C  D -

     ACGTGACCGACCGCGAGTCGGTTCGCGAGCTGCTCGGCGGCATCGGTGACGACGTCCCGC
5761 ---------+---------+---------+---------+---------+---------+ 5820
       V  T  D  R  E  S  V  R  E  L  L  G  G  I  G  D  D  V  P  L -

     TCTCGGCGGTGTTCCACGCCGCCGCCACGCTCGACGACGGCACCGTGGACACCCTCACCG
5821 ---------+---------+---------+---------+---------+---------+ 5880
```

FIG.2-14

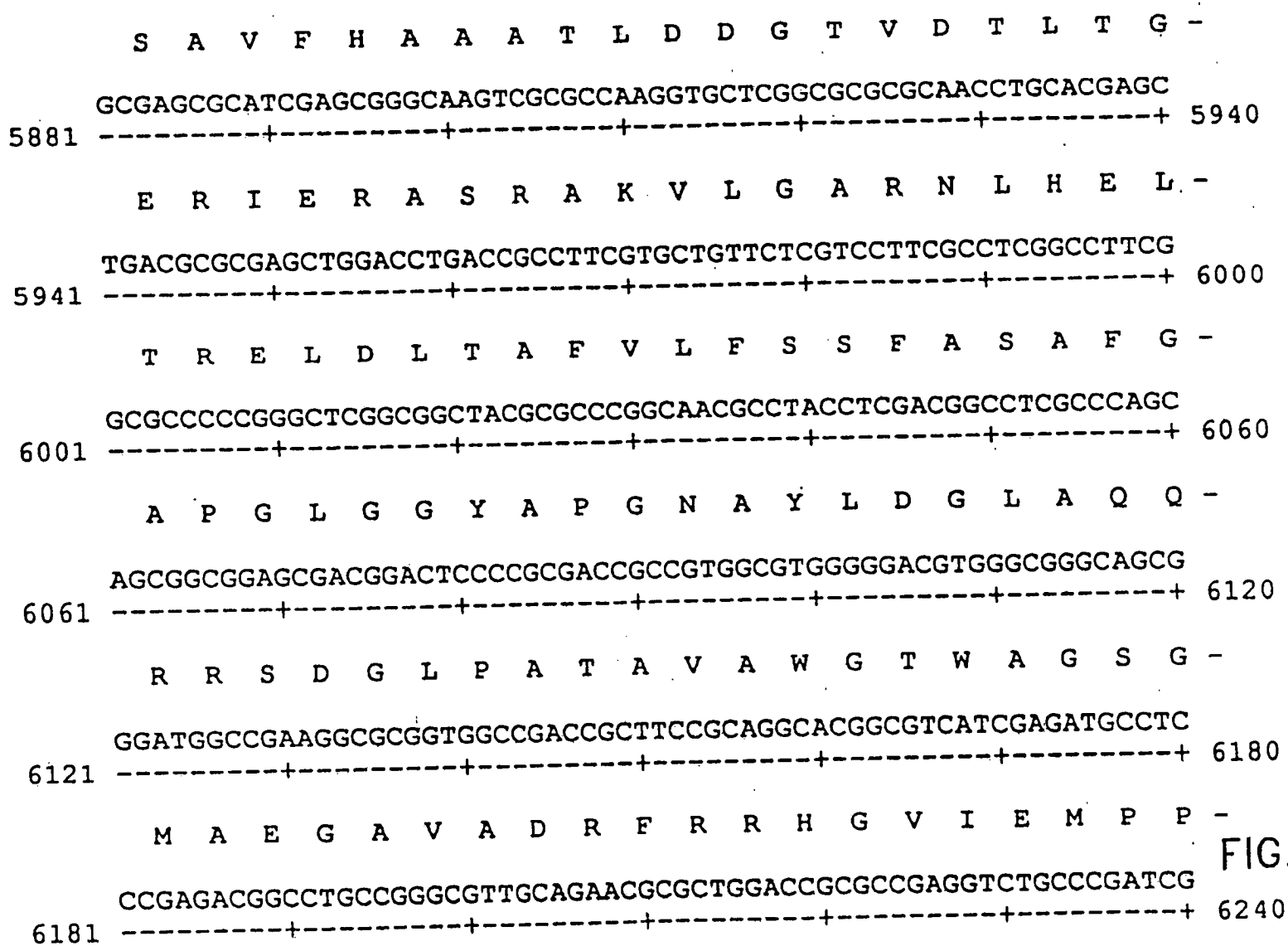

FIG.2-15

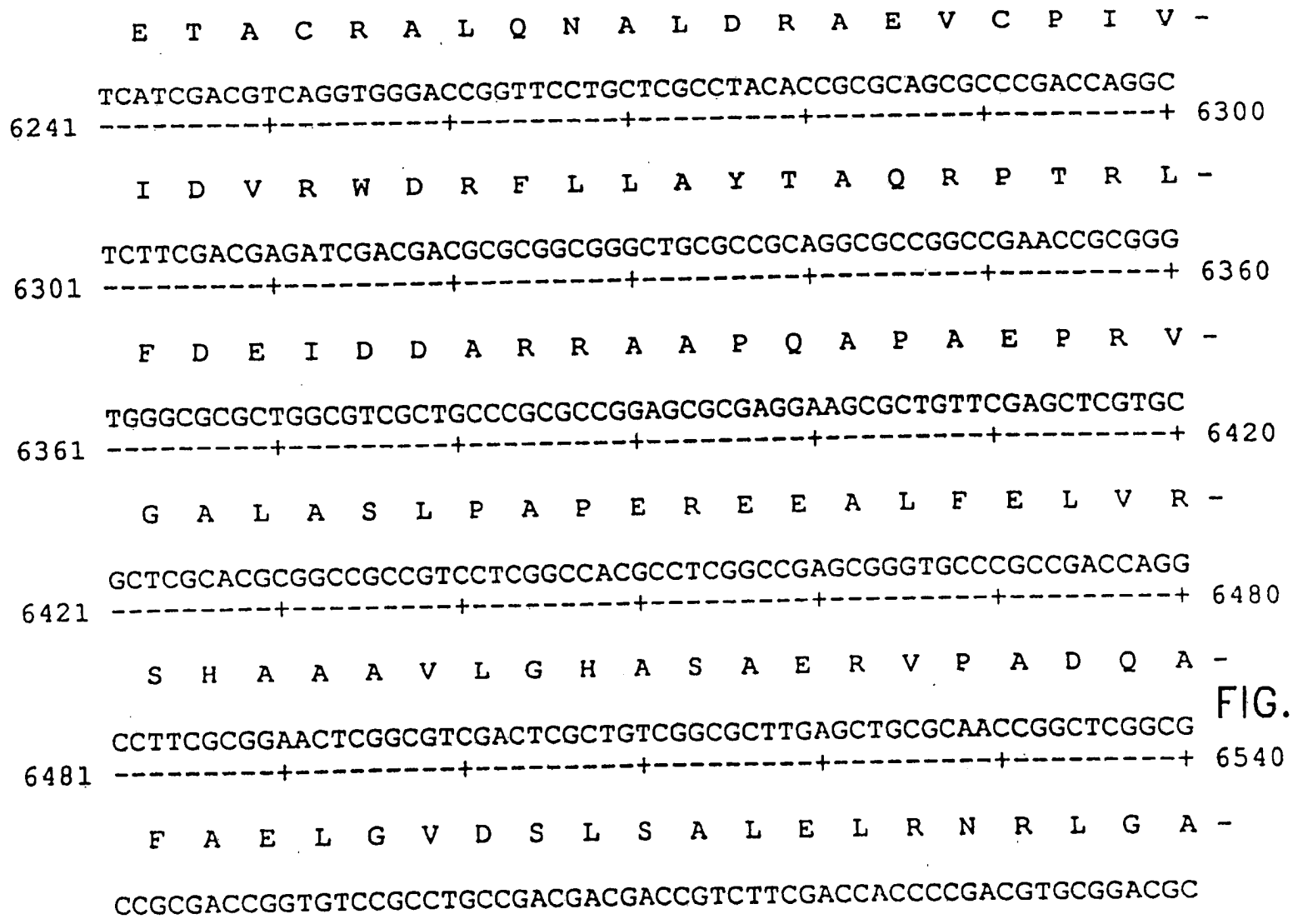
E T A C R A L Q N A L D R A E V C P I V -
TCATCGACGTCAGGTGGGACCGGTTCCTGCTCGCCTACACCGCGCAGCGCCCGACCAGGC
6241 ---------+---------+---------+---------+---------+---------+ 6300
I D V R W D R F L L A Y T A Q R P T R L -
TCTTCGACGAGATCGACGACGCGCGGCGGGCTGCGCCGCAGGCGCCGGCCGAACCGCGGG
6301 ---------+---------+---------+---------+---------+---------+ 6360
F D E I D D A R R A A P Q A P A E P R V -
TGGGCGCGCTGGCGTCGCTGCCCGCGCCGGAGCGCGAGGAAGCGCTGTTCGAGCTCGTGC
6361 ---------+---------+---------+---------+---------+---------+ 6420
G A L A S L P A P E R E E A L F E L V R -
GCTCGCACGCGGCCGCCGTCCTCGGCCACGCCTCGGCCGAGCGGGTGCCCGCCGACCAGG
6421 ---------+---------+---------+---------+---------+---------+ 6480
S H A A A V L G H A S A E R V P A D Q A -
CCTTCGCGGAACTCGGCGTCGACTCGCTGTCGGCGCTTGAGCTGCGCAACCGGCTCGGCG
6481 ---------+---------+---------+---------+---------+---------+ 6540
F A E L G V D S L S A L E L R N R L G A -
CCGCGACCGGTGTCCGCCTGCCGACGACGACCGTCTTCGACCACCCCGACGTGCGGACGC

FIG.2-16

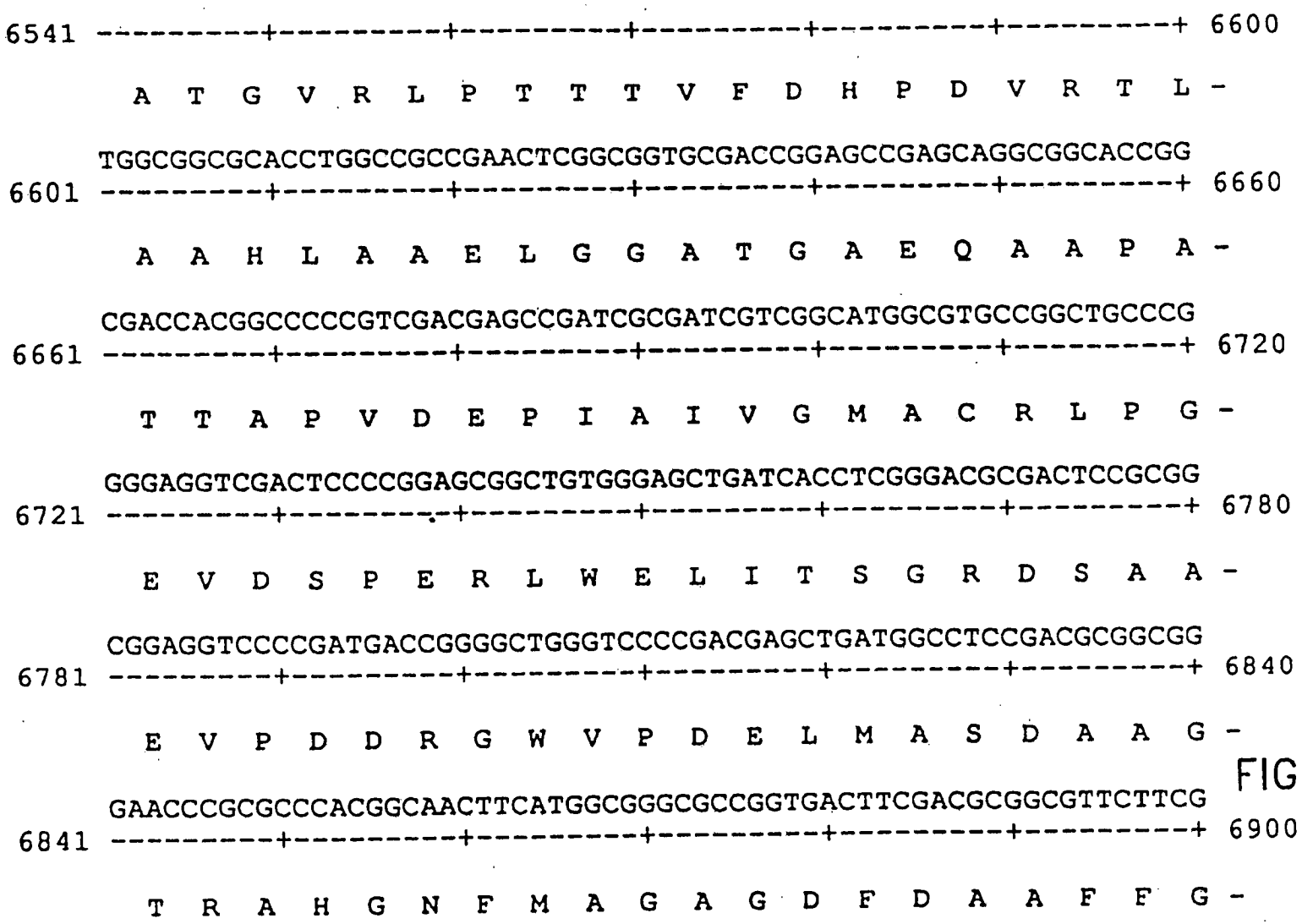

FIG.2—17

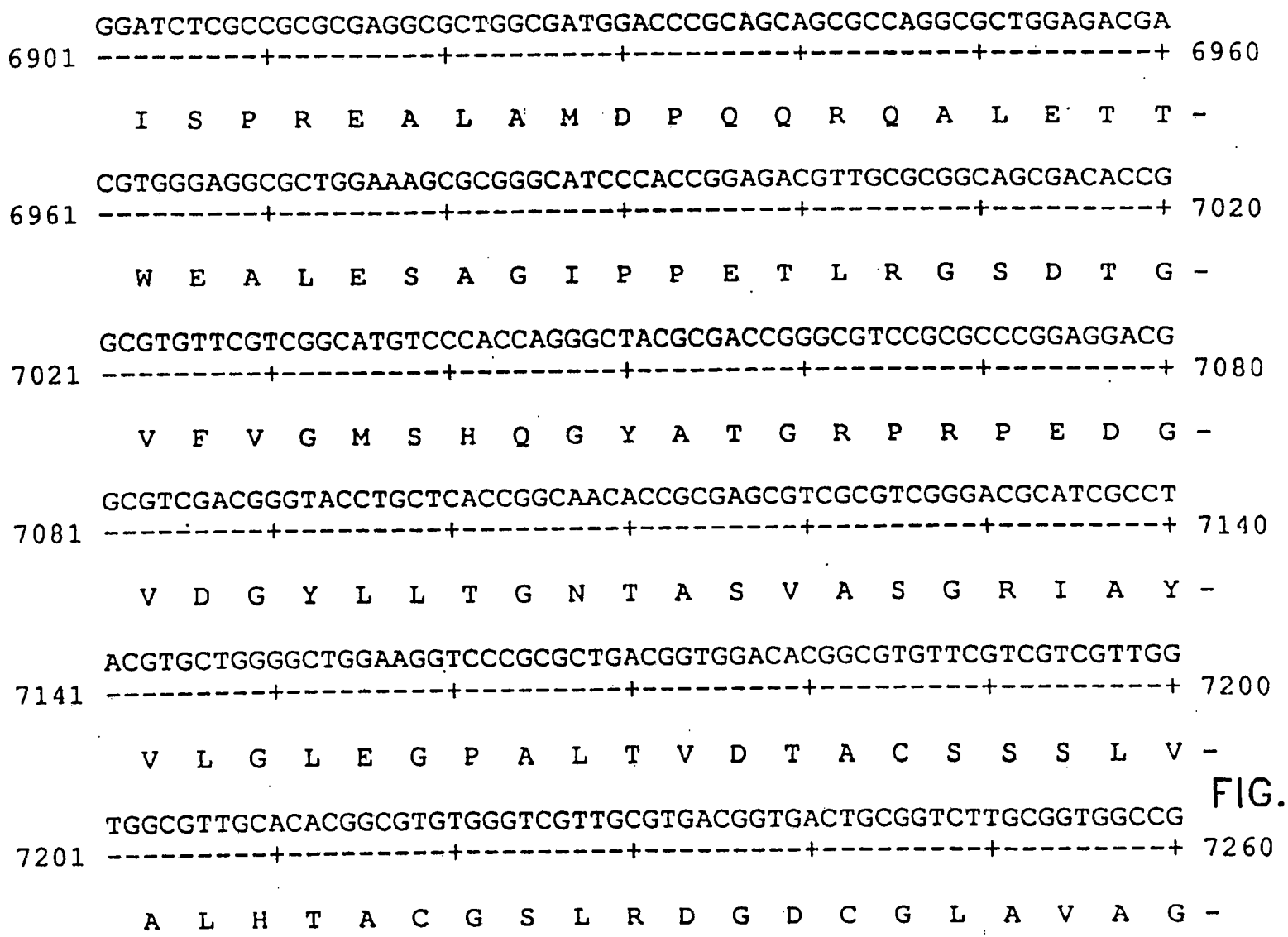
GGATCTCGCCGCGCGAGGCGCTGGCGATGGACCCGCAGCAGCGCCAGGCGCTGGAGACGA
6901 ---------+---------+---------+---------+---------+---------+ 6960
I S P R E A L A M D P Q Q R Q A L E T T -
CGTGGGAGGCGCTGGAAAGCGCGGGCATCCCACCGGAGACGTTGCGCGGCAGCGACACCG
6961 ---------+---------+---------+---------+---------+---------+ 7020
W E A L E S A G I P P E T L R G S D T G -
GCGTGTTCGTCGGCATGTCCCACCAGGGCTACGCGACCGGGCGTCCGCGCCCGGAGGACG
7021 ---------+---------+---------+---------+---------+---------+ 7080
V F V G M S H Q G Y A T G R P R P E D G -
GCGTCGACGGGTACCTGCTCACCGGCAACACCGCGAGCGTCGCGTCGGGACGCATCGCCT
7081 ---------+---------+---------+---------+---------+---------+ 7140
V D G Y L L T G N T A S V A S G R I A Y -
ACGTGCTGGGGCTGGAAGGTCCCGCGCTGACGGTGGACACGGCGTGTTCGTCGTCGTTGG
7141 ---------+---------+---------+---------+---------+---------+ 7200
V L G L E G P A L T V D T A C S S S L V -
TGGCGTTGCACACGGCGTGTGGGTCGTTGCGTGACGGTGACTGCGGTCTTGCGGTGGCCG
7201 ---------+---------+---------+---------+---------+---------+ 7260
A L H T A C G S L R D G D C G L A V A G -

FIG.2—18

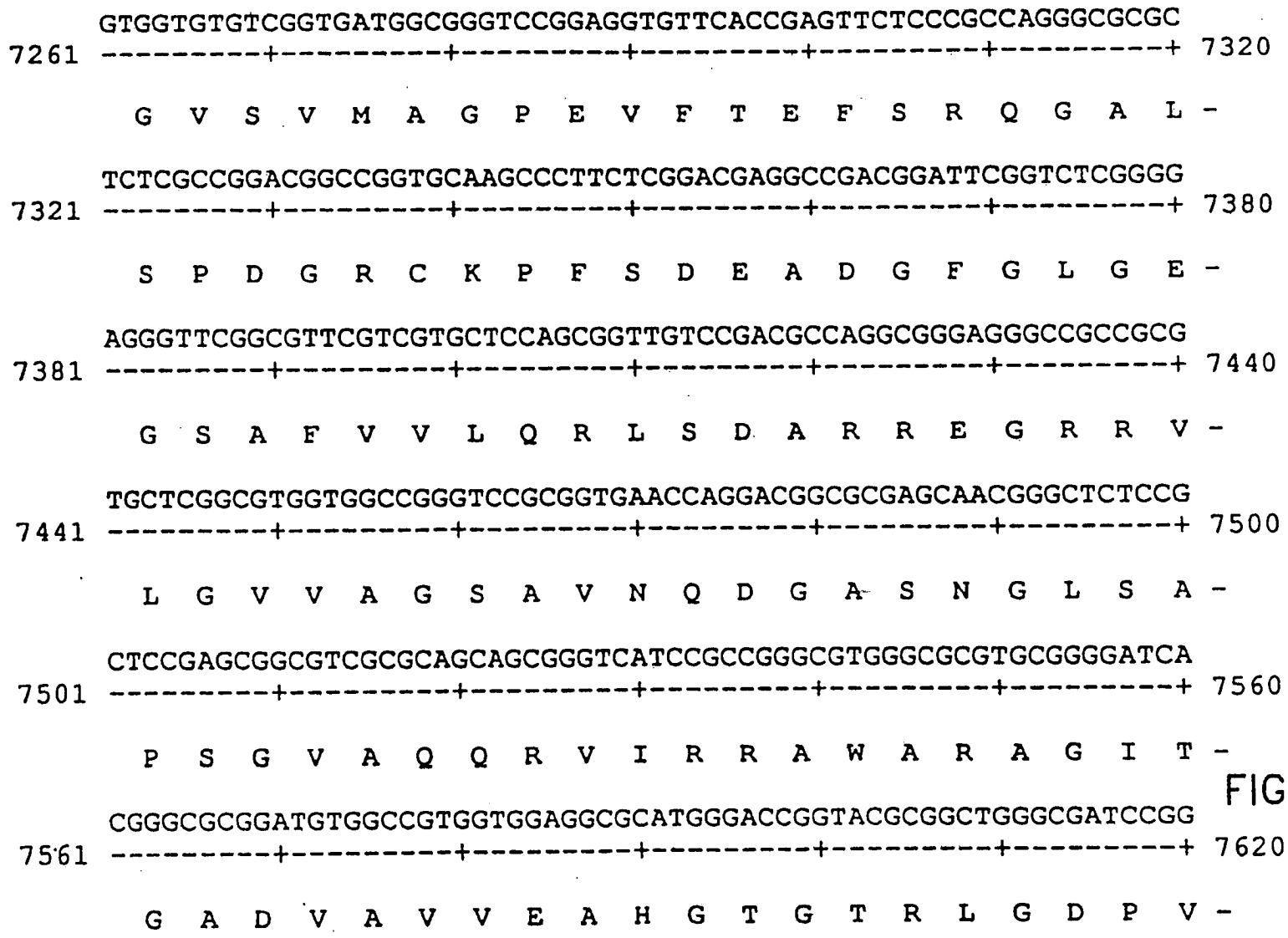

FIG.2-19

```
    TGGAGGCGTCGGCGTTGCTGGCTACTTACGGCAAGTCGCGCGGGTCGTCGGGCCCGGTGC
7621 ---------+---------+---------+---------+---------+---------+ 7680
      E  A  S  A  L  L  A  T  Y  G  K  S  R  G  S  S  G  P  V  L -

    TGCTGGGTTCGGTGAAGTCGAACATCGGTCACGCGCAGGCGGCCGCGGGTGTCGCGGGCG
7681 ---------+---------+---------+---------+---------+---------+ 7740
      L  G  S  V  K  S  N  I  G  H  A  Q  A  A  A  G  V  A  G  V -

    TGATCAAGGTGCTGCTCGGCCTGGAACGCGGTGTGGTGCCCCCGATGCTGTGCCGGGGCG
7741 ---------+---------+---------+---------+---------+---------+ 7800
      I  K  V  L  L  G  L  E  R  G  V  V  P  P  M  L  C  R  G  E -

    AGAGGTCGGGCCTCATCGACTGGTCCTCCGGCGAGATCGAGCTCGCAGACGGCGTGCGGG
7801 ---------+---------+---------+---------+---------+---------+ 7860
      R  S  G  L  I  D  W  S  S  G  E  I  E  L  A  D  G  V  R  E -

    AGTGGTCGCCCGCCGCGGACGGGGTGCGCCGGGCAGGTGTGTCGGCGTTCGGGGTGAGCG
7861 ---------+---------+---------+---------+---------+---------+ 7920
      W  S  P  A  A  D  G  V  R  R  A  G  V  S  A  F  G  V  S  G -

    GGACGAACGCGCACGTGATCATCGCCGAGCCGCCGGAACCGGAGCCCGTGCCGCAACCGC
7921 ---------+---------+---------+---------+---------+---------+ 7980
```

FIG.2-20

```
      T  N  A  H  V  I  I  A  E  P  P  E  P  E  P  V  P  Q  P  R -

    GACGCATGCTGCCCGCGACCGGGGTGGTGCCGGTCGTGCTGTCGGCCAGGACCGGGGCGG
7981 ---------+---------+---------+---------+---------+---------+ 8040

      R  M  L  P  A  T  G  V  V  P  V  V  L  S  A  R  T  G  A  A -

    CGTTGCGGGCGCAGGCCGGCAGGCTCGCCGACCACCTCGCCGCGCATCCCGGGATCGCAC
8041 ---------+---------+---------+---------+---------+---------+ 8100

      L  R  A  Q  A  G  R  L  A  D  H  L  A  A  H  P  G  I  A  P -

    CGGCCGACGTGAGCTGGACGATGGCGCGGGCCCGCCAGCACTTCGAGGAGCGGGCCGCGG
8101 ---------+---------+---------+---------+---------+---------+ 8160

      A  D  V  S  W  T  M  A  R  A  R  Q  H  F  E  E  R  A  A  V -

    TGCTCGCCGCCGACACCGCCGAGGCCGTGCACCGGTTGCGGGCGGTGGCCGACGGCGCGG
8161 ---------+---------+---------+---------+---------+---------+ 8220

      L  A  A  D  T  A  E  A  V  H  R  L  R  A  V  A  D  G  A  V -

    TGGTTCCCGGTGTTGTCACCGGCAGTGCCTCCGACGGTGGTTCAGTGTTCGTCTTCCCTG
8221 ---------+---------+---------+---------+---------+---------+ 8280

      V  P  G  V  V  T  G  S  A  S  D  G  G  S  V  F  V  F  P  G -

    GGCAGGGTGCCCAGTGGGAAGGCATGGCGCGGGAGTTGTTGCCGGTTCCCGTCTTCGCCG
8281 ---------+---------+---------+---------+---------+---------+ 8340
```

FIG.2-21

```
      Q  G  A  Q  W  E  G  M  A  R  E  L  L  P  V  P  V  F  A  E -

    AGTCGATCGCCGAGTGCGATGCGGTGTTGTCGGAGGTGGCCGGATTCTCGGTGTCCGAGG
8341 ---------+---------+---------+---------+---------+---------+ 8400

      S  I  A  E  C  D  A  V  L  S  E  V  A  G  F  S  V  S  E  V -

    TGCTGGAGCCACGTCCGGACGCGCCGTCGCTGGAGCGGGTCGACGTGGTGCAGCCGGTGC
8401 ---------+---------+---------+---------+---------+---------+ 8460

      L  E  P  R  P  D  A  P  S  L  E  R  V  D  V  V  Q  P  V  L -

    TGTTCGCGGTGATGGTGTCGCTGGCGCGGTTGTGGCGTGCCTGCGGTGCCGTTCCTTCGG
8461 ---------+---------+---------+---------+---------+---------+ 8520

      F  A  V  M  V  S  L  A  R  L  W  R  A  C  G  A  V  P  S  A -

    CCGTCATAGGGCACTCGCAGGGTGAGATCGCCGCCGCGGTGGTGGCGGGAGCGTTGTCGC
8521 ---------+---------+---------+---------+---------+---------+ 8580

      V  I  G  H  S  Q  G  E  I  A  A  A  V  V  A  G  A  L  S  L -

    TGGAGGACGGCATGCGCGTCGTCGCCCGCCGGTCGAGGGCGGTGCGTGCGGTCGCGGGCC
8581 ---------+---------+---------+---------+---------+---------+ 8640

      E  D  G  M  R  V  V  A  R  R  S  R  A  V  R  A  V  A  G  R -
```

FIG.2-22

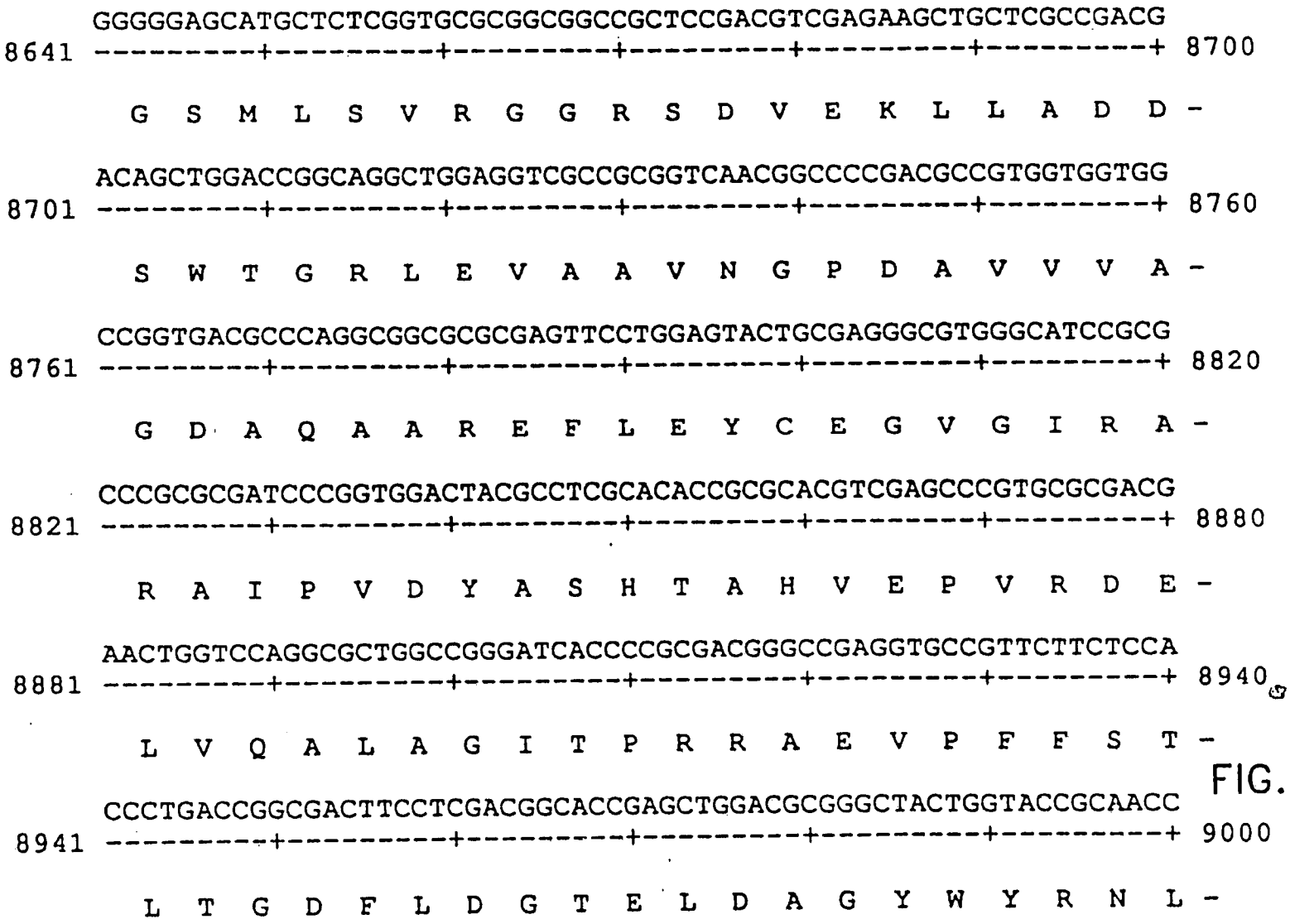
GGGGGAGCATGCTCTCGGTGCGCGGCGGCCGCTCCGACGTCGAGAAGCTGCTCGCCGACG
8641 ---------+---------+---------+---------+---------+---------+ 8700
G S M L S V R G G R S D V E K L L A D D -
ACAGCTGGACCGGCAGGCTGGAGGTCGCCGCGGTCAACGGCCCCGACGCCGTGGTGGTGG
8701 ---------+---------+---------+---------+---------+---------+ 8760
S W T G R L E V A A V N G P D A V V V A -
CCGGTGACGCCCAGGCGGCGCGCGAGTTCCTGGAGTACTGCGAGGGCGTGGGCATCCGCG
8761 ---------+---------+---------+---------+---------+---------+ 8820
G D A Q A A R E F L E Y C E G V G I R A -
CCCGCGCGATCCCGGTGGACTACGCCTCGCACACCGCGCACGTCGAGCCCGTGCGCGACG
8821 ---------+---------+---------+---------+---------+---------+ 8880
R A I P V D Y A S H T A H V E P V R D E -
AACTGGTCCAGGCGCTGGCCGGGATCACCCCGCGACGGGCCGAGGTGCCGTTCTTCTCCA
8881 ---------+---------+---------+---------+---------+---------+ 8940
L V Q A L A G I T P R R A E V P F F S T -
CCCTGACCGGCGACTTCCTCGACGGCACCGAGCTGGACGCGGGCTACTGGTACCGCAACC
8941 ---------+---------+---------+---------+---------+---------+ 9000
L T G D F L D G T E L D A G Y W Y R N L -

FIG.2-23

```
    TGCGTCACCCGGTGGAGTTCCACTCCGCCGTGCAGGCGCTGACCGACCAGGGATACGCGA
9001 ---------+---------+---------+---------+---------+---------+ 9060
      R  H  P  V  E  F  H  S  A  V  Q  A  L  T  D  Q  G  Y  A  T -

    CGTTCATCGAGGTCAGCCCGCACCCGGTGCTGGCGTCGAGCGTCCAGGAGACCCTCGACG
9061 ---------+---------+---------+---------+---------+---------+ 9120
      F  I  E  V  S  P  H  P  V  L  A  S  S  V  Q  E  T  L  D  D -

    ACGCCGAGTCGGACGCGGCGGTGCTCGGGACGCTGGAACGCGACGCGGGCGACGCCGACC
9121 ---------+---------+---------+---------+---------+---------+ 9180
      A  E  S  D  A  A  V  L  G  T  L  E  R  D  A  G  D  A  D  R -

    GCTTCCTCACGGCACTCGCCGACGCGCACACGCGCGGTGTCGCGGTCGACTGGGAAGCGG
9181 ---------+---------+---------+---------+---------+---------+ 9240
      F  L  T  A  L  A  D  A  H  T  R  G  V  A  V  D  W  E  A  V -

    TGCTCGGCCGGGCCGGACTGGTCGACCTGCCGGGTTATCCTTTCCAGGGCAAGCGGTTCT
9241 ---------+---------+---------+---------+---------+---------+ 9300
      L  G  R  A  G  L  V  D  L  P  G  Y  P  F  Q  G  K  R  F  W -

    GGCTGCTGCCGGACCGCACCACCCCTCGTGACGAGCTCGACGGCTGGTTCTACCGGGTCG
9301 ---------+---------+---------+---------+---------+---------+ 9360
```

FIG.2-24

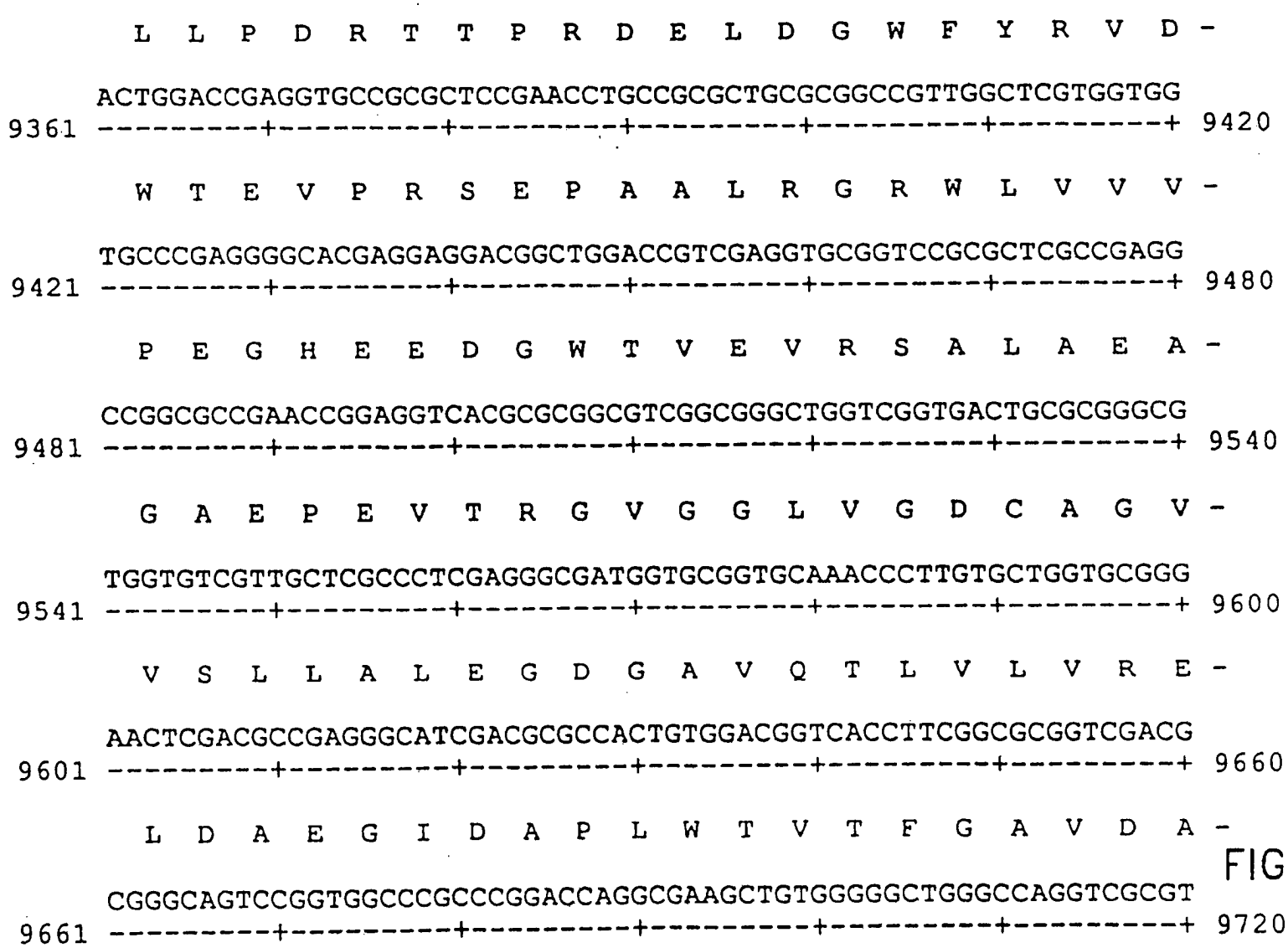

FIG.2-25

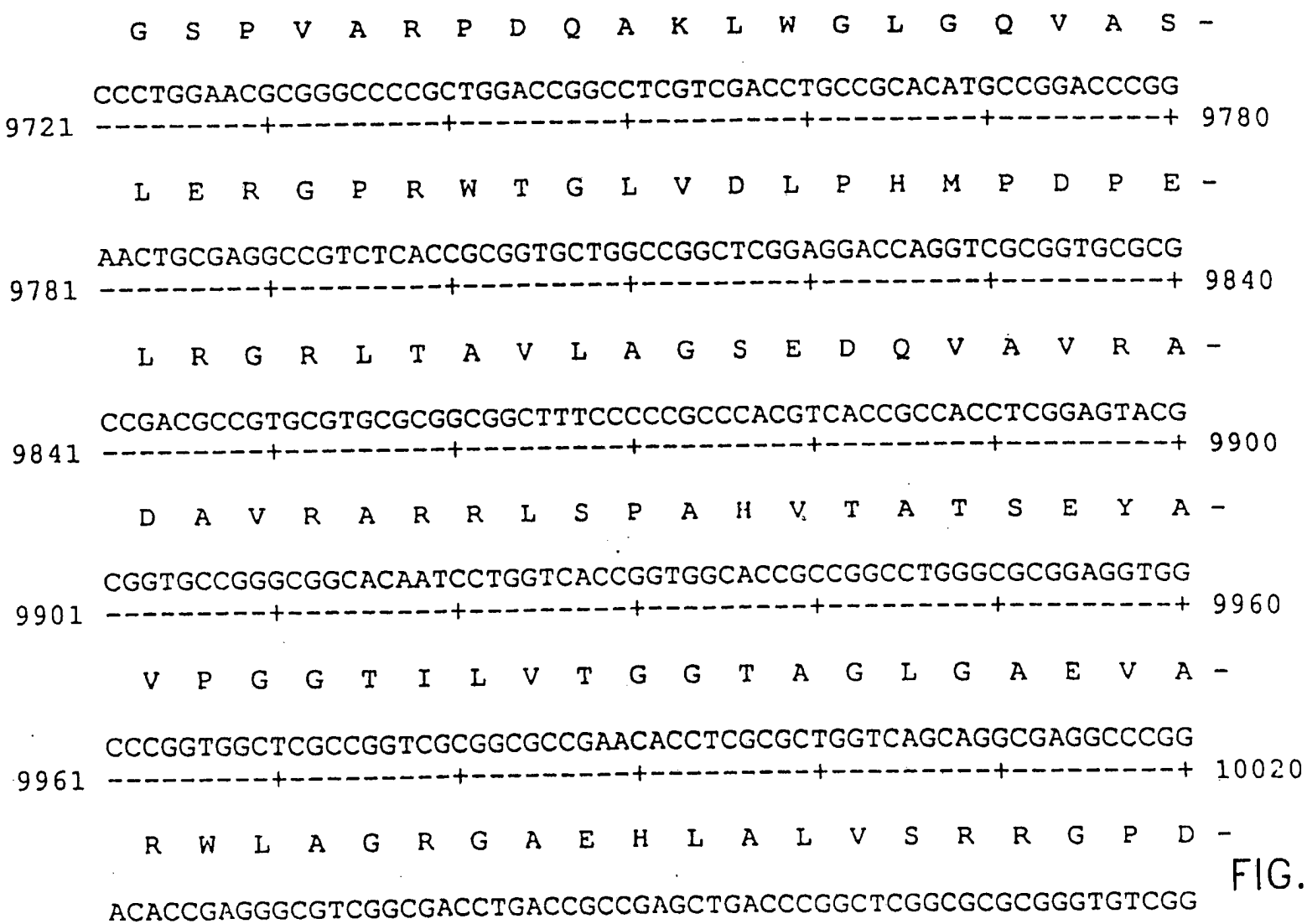
G S P V A R P D Q A K L W G L G Q V A S -
CCCTGGAACGCGGGCCCCGCTGGACCGGCCTCGTCGACCTGCCGCACATGCCGGACCCGG
9721 ---------+---------+---------+---------+---------+---------+ 9780
L E R G P R W T G L V D L P H M P D P E -
AACTGCGAGGCCGTCTCACCGCGGTGCTGGCCGGCTCGGAGGACCAGGTCGCGGTGCGCG
9781 ---------+---------+---------+---------+---------+---------+ 9840
L R G R L T A V L A G S E D Q V A V R A -
CCGACGCCGTGCGTGCGCGGCGGCTTTCCCCCGCCCACGTCACCGCCACCTCGGAGTACG
9841 ---------+---------+---------+---------+---------+---------+ 9900
D A V R A R R L S P A H V T A T S E Y A -
CGGTGCCGGGCGGCACAATCCTGGTCACCGGTGGCACCGCCGGCCTGGGCGCGGAGGTGG
9901 ---------+---------+---------+---------+---------+---------+ 9960
V P G G T I L V T G G T A G L G A E V A -
CCCGGTGGCTCGCCGGTCGCGGCGCCGAACACCTCGCGCTGGTCAGCAGGCGAGGCCCGG
9961 ---------+---------+---------+---------+---------+---------+ 10020
R W L A G R G A E H L A L V S R R G P D -
ACACCGAGGGCGTCGGCGACCTGACCGCCGAGCTGACCCGGCTCGGCGCGCGGGTGTCGG

FIG.2-26

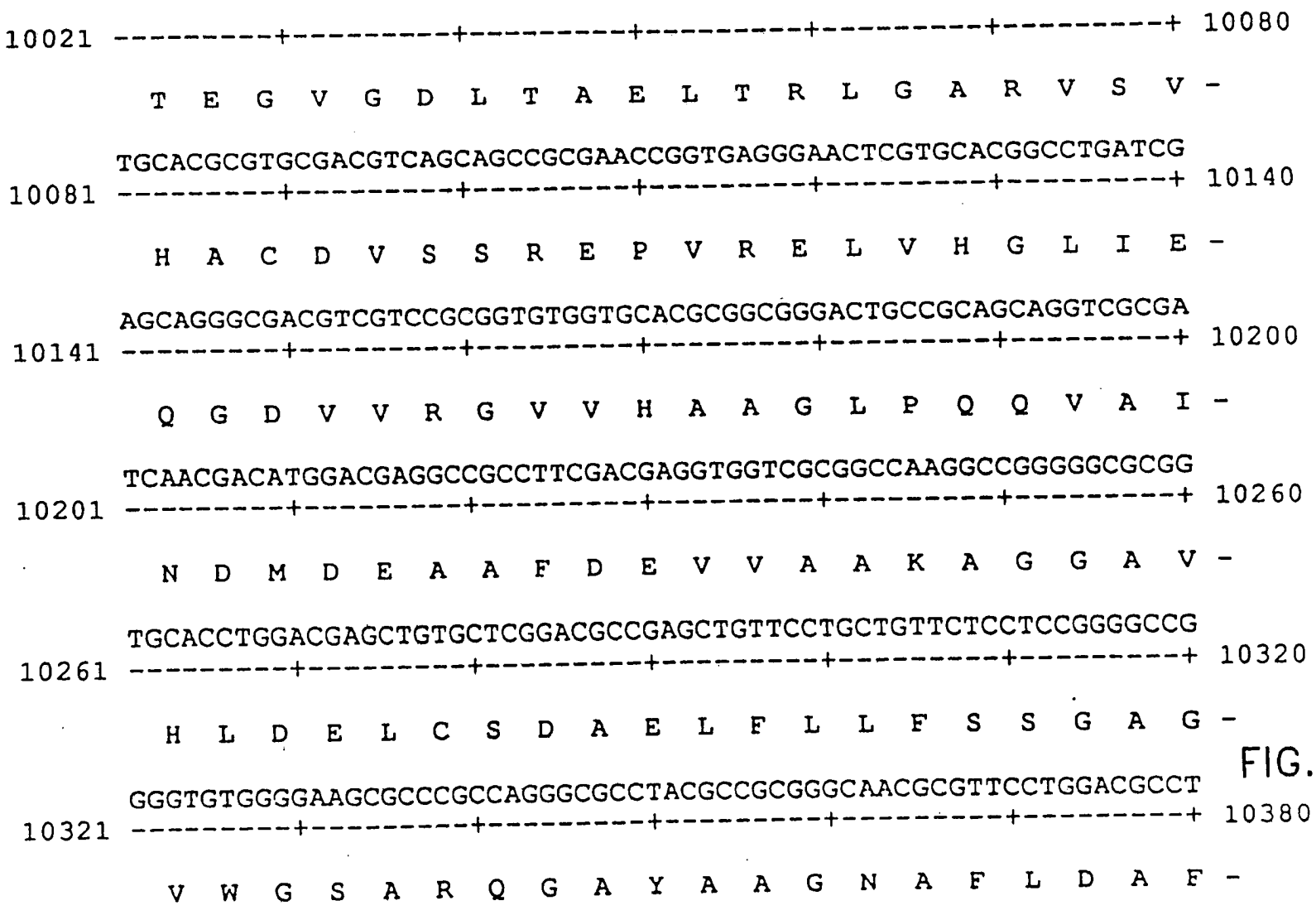

FIG.2-27

SUBSTITUTE SHEET

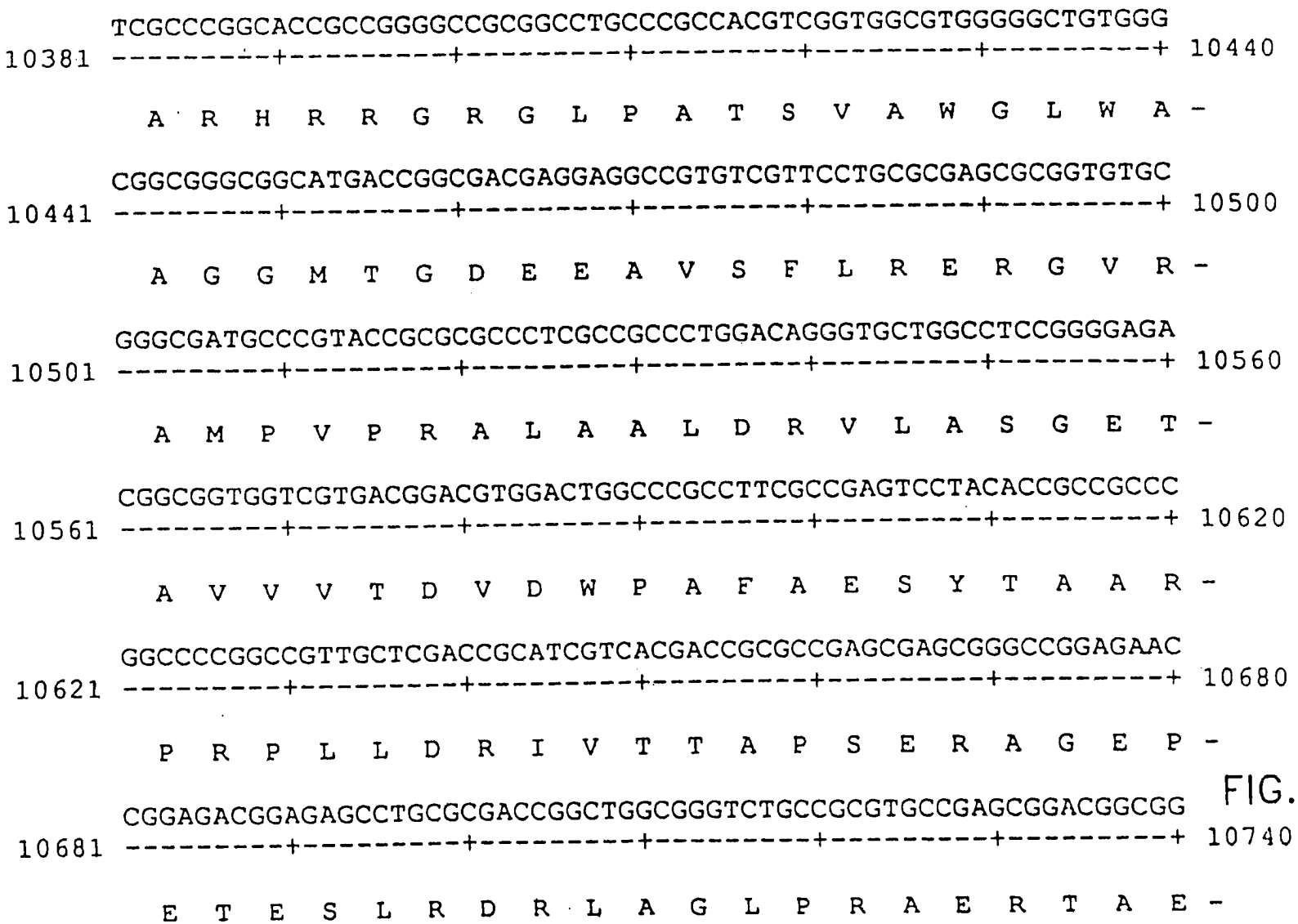

FIG.2-28

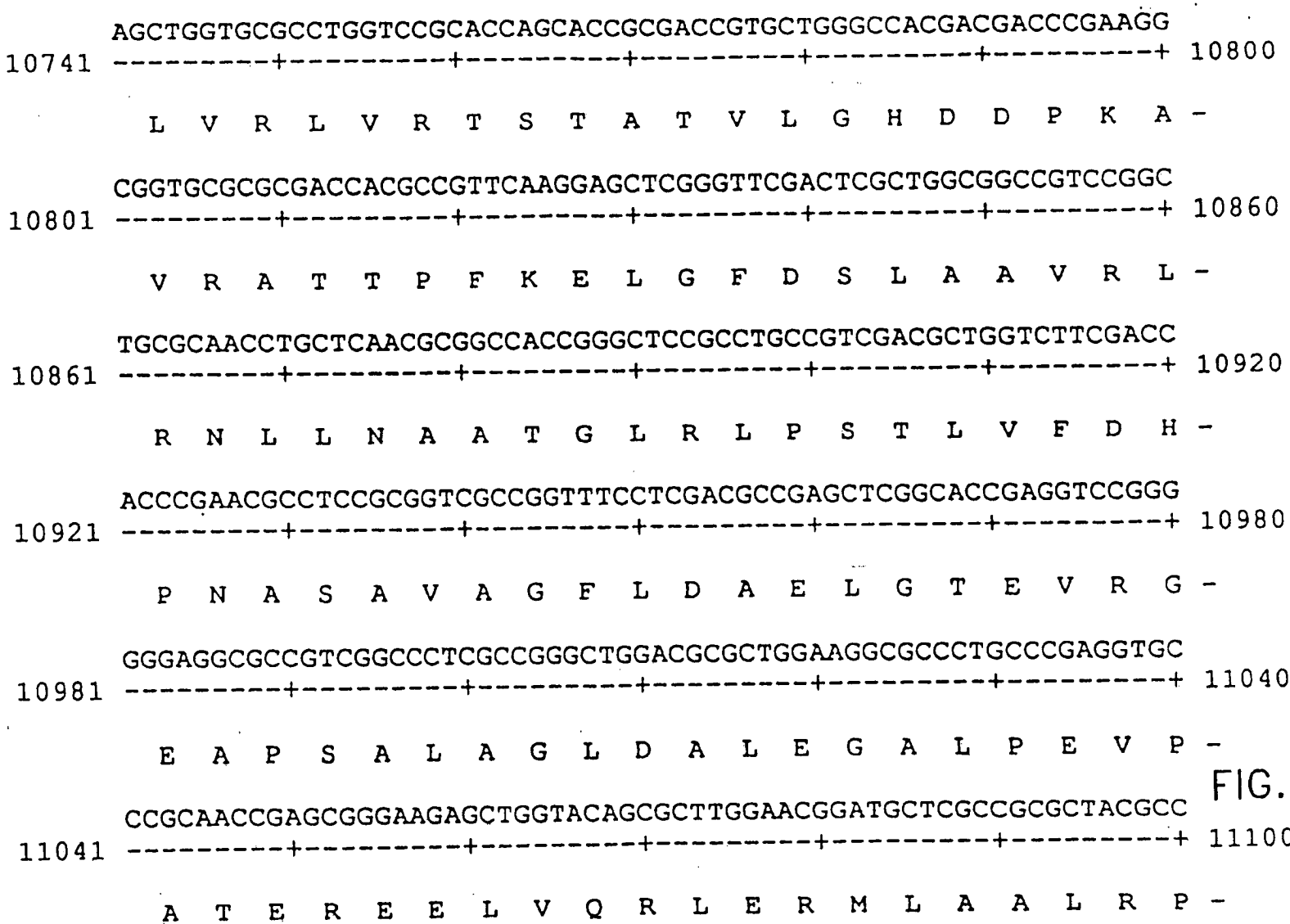

FIG.2-29

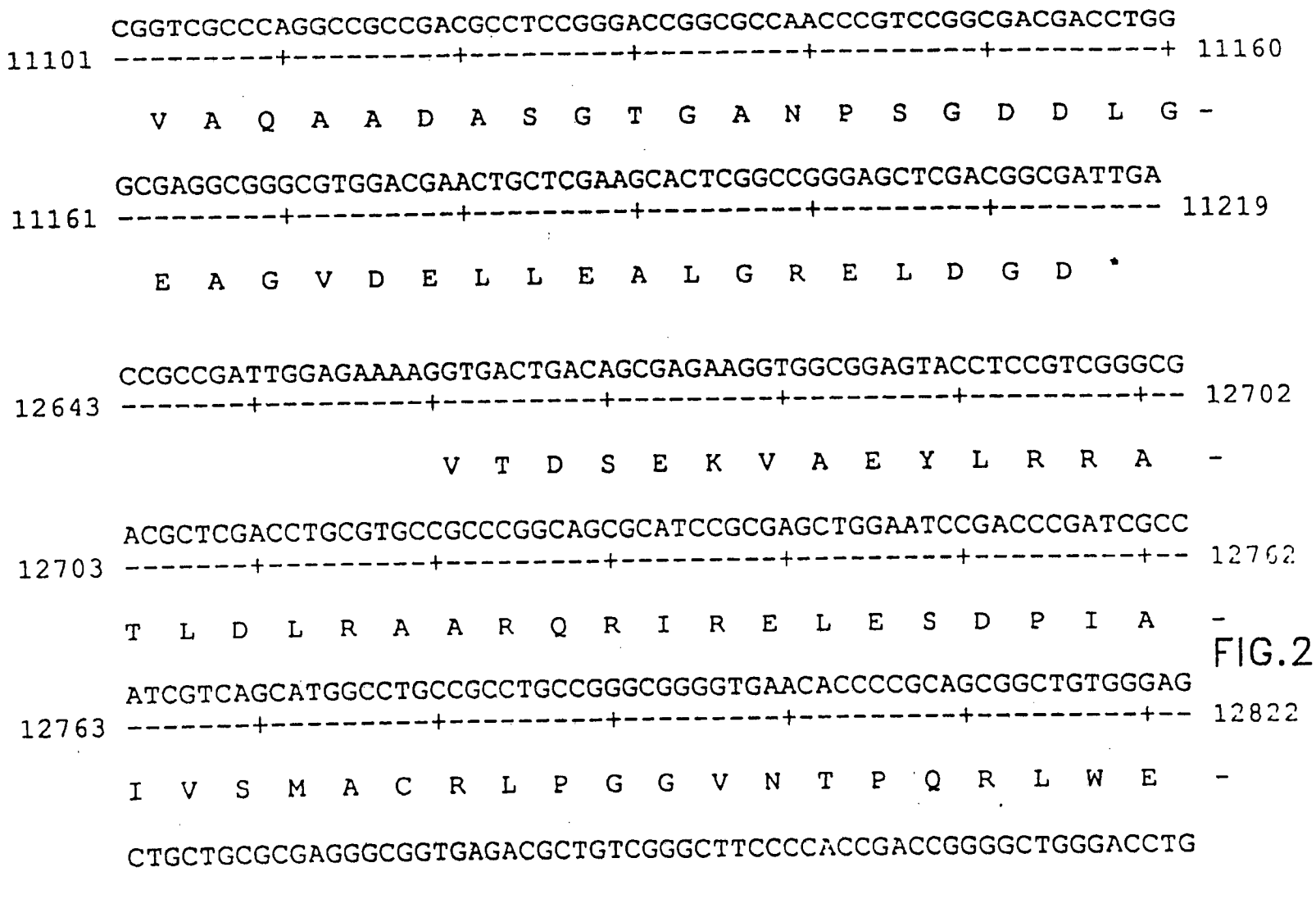

FIG.2-30

SUBSTITUTE SHEET

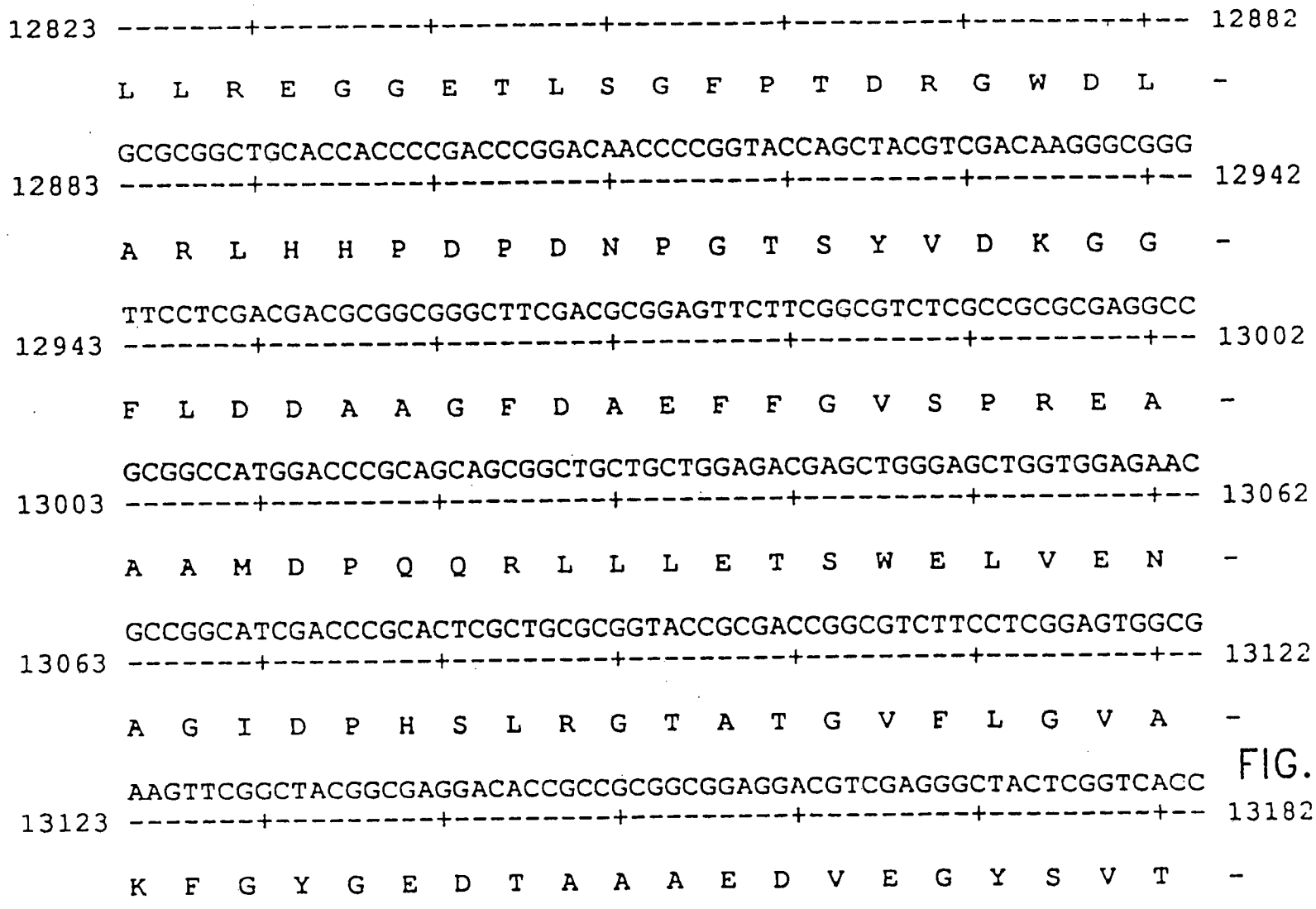

FIG.2-31

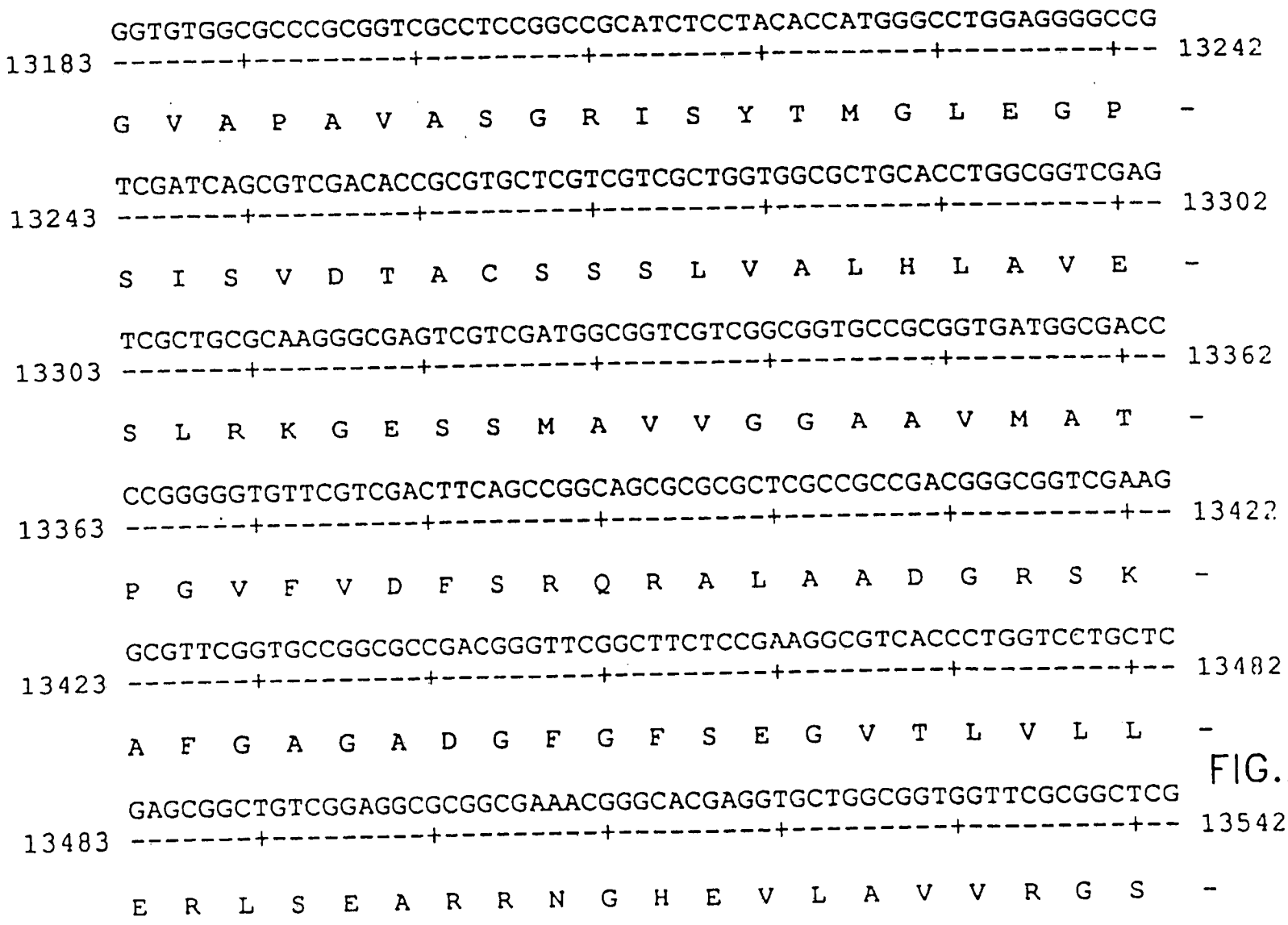
GGTGTGGCGCCCGCGGTCGCCTCCGGCCGCATCTCCTACACCATGGGCCTGGAGGGGCCG
13183 -------+---------+---------+---------+---------+---------+-- 13242
G V A P A V A S G R I S Y T M G L E G P -
TCGATCAGCGTCGACACCGCGTGCTCGTCGTCGCTGGTGGCGCTGCACCTGGCGGTCGAG
13243 -------+---------+---------+---------+---------+---------+-- 13302
S I S V D T A C S S S L V A L H L A V E -
TCGCTGCGCAAGGGCGAGTCGTCGATGGCGGTCGTCGGCGGTGCCGCGGTGATGGCGACC
13303 -------+---------+---------+---------+---------+---------+-- 13362
S L R K G E S S M A V V G G A A V M A T -
CCGGGGGTGTTCGTCGACTTCAGCCGGCAGCGCGCGCTCGCCGCCGACGGGCGGTCGAAG
13363 -------+---------+---------+---------+---------+---------+-- 13422
P G V F V D F S R Q R A L A A D G R S K -
GCGTTCGGTGCCGGCGCCGACGGGTTCGGCTTCTCCGAAGGCGTCACCCTGGTCCTGCTC
13423 -------+---------+---------+---------+---------+---------+-- 13482
A F G A G A D G F G F S E G V T L V L L -
GAGCGGCTGTCGGAGGCGCGGCGAAACGGGCACGAGGTGCTGGCGGTGGTTCGCGGCTCG
13483 -------+---------+---------+---------+---------+---------+-- 13542
E R L S E A R R N G H E V L A V V R G S -

FIG.2-32

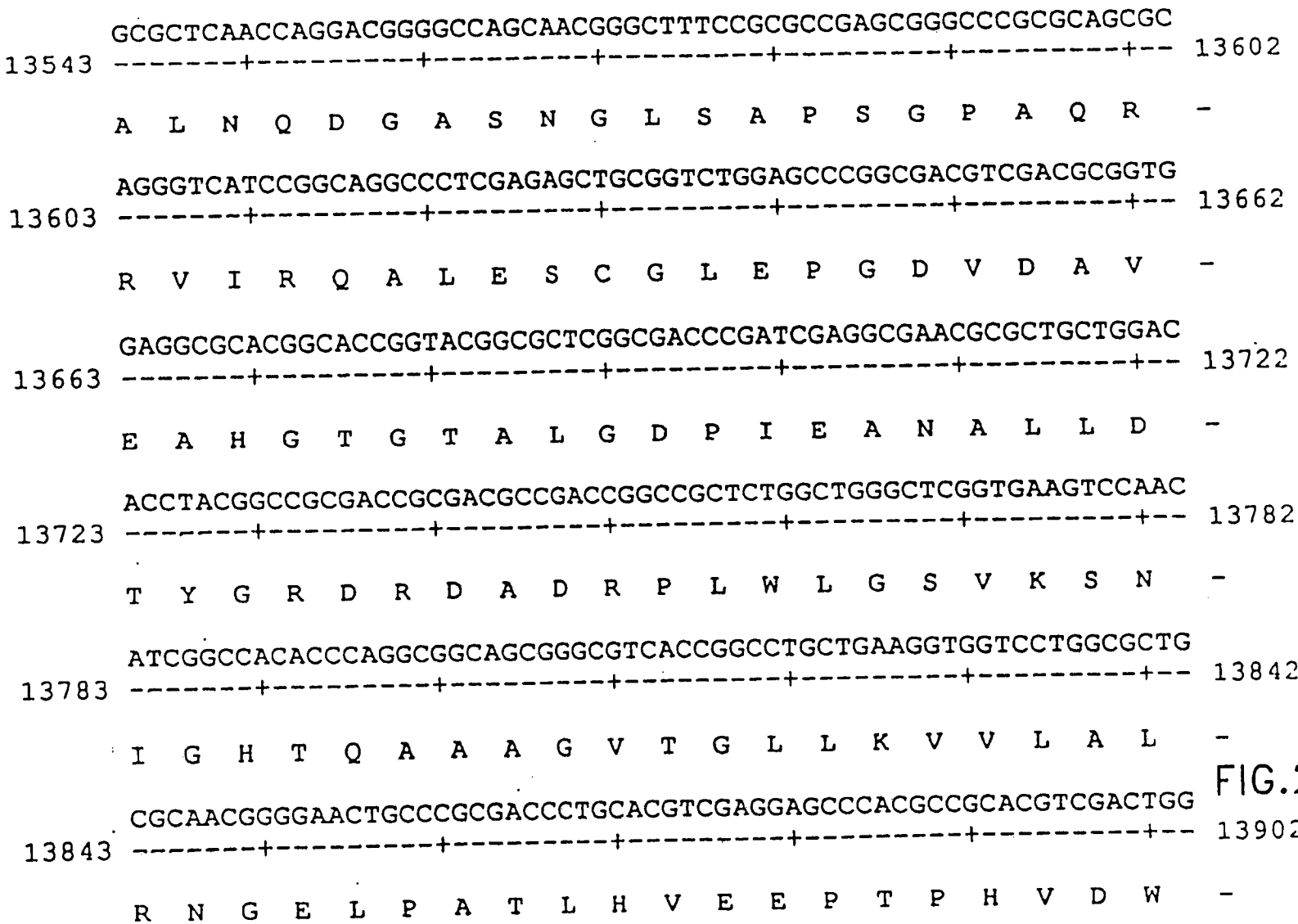

FIG.2-33

```
      TCGTCCGGCGGCGTGGCGCTGCTGGCGGGCAACCAGCCGTGGCGGCGCGGCGAGCGGACT
13903 -------+---------+---------+---------+---------+---------+-- 13962

      S  S  G  G  V  A  L  L  A  G  N  Q  P  W  R  R  G  E  R  T  -

      CGGCGCGCCCGTGTTTCCGCGTTCGGGATCAGCGGGACGAATGCGCACGTGATCGTCGAG
13963 -------+---------+---------+---------+---------+---------+-- 14022

      R  R  A  R  V  S  A  F  G  I  S  G  T  N  A  H  V  I  V  E  -

      GAAGCTCCTGAGCGCGAGCACCGGGAGACCACCGCGCACGACGGCCGACCGGTTCCGCTG
14023 -------+---------+---------+---------+---------+---------+-- 14082

      E  A  P  E  R  E  H  R  E  T  T  A  H  D  G  R  P  V  P  L  -

      GTGGTGTCCGCGCGCACGACGGCGGCGTTGCGGGCGCAGGCCGCCCAGATCGCCGAGCTG
14083 -------+---------+---------+---------+---------+---------+-- 14142

      V  V  S  A  R  T  T  A  A  L  R  A  Q  A  A  Q  I  A  E  L  -

      CTCGAACGCCCGGACGCCGACCTCGCCGGGGTCGGGCTGGGCCTGGCCACGACCCGCGCC
14143 -------+---------+---------+---------+---------+---------+-- 14202

      L  E  R  P  D  A  D  L  A  G  V  G  L  G  L  A  T  T  R  A  -

      CGCCACGAGCACCGCGCCGCCGTGGTGGCATCGACCCGCGAGGAAGCGGTGCGCGGACTG
14203 -------+---------+---------+---------+---------+---------+-- 14262
```

FIG.2-34

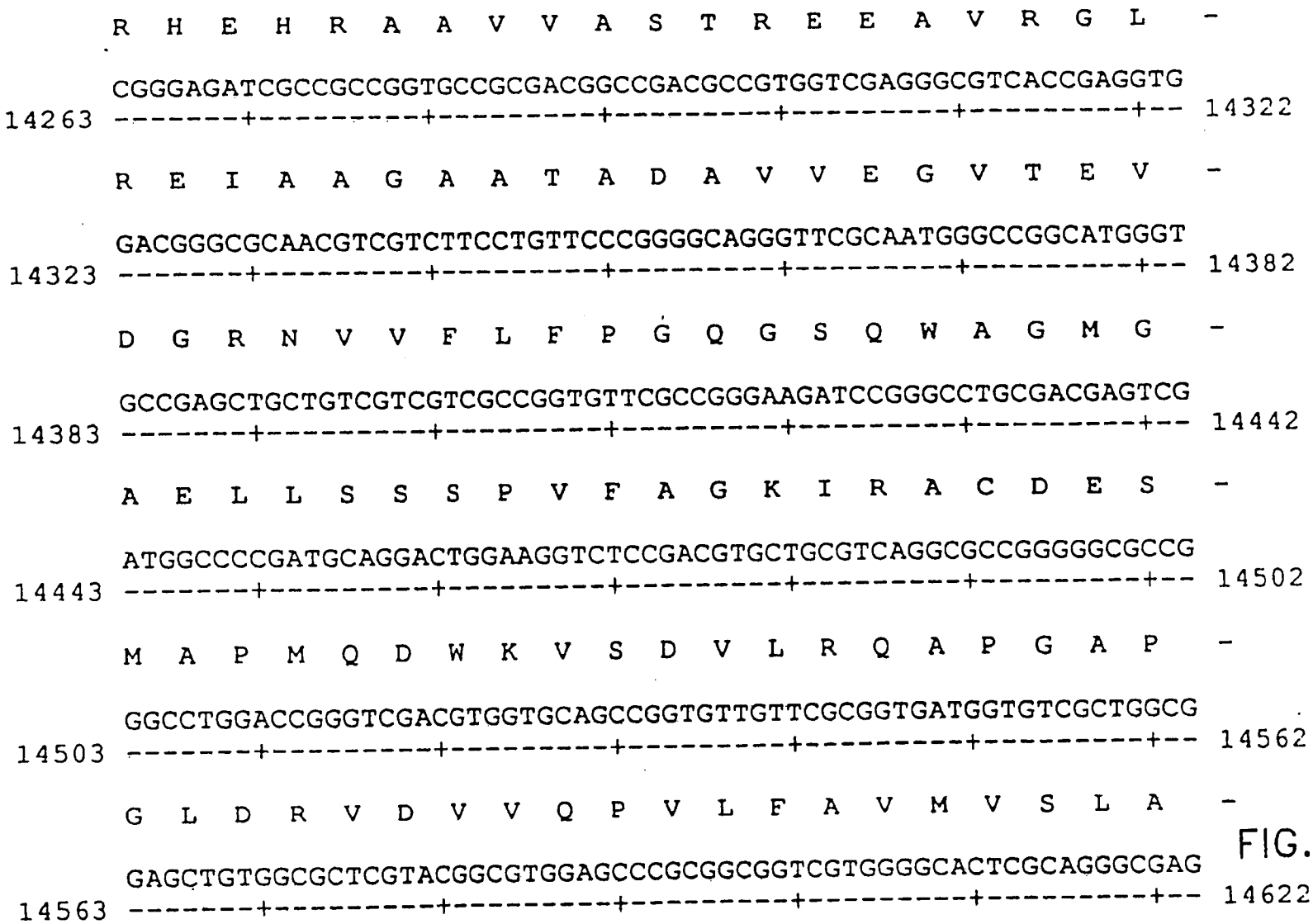

FIG.2-35

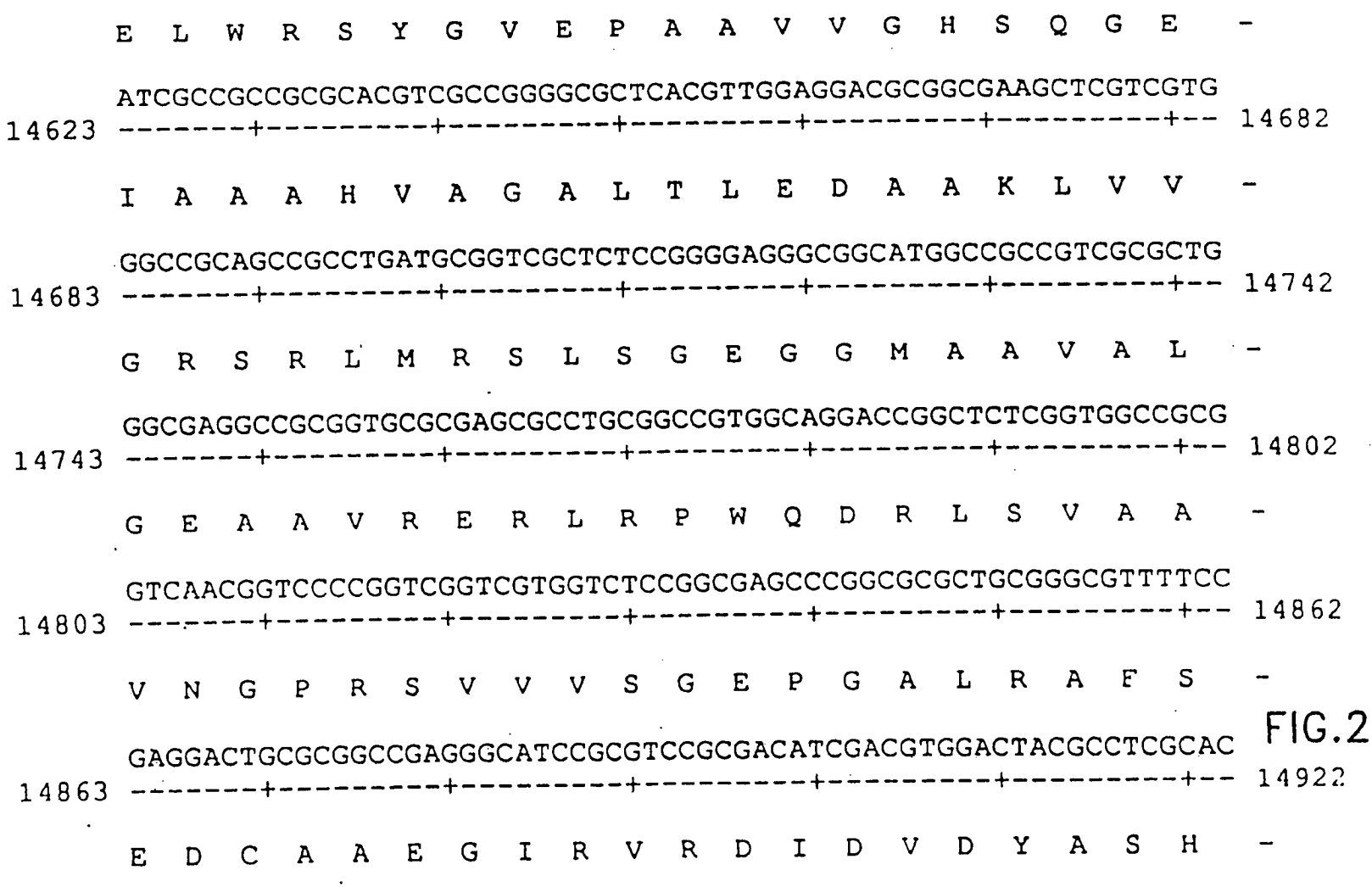

FIG.2-36

SUBSTITUTE SHEET

```
      TCGCCGCAGATCGAGCGGGTCCGCGAGGAACTCCTCGAAACGACCGGCGACATCGCGCCG
14923 -------+---------+---------+---------+---------+---------+-- 14982

      S  P  Q  I  E  R  V  R  E  E  L  L  E  T  T  G  D  I  A  P  -

      CGCCCGGCGCGGGTGACGTTCCACTCCACTGTGGAGTCGCGGTCTATGGACGGCACCGAG
14983 -------+---------+---------+---------+---------+---------+-- 15042

      R  P  A  R  V  T  F  H  S  T  V  E  S  R  S  M  D  G  T  E  -

      CTGGATGCCCGGTACTGGTACCGCAACCTGCGCGAGACGGTGCGCTTCGCCGACGCCGTG
15043 -------+---------+---------+---------+---------+---------+-- 15102

      L  D  A  R  Y  W  Y  R  N  L  R  E  T  V  R  F  A  D  A  V  -

      ACGCGGCTGGCGGAGTCGGGATACGACGCGTTCATCGAGGTCAGCCCGCATCCGGTCGTG
15103 -------+---------+---------+---------+---------+---------+-- 15162

      T  R  L  A  E  S  G  Y  D  A  F  I  E  V  S  P  H  P  V  V  -

      GTCCAGGCCGTCGAGGAGGCGGTCGAAGAGGCTGACGGTGCCGAAGACGCGGTCGTAGTC
15163 -------+---------+---------+---------+---------+---------+-- 15222

      V  Q  A  V  E  E  A  V  E  E  A  D  G  A  E  D  A  V  V  V  -

      GGCTCGCTGCACCGCGACGGCGGTGACCTCTCGGCCTTCCTGCGGTCGATGGCCACCGCG
15223 -------+---------+---------+---------+---------+---------+-- 15282

      G  S  L  H  R  D  G  G  D  L  S  A  F  L  R  S  M  A  T  A  -
```

FIG.2-37

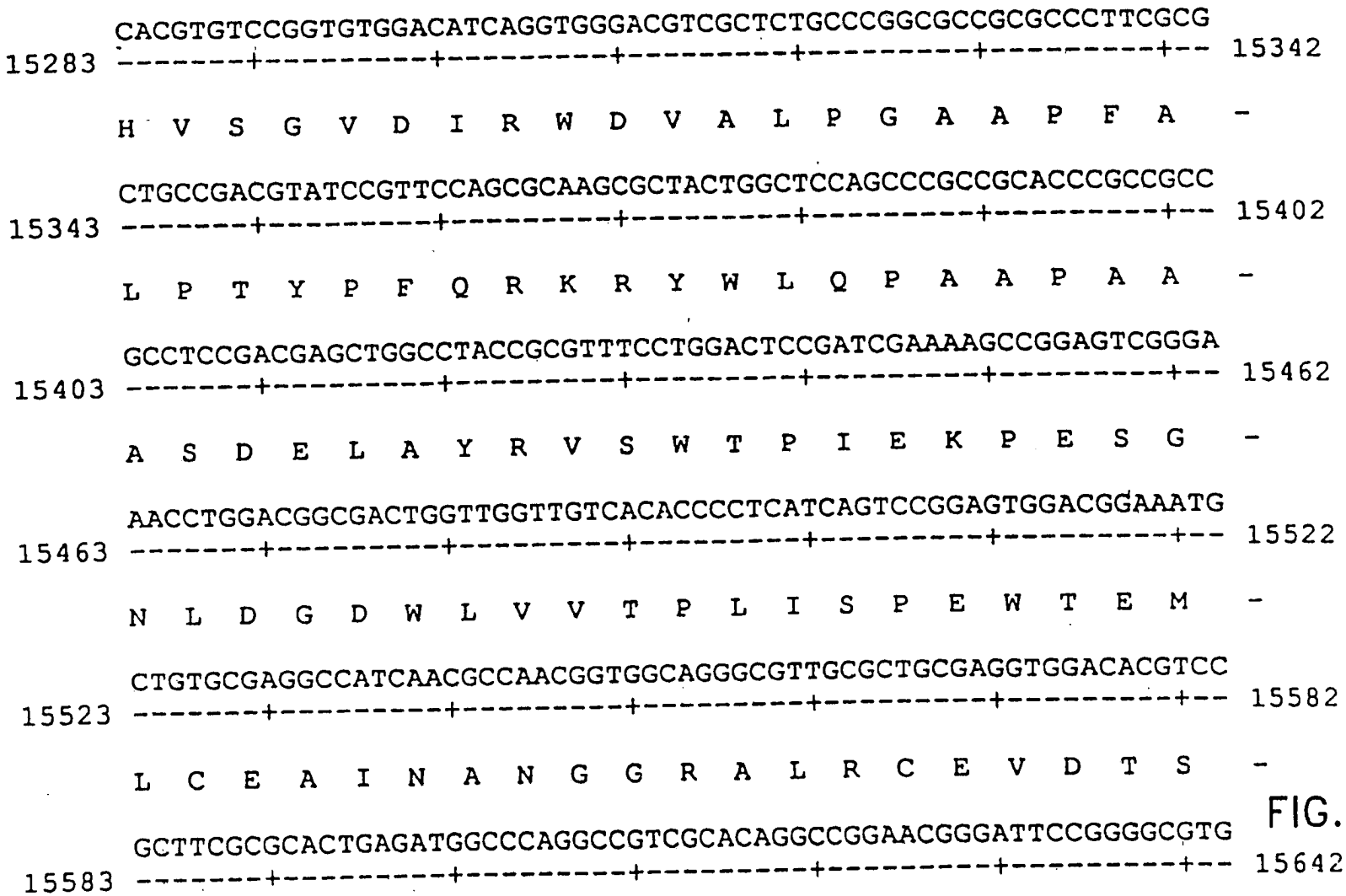

FIG.2-38

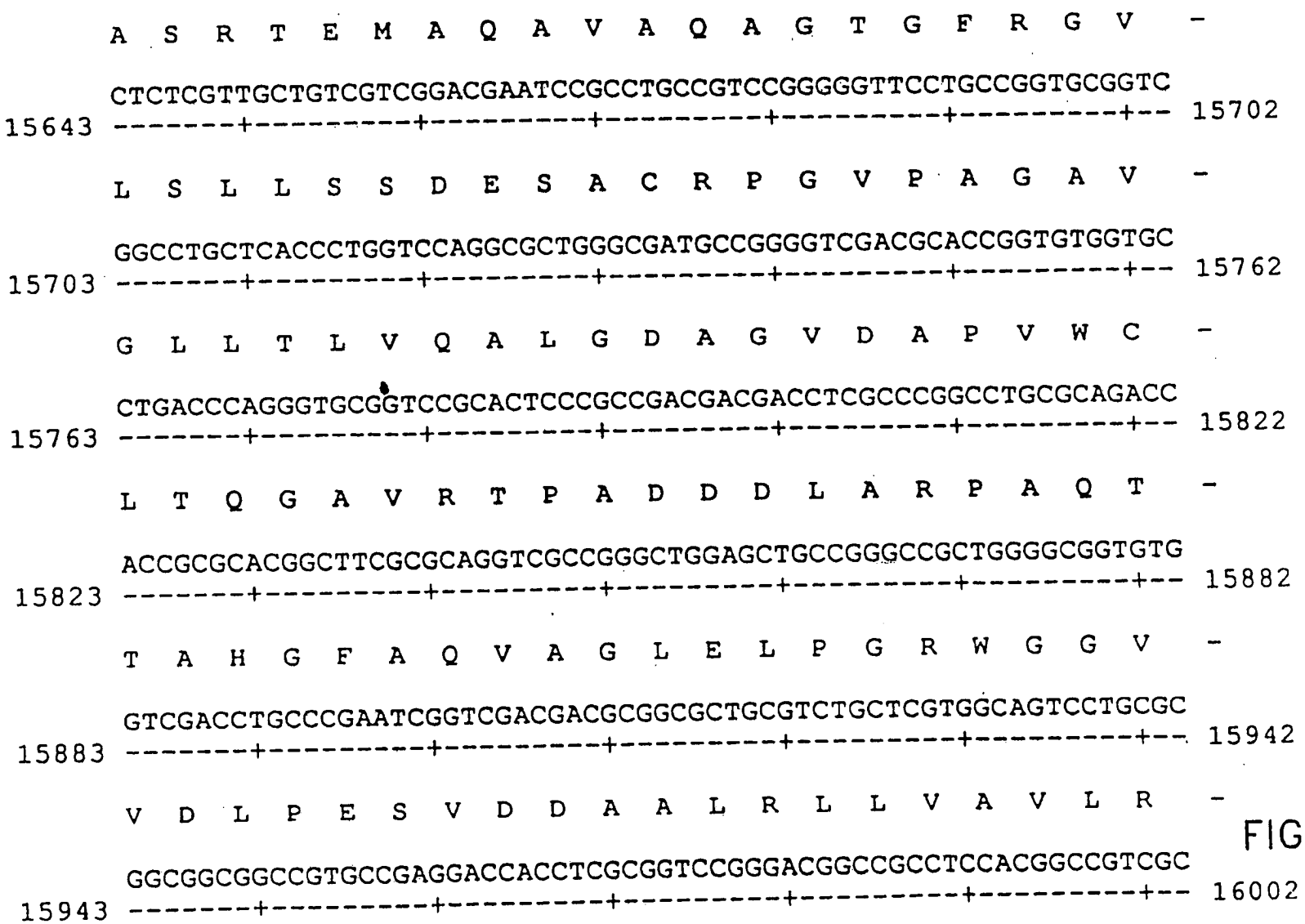
A S R T E M A Q A V A Q A G T G F R G V -
CTCTCGTTGCTGTCGTCGGACGAATCCGCCTGCCGTCCGGGGGGTTCCTGCCGGTGCGGTC
15643 ------+---------+---------+---------+---------+---------+-- 15702
L S L L S S D E S A C R P G V P A G A V -
GGCCTGCTCACCCTGGTCCAGGCGCTGGGCGATGCCGGGGTCGACGCACCGGTGTGGTGC
15703 ------+---------+---------+---------+---------+---------+-- 15762
G L L T L V Q A L G D A G V D A P V W C -
CTGACCCAGGGTGCGGTCCGCACTCCCGCCGACGACGACCTCGCCCGGCCTGCGCAGACC
15763 ------+---------+---------+---------+---------+---------+-- 15822
L T Q G A V R T P A D D D L A R P A Q T -
ACCGCGCACGGCTTCGCGCAGGTCGCCGGGCTGGAGCTGCCGGGCCGCTGGGGCGGTGTG
15823 ------+---------+---------+---------+---------+---------+-- 15882
T A H G F A Q V A G L E L P G R W G G V -
GTCGACCTGCCCGAATCGGTCGACGACGCGGCGCTGCGTCTGCTCGTGGCAGTCCTGCGC
15883 ------+---------+---------+---------+---------+---------+-- 15942
V D L P E S V D D A A L R L L V A V L R -
GGCGGCGGCCGTGCCGAGGACCACCTCGCGGTCCGGGACGGCCGCCTCCACGGCCGTCGC
15943 ------+---------+---------+---------+---------+---------+-- 16002

FIG.2-39

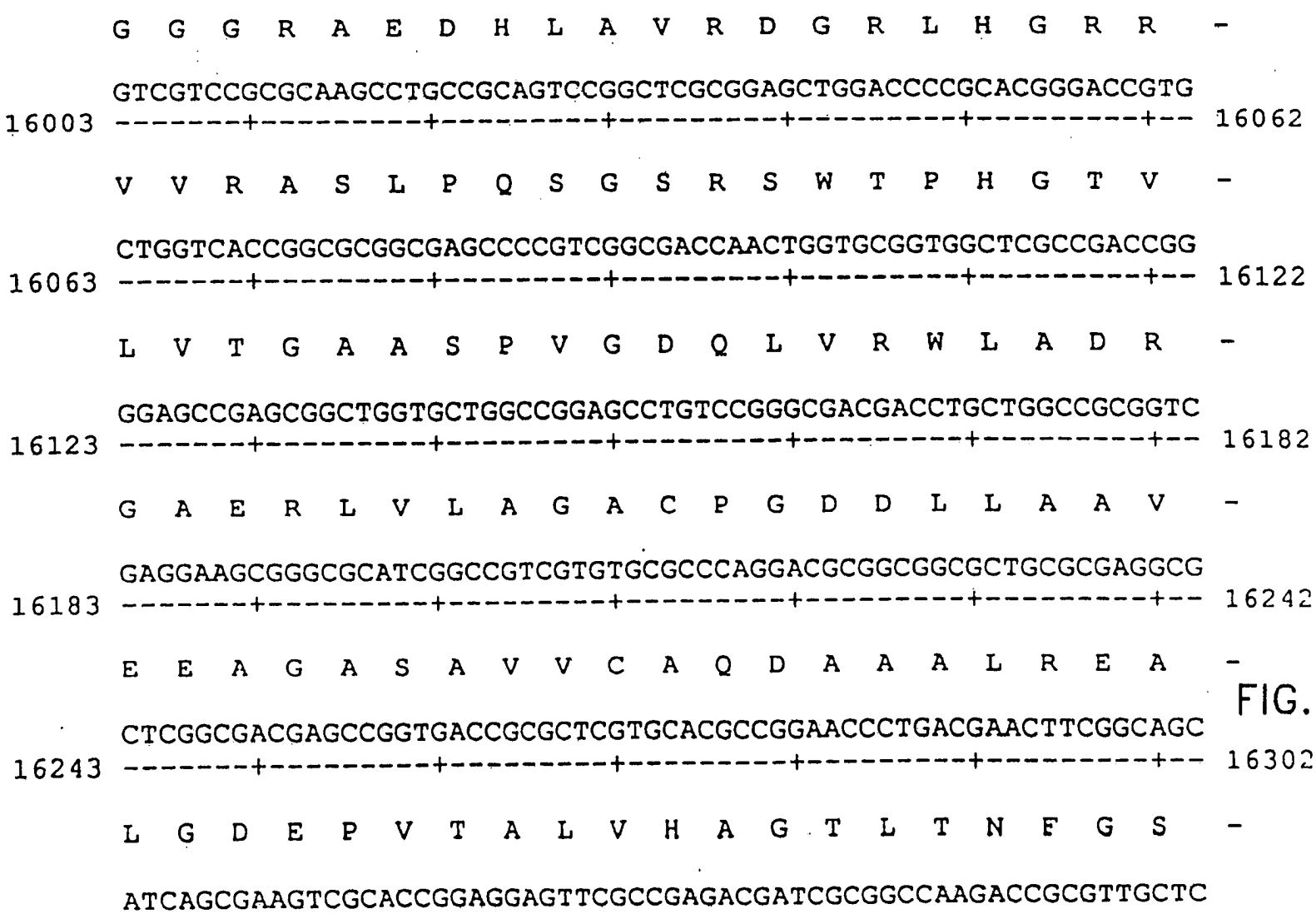

FIG.2-40

SUBSTITUTE SHEET

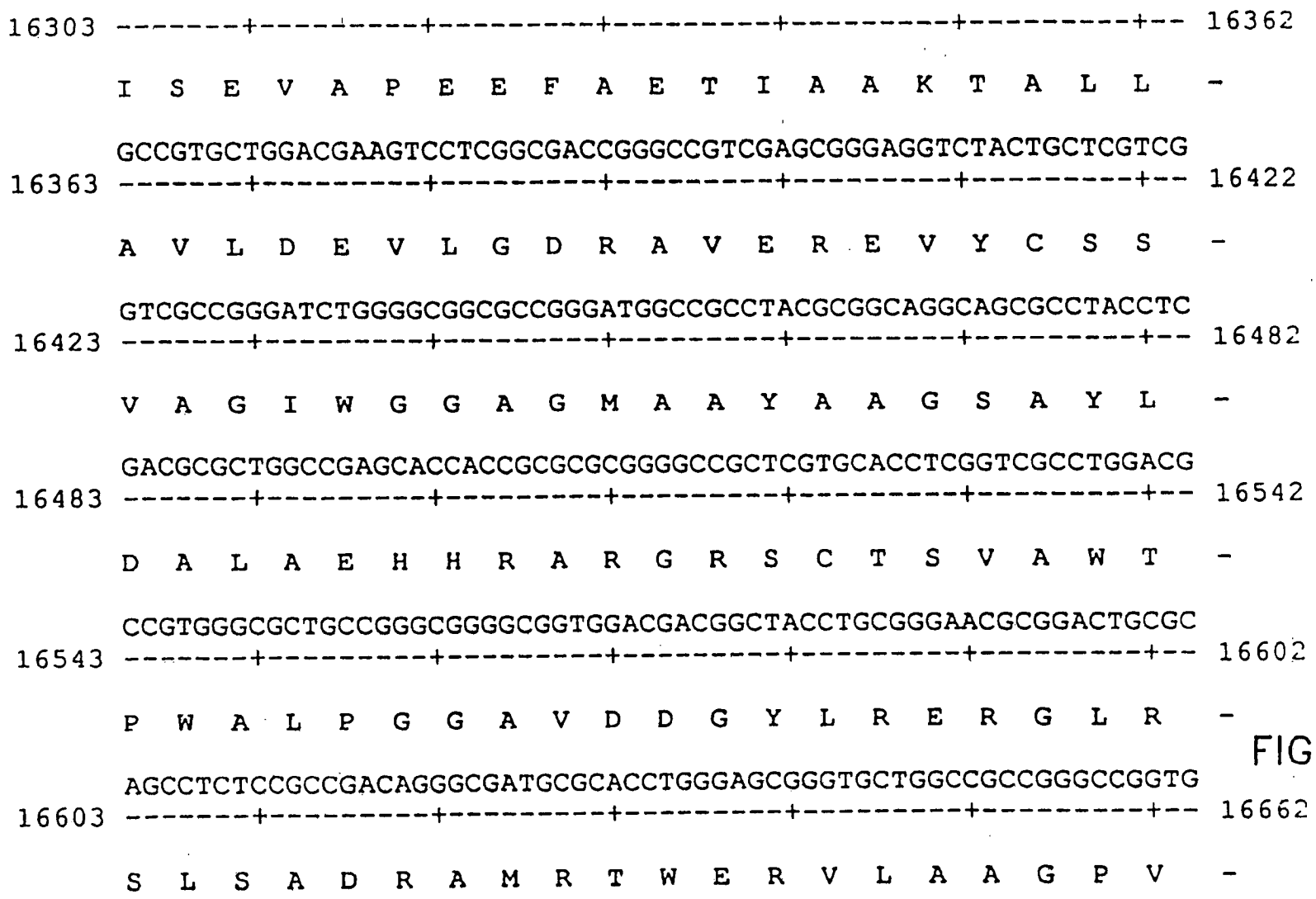
16303 -------+---------+---------+---------+---------+---------+---------+-- 16362
I S E V A P E E F A E T I A A K T A L L -
GCCGTGCTGGACGAAGTCCTCGGCGACCGGGCCGTCGAGCGGGAGGTCTACTGCTCGTCG
16363 -------+---------+---------+---------+---------+---------+---------+-- 16422
A V L D E V L G D R A V E R E V Y C S S -
GTCGCCGGGATCTGGGGCGGCGCCGGGATGGCCGCCTACGCGGCAGGCAGCGCCTACCTC
16423 -------+---------+---------+---------+---------+---------+---------+-- 16482
V A G I W G G A G M A A Y A A G S A Y L -
GACGCGCTGGCCGAGCACCACCGCGCGCGGGGCCGCTCGTGCACCTCGGTCGCCTGGACG
16483 -------+---------+---------+---------+---------+---------+---------+-- 16542
D A L A E H H R A R G R S C T S V A W T -
CCGTGGGCGCTGCCGGGCGGGGCGGTGGACGACGGCTACCTGCGGGAACGCGGACTGCGC
16543 -------+---------+---------+---------+---------+---------+---------+-- 16602
P W A L P G G A V D D G Y L R E R G L R -
AGCCTCTCCGCCGACAGGGCGATGCGCACCTGGGAGCGGGTGCTGGCCGCCGGGCCGGTG
16603 -------+---------+---------+---------+---------+---------+---------+-- 16662
S L S A D R A M R T W E R V L A A G P V -

FIG.2—41

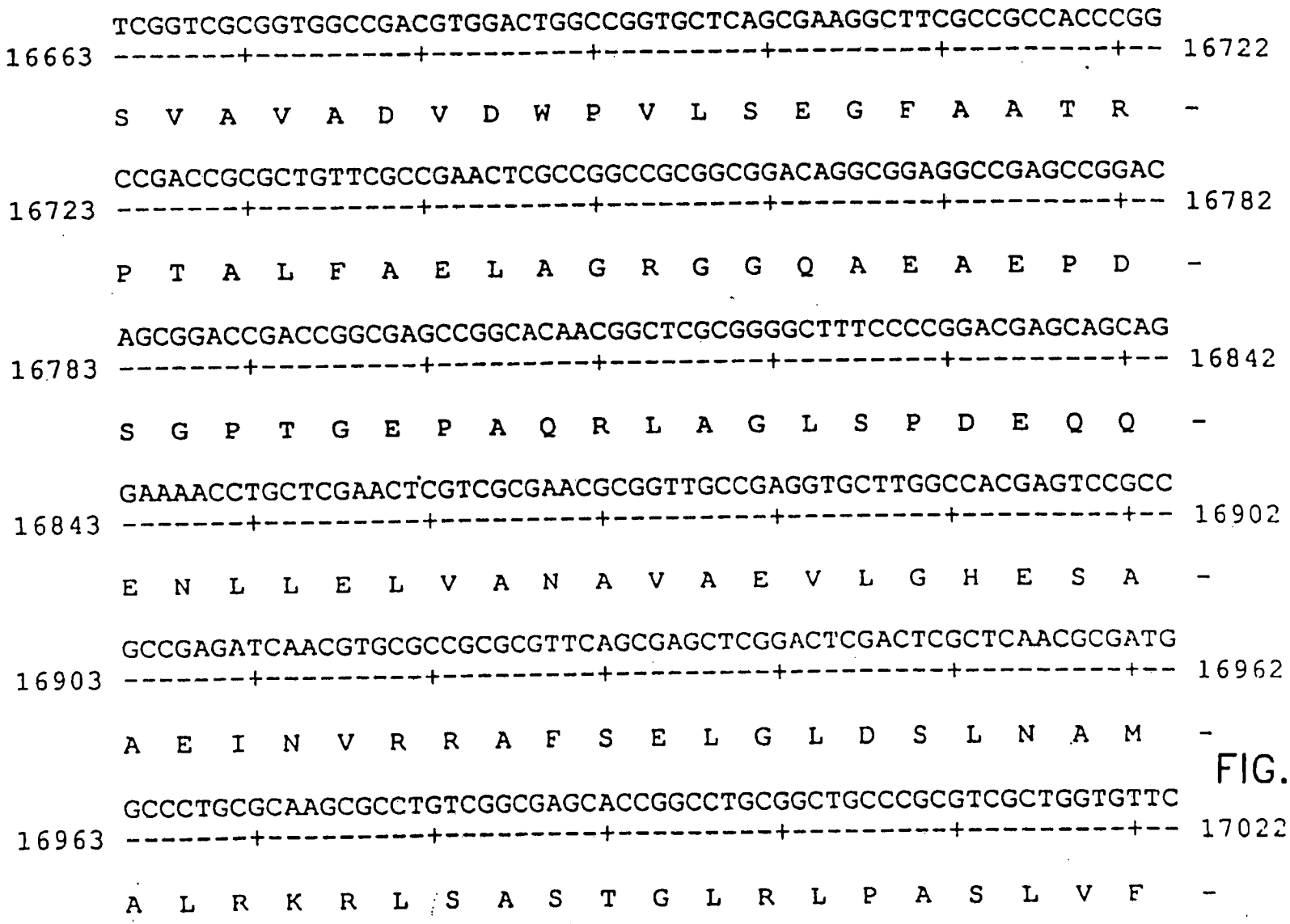

FIG.2-42

```
      GACCACCCCACCGTCACCGCGCTCGCGCAGCACCTGCGCGCCCGGCTCGTCGGTGACGCC
17023 -------+---------+---------+---------+---------+---------+--- 17082

      D  H  P  T  V  T  A  L  A  Q  H  L  R  A  R  L  V  G  D  A   -

      GACCAGGCCGCGGTGCGCGTCGTCGGCGCGGCCGACGAGTCCGAGCCCATCGCCATCGTC
17083 -------+---------+---------+---------+---------+---------+--- 17142

      D  Q  A  A  V  R  V  V  G  A  A  D  E  S  E  P  I  A  I  V   -

      GGCATCGGCTGCCGTTTCCCCGGCGGCATCGGCTCGCCCGAGCAGTTGTGGCGGGTGCTG
17143 -------+---------+---------+---------+---------+---------+--- 17202

      G  I  G  C  R  F  P  G  G  I  G  S  P  E  Q  L  W  R  V  L   -

      GCCGAGGGCGCGAACCTCACCACCGGCTTCCCGGCCGACCGGGGCTGGGACATCGGGCGG
17203 -------+---------+---------+---------+---------+---------+--- 17262

      A  E  G  A  N  L  T  T  G  F  P  A  D  R  G  W  D  I  G  R   -

      CTCTACCACCCGGACCCGGACAACCCCGGCACCAGCTACGTGGACAAGGGCGGGTTCCTC
17263 -------+---------+---------+---------+---------+---------+--- 17322

      L  Y  H  P  D  P  D  N  P  G  T  S  Y  V  D  K  G  G  F  L   -

      ACCGACGCGGCGGATTTCGACCCGGGCTTCTTCGGCATCACGCCCCGCGAAGCGCTGGCG
17323 -------+---------+---------+---------+---------+---------+--- 17382

      T  D  A  A  D  F  D  P  G  F  F  G  I  T  P  R  E  A  L  A   -
```

FIG.2-43

```
      ATGGACCCGCAGCAGCGCCTCATGCTGGAGACGGCGTGGGAGGCAGTGGAACGCGCGGGC
17383 -------+---------+---------+---------+---------+---------+-- 17442

      M  D  P  Q  Q  R  L  M  L  E  T  A  W  E  A  V  E  R  A  G   -

      ATCGACCCCGACGCCCTGCGAGGCACCGACACCGGCGTCTTCGTCGGCATGAACGGCCAG
17443 -------+---------+---------+---------+---------+---------+-- 17502

      I  D  P  D  A  L  R  G  T  D  T  G  V  F  V  G  M  N  G  Q   -

      TCCTACATGCAGCTGCTGGCCGGTGAGGCCGAACGCGTCGACGGCTACCAGGGCCTCGGA
17503 -------+---------+---------+---------+---------+---------+-- 17562

      S  Y  M  Q  L  L  A  G  E  A  E  R  V  D  G  Y  Q  G  L  G   -

      AACTCCGCGAGCGTGCTCTCCGGGCGCATCGCCTACACCTTCGGCTGGGAGGGCCCGGCG
17563 -------+---------+---------+---------+---------+---------+-- 17622

      N  S  A  S  V  L  S  G  R  I  A  Y  T  F  G  W  E  G  P  A   -

      CTGACGGTGGACACCGCGTGCTCGTCCTCGCTGGTCGGCATCCACCTCGCGATGCAGGCG
17623 -------+---------+---------+---------+---------+---------+-- 17682

      L  T  V  D  T  A  C  S  S  S  L  V  G  I  H  L  A  M  Q  A   -

      CTGCGGCGCGGTGAGTGCTCCCTGGCGCTGGCCGGCGGCGTCACGGTCATGTCCGACCCG
17683 -------+---------+---------+---------+---------+---------+-- 17742
```

FIG.2—44

```
      L  R  R  G  E  C  S  L  A  L  A  G  G  V  T  V  M  S  D  P  -

      TACACCTTCGTCGACTTCAGCACGCAGCGCGGGCTCGCCTCCGACGGTCGCTGCAAGGCG
17743 -------+---------+---------+---------+---------+---------+-- 17802

      Y  T  F  V  D  F  S  T  Q  R  G  L  A  S  D  G  R  C  K  A  -

      TTCTCCGCGCGGGCCGACGGCTTCGCGCTGTCGGAAGGCGTCGCCGCGCTGGTGCTGGAG
17803 -------+---------+---------+---------+---------+---------+-- 17862

      F  S  A  R  A  D  G  F  A  L  S  E  G  V  A  A  L  V  L  E  -

      CCGCTTTCCCGGGCGCGCGCCAACGGGCACCAGGTGCTGGCCGTGCTGCGCGGCAGCGCG
17863 -------+---------+---------+---------+---------+---------+-- 17922

      P  L  S  R  A  R  A  N  G  H  Q  V  L  A  V  L  R  G  S  A  -

      GTCAACCAGGACGGTGCCAGCAACGGTCTCGCCGCTCCCAACGGCCCGTCGCAGGAGCGG
17923 -------+---------+---------+---------+---------+---------+-- 17982

      V  N  Q  D  G  A  S  N  G  L  A  A  P  N  G  P  S  Q  E  R  -

      GTGATCCGGCAGGCGCTCGCCGCTTCGGGCGTGCCGGCCGCGGACGTCGACGTCGTGGAG
17983 -------+---------+---------+---------+---------+---------+-- 18042

      V  I  R  Q  A  L  A  A  S  G  V  P  A  A  D  V  D  V  V  E  -
```

FIG.2-45

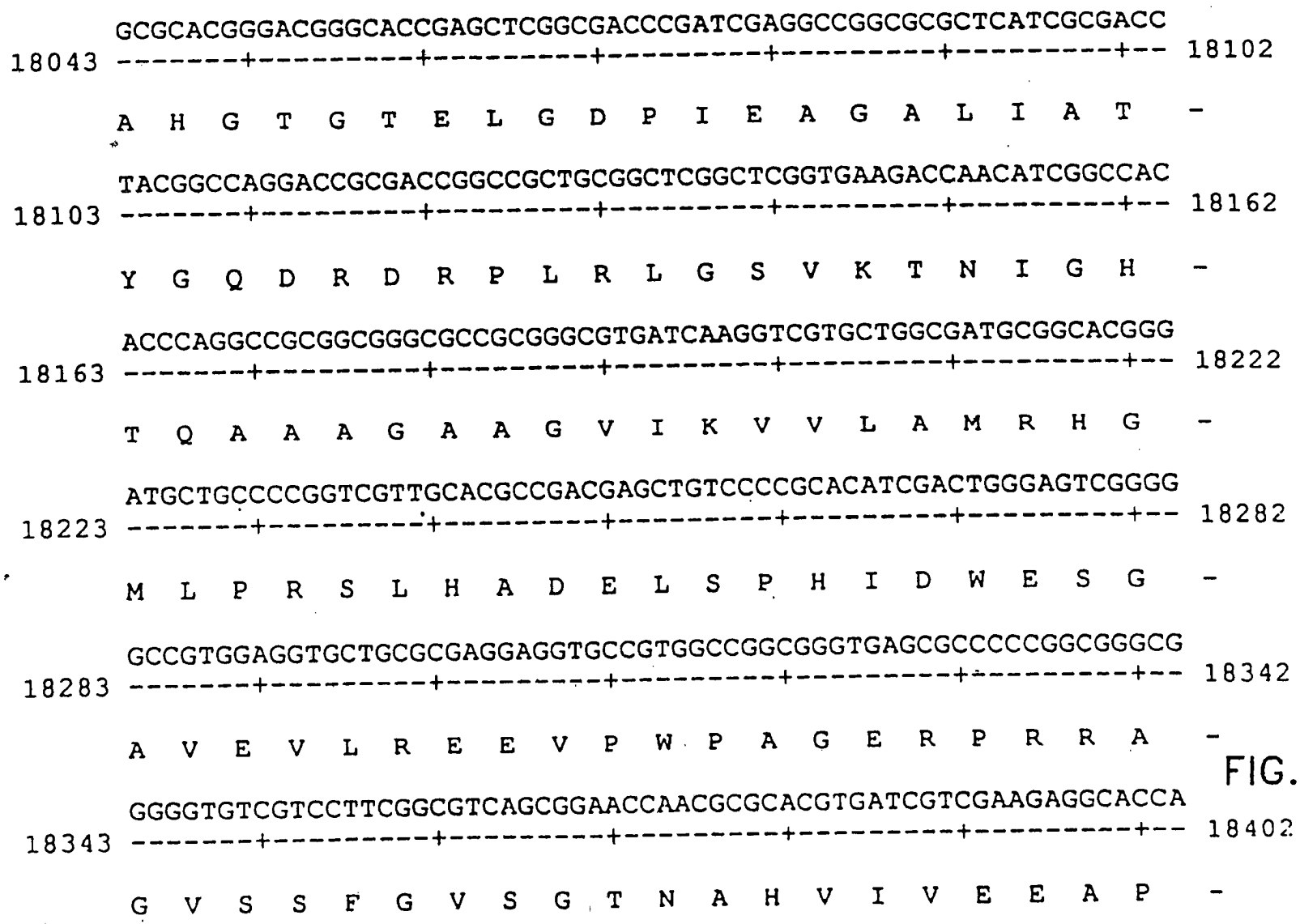
GCGCACGGGACGGGCACCGAGCTCGGCGACCCGATCGAGGCCGGCGCGCTCATCGCGACC
18043 -------+---------+---------+---------+---------+---------+-- 18102
A H G T G T E L G D P I E A G A L I A T -
TACGGCCAGGACCGCGACCGGCCGCTGCGGCTCGGCTCGGTGAAGACCAACATCGGCCAC
18103 -------+---------+---------+---------+---------+---------+-- 18162
Y G Q D R D R P L R L G S V K T N I G H -
ACCCAGGCCGCGGCGGGCGCCGCGGGCGTGATCAAGGTCGTGCTGGCGATGCGGCACGGG
18163 -------+---------+---------+---------+---------+---------+-- 18222
T Q A A A G A A G V I K V V L A M R H G -
ATGCTGCCCCGGTCGTTGCACGCCGACGAGCTGTCCCCGCACATCGACTGGGAGTCGGGG
18223 -------+---------+---------+---------+---------+---------+-- 18282
M L P R S L H A D E L S P H I D W E S G -
GCCGTGGAGGTGCTGCGCGAGGAGGTGCCGTGGCCGGCGGGTGAGCGCCCCCGGCGGGCG
18283 -------+---------+---------+---------+---------+---------+-- 18342
A V E V L R E E V P W P A G E R P R R A -
GGGGTGTCGTCCTTCGGCGTCAGCGGAACCAACGCGCACGTGATCGTCGAAGAGGCACCA
18343 -------+---------+---------+---------+---------+---------+-- 18402
G V S S F G V S G T N A H V I V E E A P -

FIG.2-46

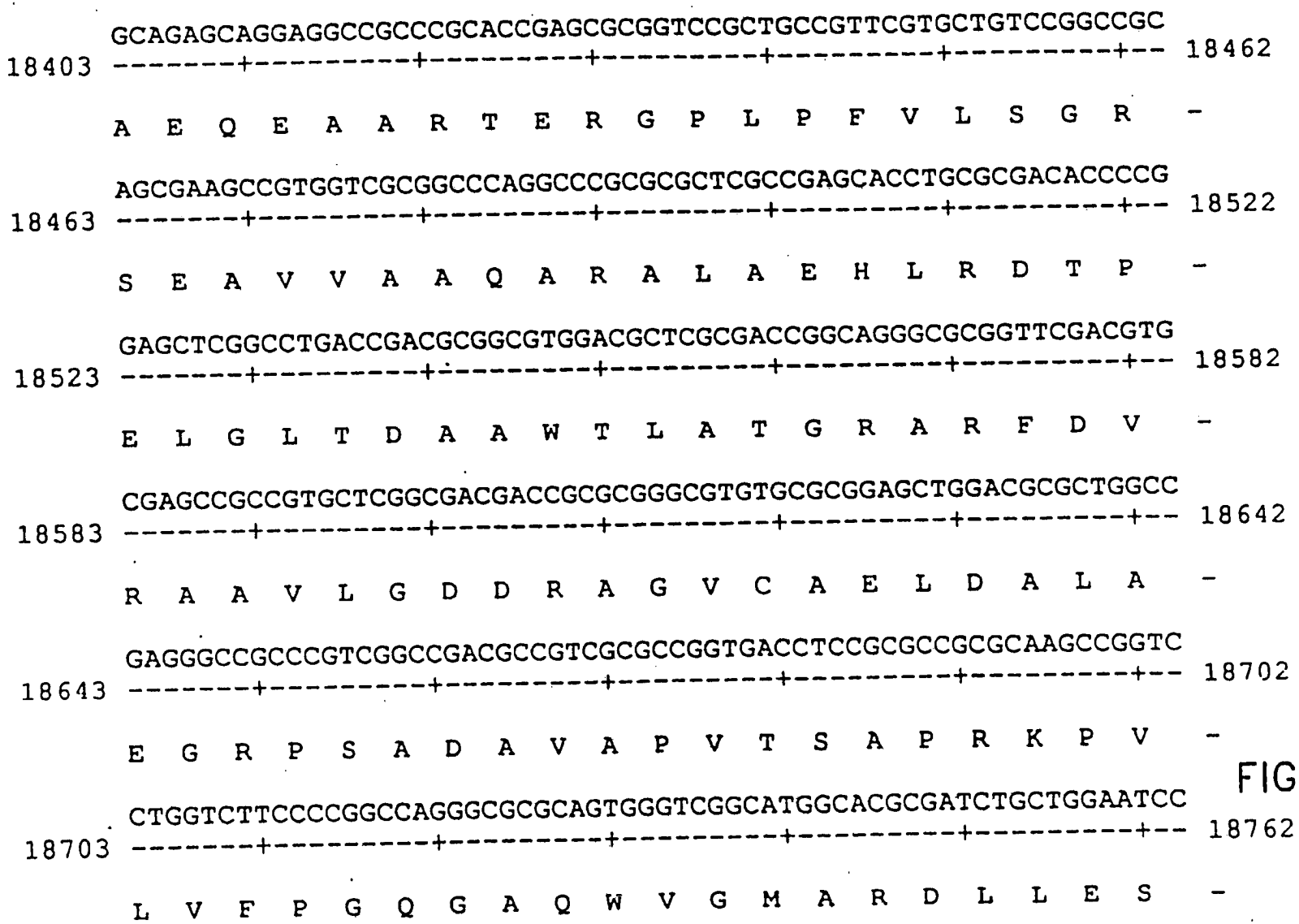
GCAGAGCAGGAGGCCGCCCGCACCGAGCGCGGTCCGCTGCCGTTCGTGCTGTCCGGCCGC
18403 -------+---------+---------+---------+---------+---------+-- 18462
A E Q E A A R T E R G P L P F V L S G R -
AGCGAAGCCGTGGTCGCGGCCCAGGCCCGCGCGCTCGCCGAGCACCTGCGCGACACCCCG
18463 -------+---------+---------+---------+---------+---------+-- 18522
S E A V V A A Q A R A L A E H L R D T P -
GAGCTCGGCCTGACCGACGCGGCGTGGACGCTCGCGACCGGCAGGGCGCGGTTCGACGTG
18523 -------+---------+---------+---------+---------+---------+-- 18582
E L G L T D A A W T L A T G R A R F D V -
CGAGCCGCCGTGCTCGGCGACGACCGCGCGGGCGTGTGCGCGGAGCTGGACGCGCTGGCC
18583 -------+---------+---------+---------+---------+---------+-- 18642
R A A V L G D D R A G V C A E L D A L A -
GAGGGCCGCCCGTCGGCCGACGCCGTCGCGCCGGTGACCTCCGCGCCGCGCAAGCCGGTC
18643 -------+---------+---------+---------+---------+---------+-- 18702
E G R P S A D A V A P V T S A P R K P V -
CTGGTCTTCCCCGGCCAGGGCGCGCAGTGGGTCGGCATGGCACGCGATCTGCTGGAATCC
18703 -------+---------+---------+---------+---------+---------+-- 18762
L V F P G Q G A Q W V G M A R D L L E S -

FIG.2-47

```
                TCCGAGGTGTTCGCCGAGTCGATGAGCCGGTGCGCCGAGGCGCTCTCGCCGCACACCGAC
18763 ----------+----------+----------+----------+----------+----------+---- 18822
                S  E  V  F  A  E  S  M  S  R  R  C  A  E  E  A  L  S  P  H  T  D  -

                TGGAAGTTGCTCGACGTCGTCCGCGGCGACGGCGGTCCCGACCCGCACGAGCGCGTCGAC
18823 ----------+----------+----------+----------+----------+----------+---- 18882
                W  K  L  L  D  V  V  R  G  D  G  G  P  D  P  H  E  R  V  D  -

                GTGCTCCAGCCGGTGCTCTTCTCGATCATGGTCTCGCTGGCCGAGCTGTGGCGCGCGCAC
18883 ----------+----------+----------+----------+----------+----------+---- 18942
                V  L  Q  P  V  L  F  S  I  M  V  S  L  A  E  L  W  R  A  H  -

                GGCGTGACCCCGGCCGCGTCGTCGGCCACTCGCAGGGCGAGATCGCCGCGGCGCACGTG
18943 ----------+----------+----------+----------+----------+----------+---- 19002
                G  V  T  P  A  A  V  V  V  G  H  S  Q  G  E  I  A  A  A  H  V  -

                GCGGGCGCGCTGTCGCTGGAAGCCGCCGCGAAGGTGGTGGCCCTGCGCAGCCAGGTGTTG
19003 ----------+----------+----------+----------+----------+----------+---- 19062
                A  G  A  L  S  L  E  A  A  A  K  V  V  V  A  L  R  S  Q  V  L  -

                CGCGAGCTCGACGACCAGGGCGGCATGGTGTCGGTCGGCGCGTCCCGCGACGAGCTGGAG
19063 ----------+----------+----------+----------+----------+----------+---- 19122
```

FIG.2-48

```
      R  E  L  D  D  Q  G  G  M  V  S  V  G  A  S  R  D  E  L  E   -

      ACCGTGCTCGCGCGCTGGGACGGCCGTGTCGCGGTGGCCGCCGTGAACGGGCCTGGCACC
19123 -------+---------+---------+---------+---------+---------+--- 19182

      T  V  L  A  R  W  D  G  R  V  A  V  A  A  V  N  G  P  G  T   -

      AGCGTCGTTGCCGGGCCGACCGCGGAGCTGGACGAGTTCTTCGCCGAGGCCGAGGCGCGG
19183 -------+---------+---------+---------+---------+---------+--- 19242

      S  V  V  A  G  P  T  A  E  L  D  E  F  F  A  E  A  E  A  R   -

      GAGATGAAGCCGCGCCGGATCGCCGTGCGCTACGCCTCCCACTCCCCGGAGGTGGCGCGC
19243 -------+---------+---------+---------+---------+---------+--- 19302

      E  M  K  P  R  R  I  A  V  R  Y  A  S  H  S  P  E  V  A  R   -

      ATCGAGGACCGGCTCGCGGCCGAGCTGGGCACCATCACCGCCGTGCGGGGCTCGGTGCCG
19303 -------+---------+---------+---------+---------+---------+--- 19362

      I  E  D  R  L  A  A  E  L  G  T  I  T  A  V  R  G  S  V  P   -

      CTGCACTCCACGGTGACCGGCGAGGTCATCGACACCTCCGCGATGGACGCCTCCTACTGG
19363 -------+---------+---------+---------+---------+---------+--- 19422

      L  H  S  T  V  T  G  E  V  I  D  T  S  A  M  D  A  S  Y  W   -

      TACCGCAACCTGCGCCGACCAGTGCTCTTCGAGCAGGCGGTGCGCGGTCTGGTCGAGCAG
19423 -------+---------+---------+---------+---------+---------+--- 19482
```

FIG.2—49

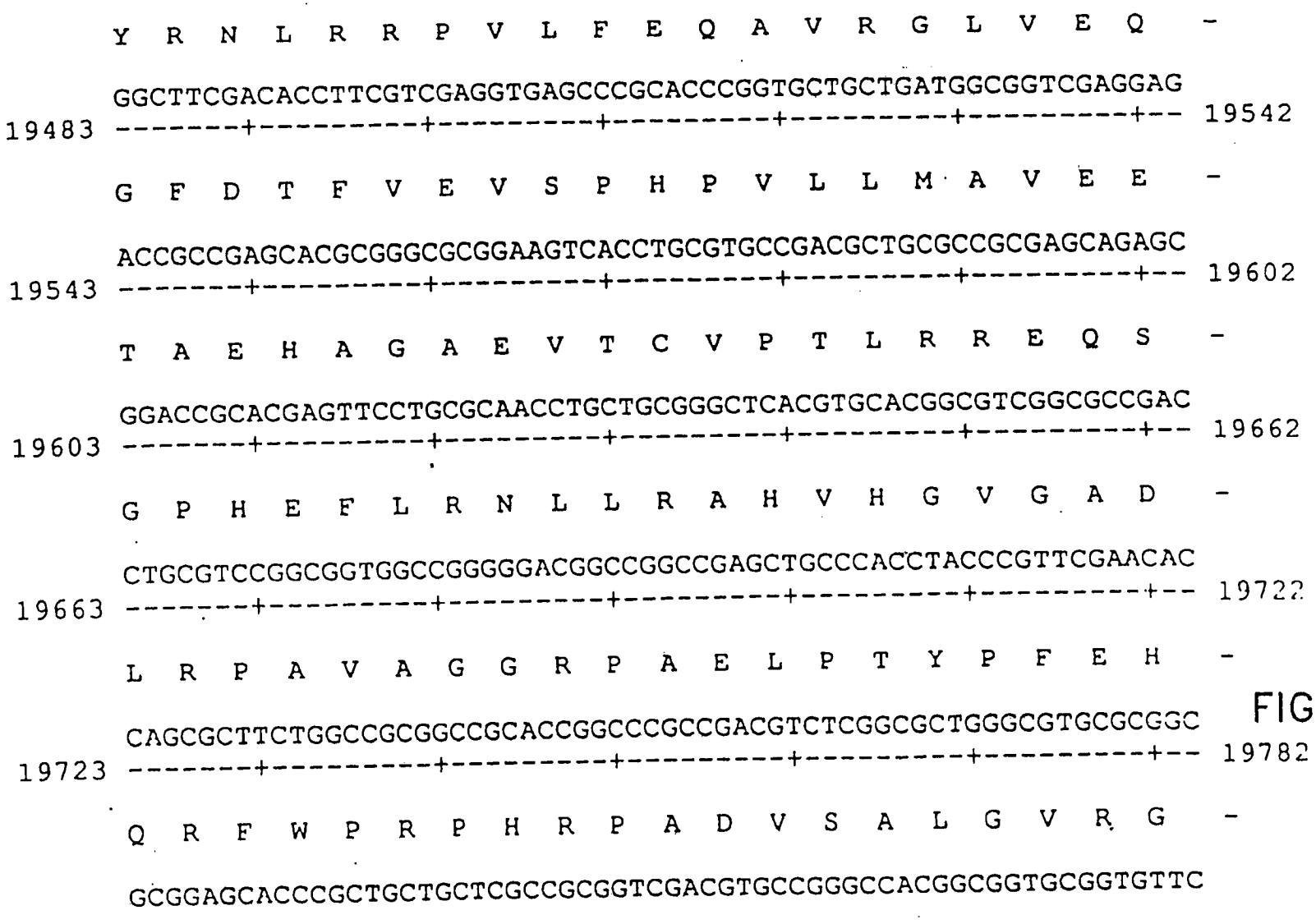

FIG.2-50

SUBSTITUTE SHEET

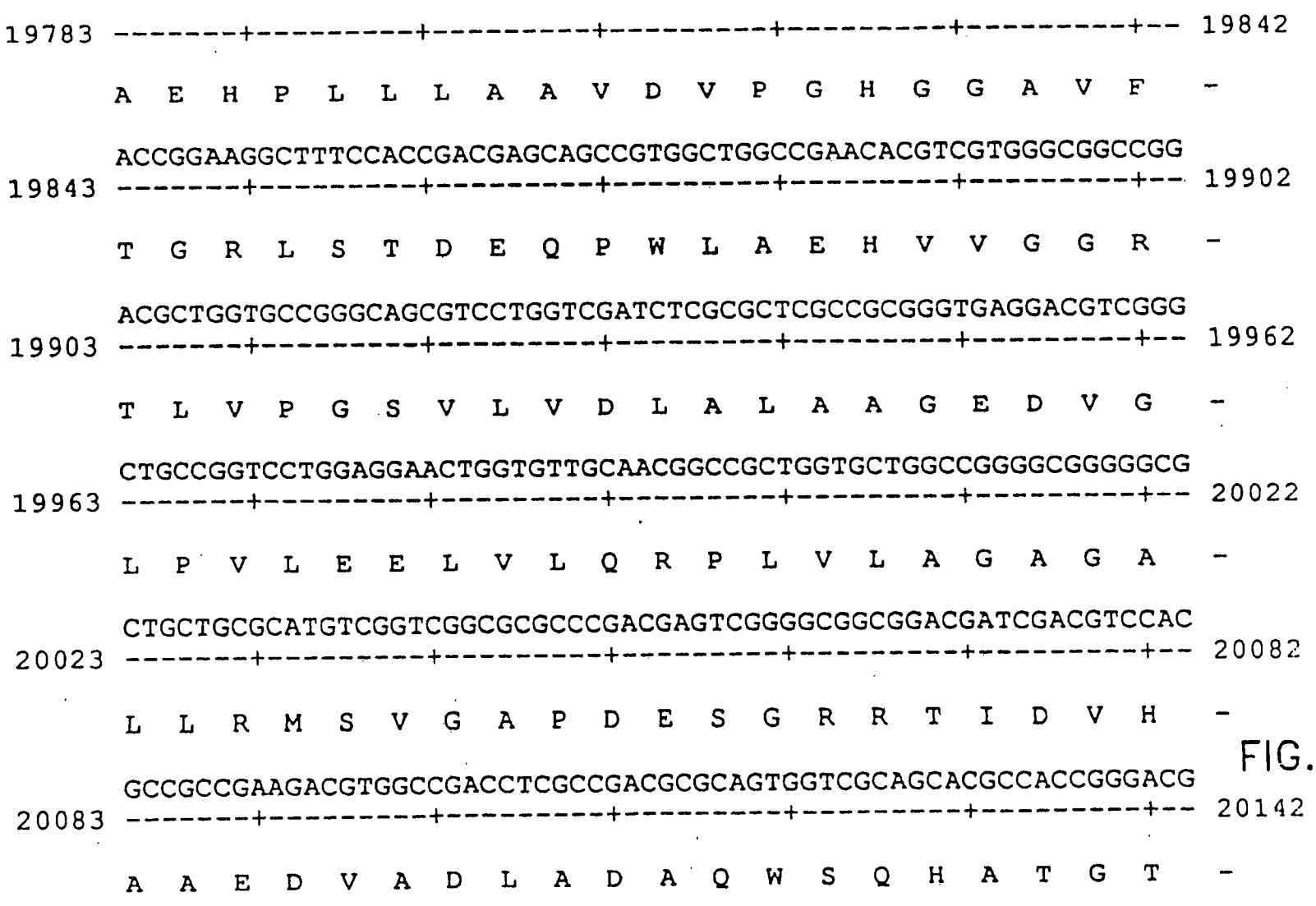

FIG.2-51

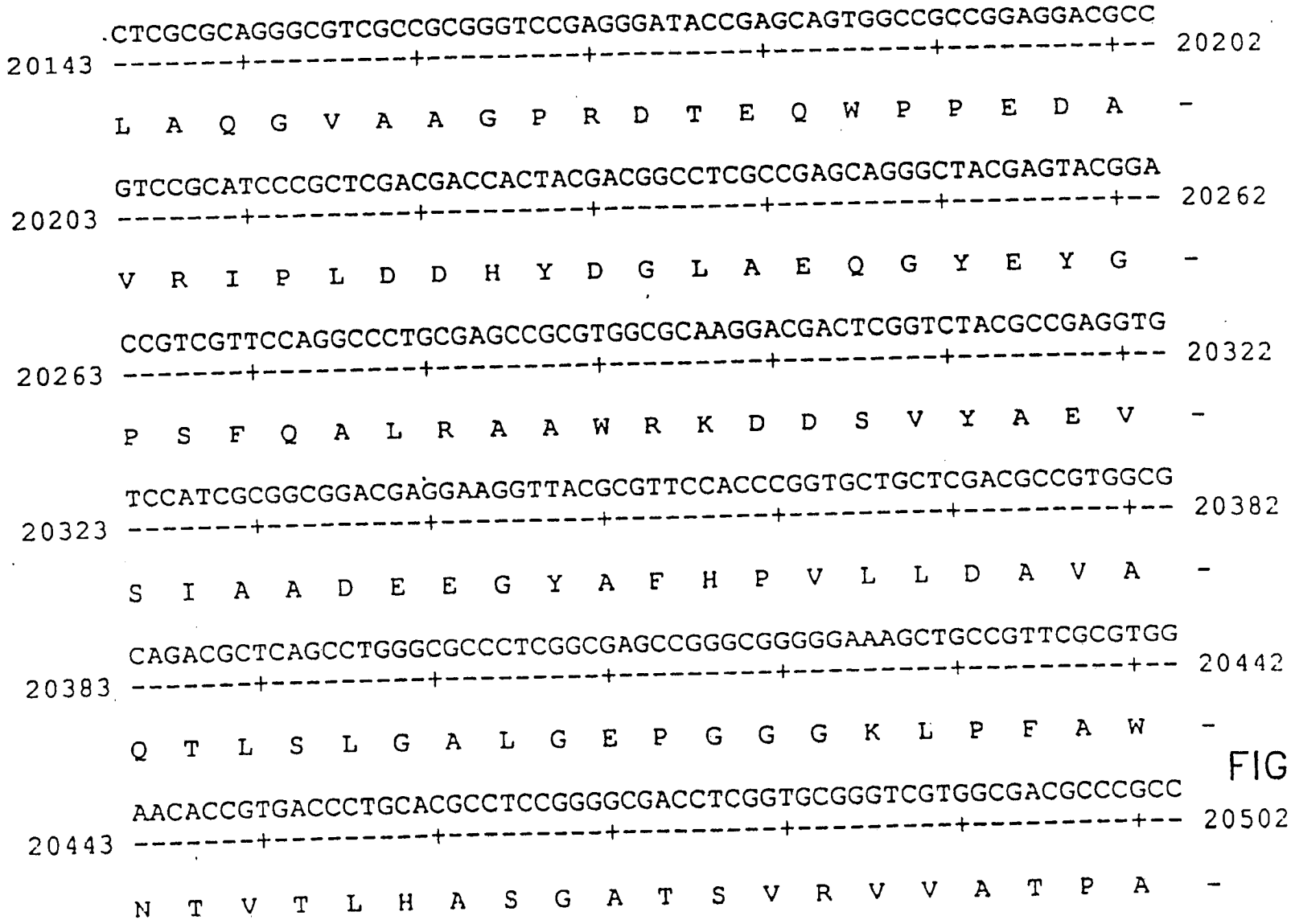

FIG.2-52

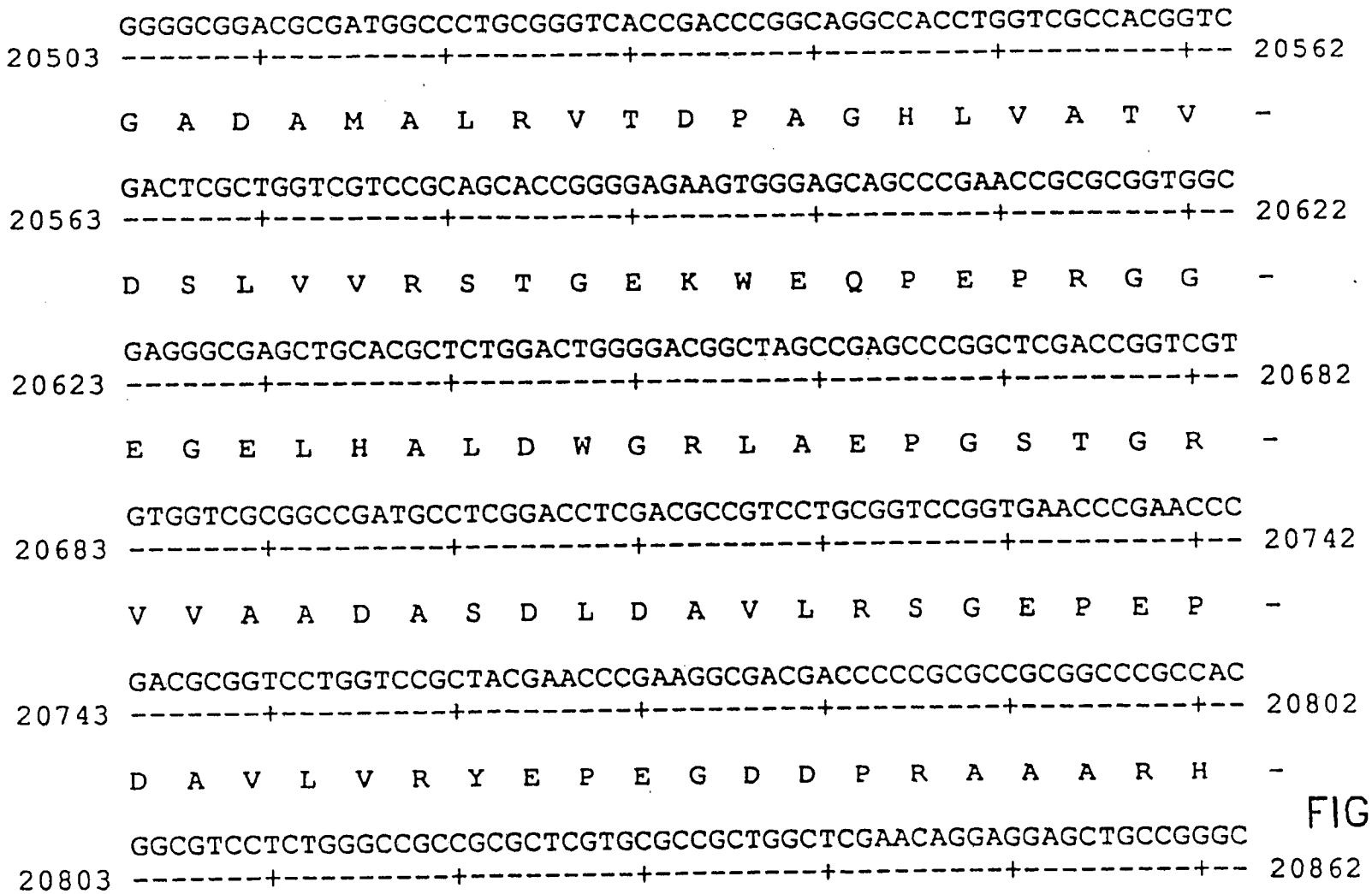
GGGGCGGACGCGATGGCCCTGCGGGTCACCGACCCGGCAGGCCACCTGGTCGCCACGGTC
20503 -------+---------+---------+---------+---------+---------+---------+-- 20562
G A D A M A L R V T D P A G H L V A T V -
GACTCGCTGGTCGTCCGCAGCACCGGGGAGAAGTGGGAGCAGCCCGAACCGCGCGGTGGC
20563 -------+---------+---------+---------+---------+---------+---------+-- 20622
D S L V V R S T G E K W E Q P E P R G G -
GAGGGCGAGCTGCACGCTCTGGACTGGGGACGGCTAGCCGAGCCCGGCTCGACCGGTCGT
20623 -------+---------+---------+---------+---------+---------+---------+-- 20682
E G E L H A L D W G R L A E P G S T G R -
GTGGTCGCGGCCGATGCCTCGGACCTCGACGCCGTCCTGCGGTCCGGTGAACCCGAACCC
20683 -------+---------+---------+---------+---------+---------+---------+-- 20742
V V A A D A S D L D A V L R S G E P E P -
GACGCGGTCCTGGTCCGCTACGAACCCGAAGGCGACGACCCCCGCGCCGCGGCCCGCCAC
20743 -------+---------+---------+---------+---------+---------+---------+-- 20802
D A V L V R Y E P E G D D P R A A A R H -
GGCGTCCTCTGGGCCGCCGCGCTCGTGCGCCGCTGGCTCGAACAGGAGGAGCTGCCGGGC
20803 -------+---------+---------+---------+---------+---------+---------+-- 20862

FIG.2-53

```
      G  V  L  W  A  A  A  L  V  R  R  W  L  E  Q  E  E  L  P  G   -

      GCGACGCTGGTCATCGCCACGTCCGGCGCGGTCACCGTGTCCGACGACGACAGCGTTCCC
20863 -------+---------+---------+---------+---------+---------+-- 20922

      A  T  L  V  I  A  T  S  G  A  V  T  V  S  D  D  D  S  V  P   -

      GAACCCGGCGCCGCCGCGATGTGGGGCGTGATCCGCTGTGCGCAGGCCGAGTCGCCGGAC
20923 -------+---------+---------+---------+---------+---------+-- 20982

      E  P  G  A  A  A  M  W  G  V  I  R  C  A  Q  A  E  S  P  D   -

      CGGTTCGTGCTCCTCGACACCGACGCGGAACCTGGGATGCTGCCTGCGGTTCCGGACAAC
20983 -------+---------+---------+---------+---------+---------+-- 21042

      R  F  V  L  L  D  T  D  A  E  P  G  M  L  P  A  V  P  D  N   -

      CCGCAGCTCGCGTTGCGCGGCGACGACGTCTTCGTGCCGCGCCTCTCGCCGCTCGCACCT
21043 -------+---------+---------+---------+---------+---------+-- 21102

      P  Q  L  A  L  R  G  D  D  V  F  V  P  R  L  S  P  L  A  P   -

      TCCGCGCTGACGCTTCCGGCAGGCACCCAACGTCTCGTGCCGGGTGACGGGGCGATCGAC
21103 -------+---------+---------+---------+---------+---------+-- 21162

      S  A  L  T  L  P  A  G  T  Q  R  L  V  P  G  D  G  A  I  D   -

      TCCGTGGCCTTCGAGCCCGCACCCGACGTCGAGCAGCCGCTCCGGGCGGGCGAGGTCCGG
21163 -------+---------+---------+---------+---------+---------+-- 21222
```

FIG.2-54

```
      S  V  A  F  E  P  A  P  D  V  E  Q  P  L  R  A  G  E  V  R  -

      GTGGACGTGCGCGCCACCGGAGTCAACTTCCGCGACGTCCTCCTCGCACTCGGCATGTAT
21223 -------+---------+---------+---------+---------+---------+-- 21282

      V  D  V  R  A  T  G  V  N  F  R  D  V  L  L  A  L  G  M  Y  -

      CCGCAGAAGGCGGACATGGGCACCGAGGCCGCCGGTGTCGTCACGGCGGTCGGACCGGAC
21283 -------+---------+---------+---------+---------+---------+-- 21342

      P  Q  K  A  D  M  G  T  E  A  A  G  V  V  T  A  V  G  P  D  -

      GTGGACGCCTTCGCGCCGGGAGACCGGGTGCTCGGCCTGTTCCAGGGAGCCTTCGCGCCG
21343 -------+---------+---------+---------+---------+---------+-- 21402

      V  D  A  F  A  P  G  D  R  V  L  G  L  F  Q  G  A  F  A  P  -

      ATCGCGGTCACCGATCACCGGCTCCTCGCACGAGTGCCGGACGGCTGGAGCGACGCCGAC
21403 -------+---------+---------+---------+---------+---------+-- 21462

      I  A  V  T  D  H  R  L  L  A  R  V  P  D  G  W  S  D  A  D  -

      GCCGCGGCCGTGCCCATCGCCTACACCACGGCGCATTACGCGCTGCACGATCTCGCGGGG
21463 -------+---------+---------+---------+---------+---------+-- 21522

      A  A  A  V  P  I  A  Y  T  T  A  H  Y  A  L  H  D  L  A  G  -

      CTGCGCGCGGGTCAGTCGGTGCTCATCCACGCAGCGGCAGGCGGTGTCGGCATGGCGGCC
21523 -------+---------+---------+---------+---------+---------+-- 21582
```

FIG.2-55

```
                L  R  A  G  G  Q  S  S  V  L  I  I  H  A  A  A  A  G  G  G  V  G  G  M  A  A  -
                GTCGCGCTGGCCCGCCGAGCGGGGGGCGGAGGTGTTGGCCACCGCCGGCCCGGCCAAGCAC
21583 ----------+---------+---------+---------+---------+---------+---------+---------+-- 21642
                V  A  L  A  R  R  A  G  A  E  E  V  L  A  T  A  G  P  A  K  H  -
                GGGACGCTGCGGGCGCTCGGTCTCGACGACGAGCACATCGCTTCCTCCCGGGAGACCGGT
21643 ----------+---------+---------+---------+---------+---------+---------+---------+-- 21702
                G  T  L  R  A  L  G  L  D  D  E  H  I  A  S  S  R  E  T  G  -
                TTCGCCCCGGAAGTTCCGGGAGCGCACCGGAGGCCGCGGCGTGGACGTGGTGCTCAACTCG
21703 ----------+---------+---------+---------+---------+---------+---------+---------+-- 21762
                F  A  R  K  F  R  E  R  T  G  G  R  G  V  D  V  V  L  N  S  -
                CTCACCGGGGAACTGCTCGACGAGTCCGCGGATCTGCTCGCCGAGGACGGCGTCTTCGTC
21763 ----------+---------+---------+---------+---------+---------+---------+---------+-- 21822
                L  T  G  E  L  L  D  E  S  A  D  L  L  A  E  D  G  V  F  V  -
                GAGATGGGCAAGACCGACCTGCGGGACGCCGGGGGCCGATACGCCCCGTTC
21823 ----------+---------+---------+---------+---------+---------+---------+---------+-- 21882
                E  M  G  K  T  D  L  R  D  A  G  D  F  R  G  R  Y  A  P  F  -
```

FIG.2-56

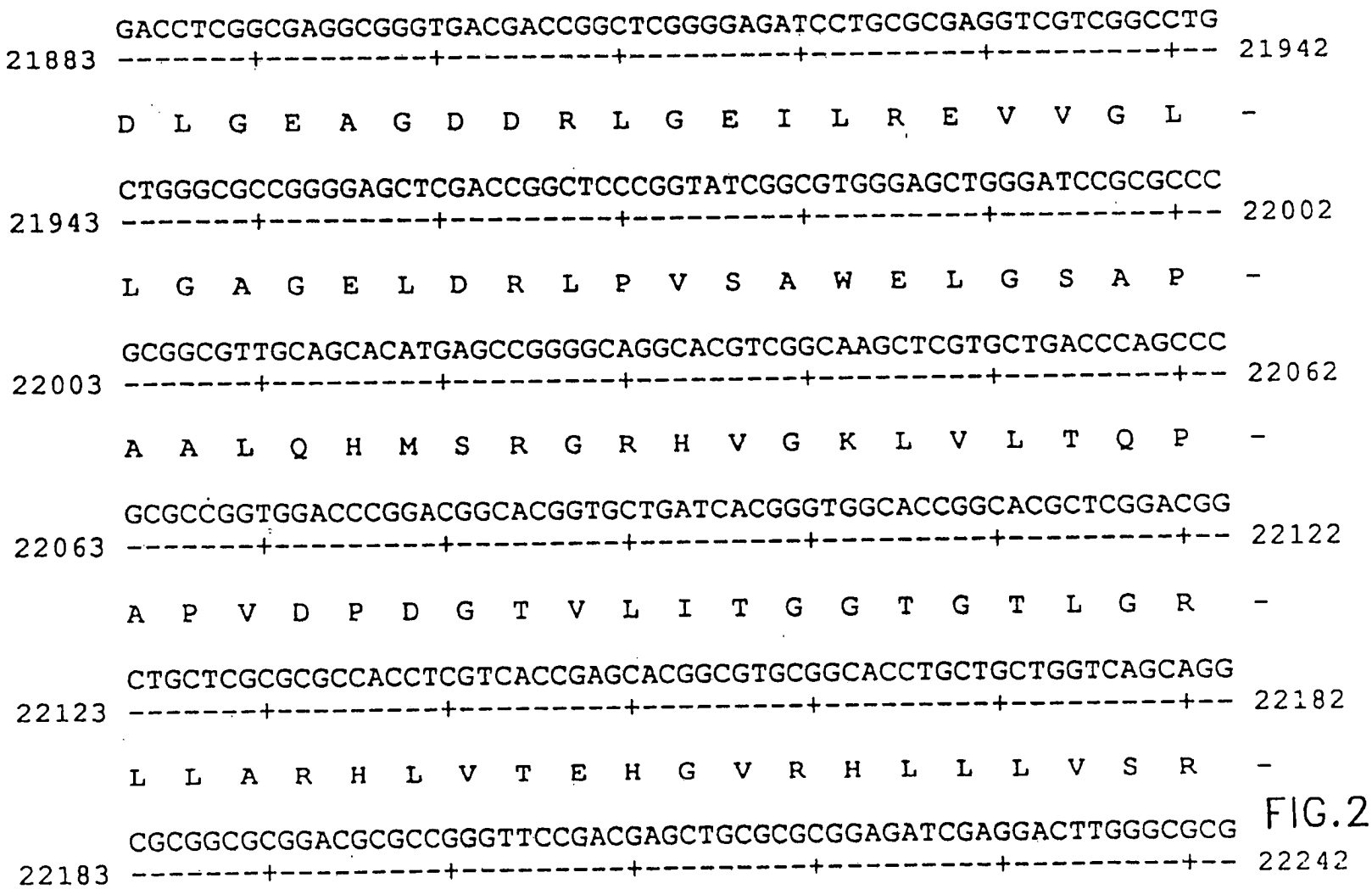
GACCTCGGCGAGGCGGGTGACGACCGGCTCGGGGAGATCCTGCGCGAGGTCGTCGGCCTG
21883 21942
D L G E A G D D R L G E I L R E V V G L -
CTGGGCGCCGGGGAGCTCGACCGGCTCCCGGTATCGGCGTGGGAGCTGGGATCCGCGCCC
21943 22002
L G A G E L D R L P V S A W E L G S A P -
GCGGCGTTGCAGCACATGAGCCGGGGCAGGCACGTCGGCAAGCTCGTGCTGACCCAGCCC
22003 22062
A A L Q H M S R G R H V G K L V L T Q P -
GCGCCGGTGGACCCGGACGGCACGGTGCTGATCACGGGTGGCACCGGCACGCTCGGACGG
22063 22122
A P V D P D G T V L I T G G T G T L G R -
CTGCTCGCGCGCCACCTCGTCACCGAGCACGGCGTGCGGCACCTGCTGCTGGTCAGCAGG
22123 22182
L L A R H L V T E H G V R H L L L V S R -
CGCGGCGCGGACGCGCCGGGTTCCGACGAGCTGCGCGCGGAGATCGAGGACTTGGGCGCG
22183 22242

FIG.2-57

```
      R  G  A  D  A  P  G  S  D  E  L  R  A  E  I  E  D  L  G  A  -

      TCCGCGGAGATCGCGGCTTGCGACACCGCCGACCGCGACGCGCTTTCGGCGCTGCTGGAC
22243 -------+---------+---------+---------+---------+---------+-- 22302

      S  A  E  I  A  A  C  D  T  A  D  R  D  A  L  S  A  L  L  D  -

      GGGCTGCCCCGGCCGCTGACCGGTGTCGTGCACGCGGCGGGTGTGCTGGCCGACGGGCTG
22303 -------+---------+---------+---------+---------+---------+-- 22362

      G  L  P  R  P  L  T  G  V  V  H  A  A  G  V  L  A  D  G  L  -

      GTCACCTCCATCGACGAGCCGGCGGTGGAGCAGGTGCTGCGCGCCAAGGTCGACGCGGCG
22363 -------+---------+---------+---------+---------+---------+-- 22422

      V  T  S  I  D  E  P  A  V  E  Q  V  L  R  A  K  V  D  A  A  -

      TGGAACCTGCACGAGCTGACCGCGAACACCGGTCTGAGCTTCTTCGTGCTGTTCTCGTCC
22423 -------+---------+---------+---------+---------+---------+-- 22482

      W  N  L  H  E  L  T  A  N  T  G  L  S  F  F  V  L  F  S  S  -

      GCGGCGTCGGTGCTAGCCGGCCCGGGGCAGGGCGTGTACGCGGCCGCGAACGAGTCGCTC
22483 -------+---------+---------+---------+---------+---------+-- 22542

      A  A  S  V  L  A  G  P  G  Q  G  V  Y  A  A  A  N  E  S  L  -

      AACGCGCTGGCTGCCCTCCGGAGGACGCGCGGCCTTCCCGCGAAGGCGCTCGGATGGGGA
22543 -------+---------+---------+---------+---------+---------+-- 22602
```

FIG.2-58

```
      N  A  L  A  A  L  R  R  T  R  G  L  P  A  K  A  L  G  W  G  -

      CTGTGGGCGCAGGCCAGCGAGATGACCAGCGGACTCGGCGACCGCATCGCCCGGACCGGG
22603 -------+---------+---------+---------+---------+---------+-- 22662

      L  W  A  Q  A  S  E  M  T  S  G  L  G  D  R  I  A  R  T  G  -

      GTCGCCGCGCTGCCGACCGAGCGGGCGCTCGCACTGTTCGACAGCGCCCTGCGCCGCGGC
22663 -------+---------+---------+---------+---------+---------+-- 22722

      V  A  A  L  P  T  E  R  A  L  A  L  F  D  S  A  L  R  R  G  -

      GGTGAGGTCGTGTTCCCGCTGTCCATCAACCGTTCCGCGCTGCGCAGGGCCGAGTTCGTG
22723 -------+---------+---------+---------+---------+---------+-- 22782

      G  E  V  V  F  P  L  S  I  N  R  S  A  L  R  R  A  E  F  V  -

      CCGGAGGTCCTGCGCGGCATGGTCAGGGCGAAGCTGCGCGCCGCCGGGCAGGCCGAGGCG
22783 -------+---------+---------+---------+---------+---------+-- 22842

      P  E  V  L  R  G  M  V  R  A  K  L  R  A  A  G  Q  A  E  A  -

      GCAGGGCCGAACGTGGTCGACCGGCTCGCCGGTCGGTCCGAGTCCGACCAGGTCGCCGGG
22843 -------+---------+---------+---------+---------+---------+-- 22902

      A  G  P  N  V  V  D  R  L  A  G  R  S  E  S  D  Q  V  A  G  -

      CTGGCCGAACTGGTGCGTTCACACGCGGCGGCGGTCTCCGGGTACGGCTCGGCCGACCAG
22903 -------+---------+---------+---------+---------+---------+-- 22962
```

FIG.2-59

```
      L  A  E  L  V  R  S  H  A  A  A  V  S  G  Y  G  S  A  D  Q  -

      CTCCCCGAGCGCAAGGCGTTCAAGGACCTCGGTTTCGACTCGCTGGCCGCGGTGGAGCTG
22963 -------+---------+---------+---------+---------+---------+-- 23022

      L  P  E  R  K  A  F  K  D  L  G  F  D  S  L  A  A  V  E  L  -

      CGCAACCGCCTCGGTACCGCGACCGGCGTGCGGCTGCCCAGCACGTTGGTGTTCGACCAC
23023 -------+---------+---------+---------+---------+---------+-- 23082

      R  N  R  L  G  T  A  T  G  V  R  L  P  S  T  L  V  F  D  H  -

      CCGACTCCGCTGGCGGTGGCCGAACACCTGCGGGACAGGCTGTTCGCGGCCTCACCGGCG
23083 -------+---------+---------+---------+---------+---------+-- 23142

      P  T  P  L  A  V  A  E  H  L  R  D  R  L  F  A  A  S  P  A  -

      GTGGACATCGGCGACCGGCTGGACGAGCTGGAGAAGGCGCTCGAAGCCCTGTCCGCCGAG
23143 -------+---------+---------+---------+---------+---------+-- 23202

      V  D  I  G  D  R  L  D  E  L  E  K  A  L  E  A  L  S  A  E  -

      GACGGGCACGACGACGTGGGCCAGCGCCTGGAGTCGCTGCTGCGCCGGTGGAACAGCAGG
23203 -------+---------+---------+---------+---------+---------+-- 23262

      D  G  H  D  D  V  G  Q  R  L  E  S  L  L  R  R  W  N  S  R  -

      CGGGCGGACGCCCCGAGCACGTCCGCGATCAGCGAGGACGCCAGTGACGACGAGCTGTTC
```

FIG.2-60

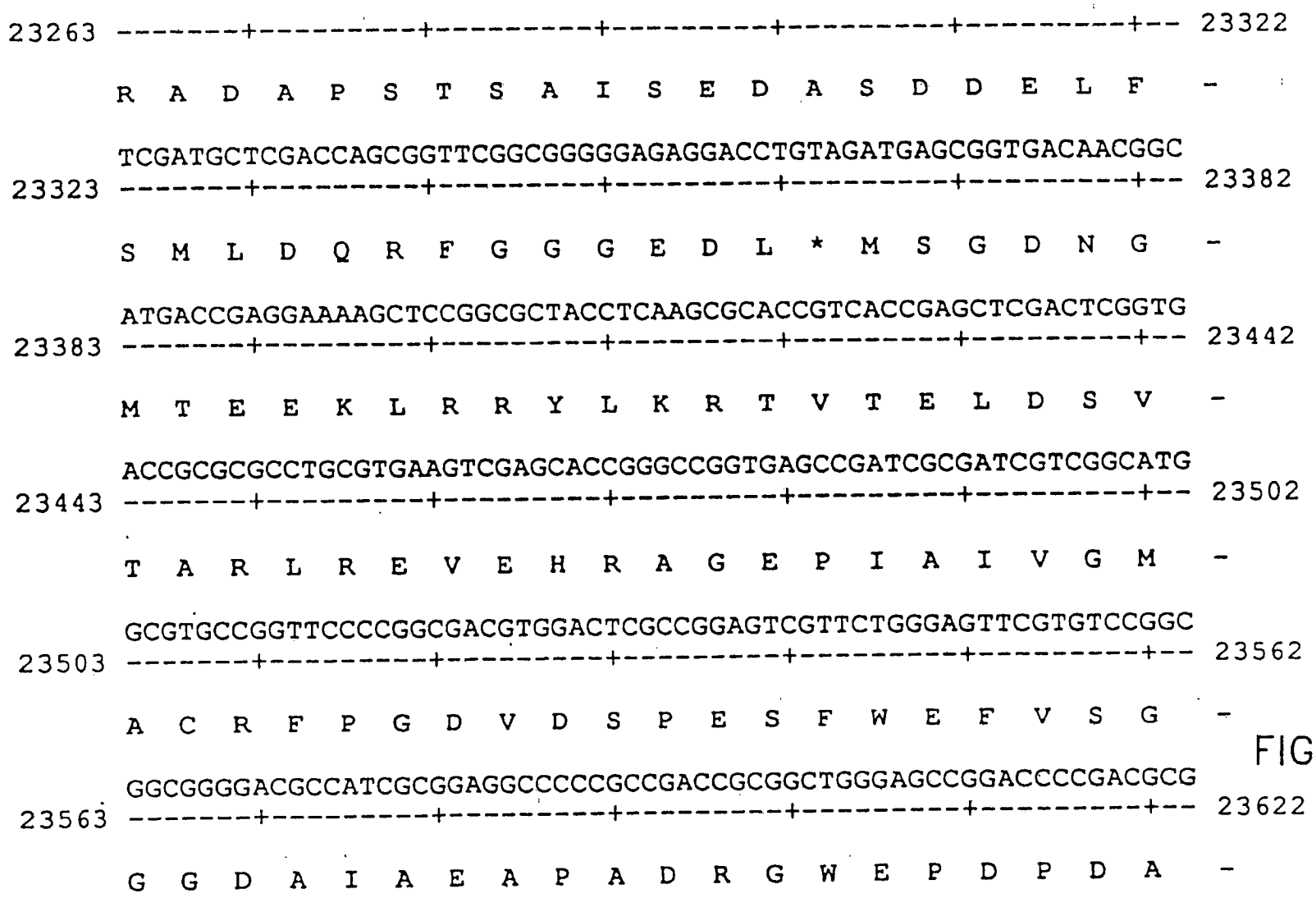
23263 -------+---------+---------+---------+---------+---------+-- 23322
R A D A P S T S A I S E D A S D D E L F -
TCGATGCTCGACCAGCGGTTCGGCGGGGGAGAGGACCTGTAGATGAGCGGTGACAACGGC
23323 -------+---------+---------+---------+---------+---------+-- 23382
S M L D Q R F G G G E D L * M S G D N G -
ATGACCGAGGAAAAGCTCCGGCGCTACCTCAAGCGCACCGTCACCGAGCTCGACTCGGTG
23383 -------+---------+---------+---------+---------+---------+-- 23442
M T E E K L R R Y L K R T V T E L D S V -
ACCGCGCGCCTGCGTGAAGTCGAGCACCGGGCCGGTGAGCCGATCGCGATCGTCGGCATG
23443 -------+---------+---------+---------+---------+---------+-- 23502
T A R L R E V E H R A G E P I A I V G M -
GCGTGCCGGTTCCCCGGCGACGTGGACTCGCCGGAGTCGTTCTGGGAGTTCGTGTCCGGC
23503 -------+---------+---------+---------+---------+---------+-- 23562
A C R F P G D V D S P E S F W E F V S G -
GGCGGGGACGCCATCGCGGAGGCCCCCGCCGACCGCGGCTGGGAGCCGGACCCCGACGCG
23563 -------+---------+---------+---------+---------+---------+-- 23622
G G D A I A E A P A D R G W E P D P D A -

FIG.2-61

```
      CGGCTGGGCGGGATGCTCGCGGCCGCGGGCGACTTCGACGCGGGCTTCTTCGGGATCTCG
23623 -------+---------+---------+---------+---------+---------+-- 23682

      R  L  G  G  M  L  A  A  A  G  D  F  D  A  G  F  F  G  I  S  -

      CCGCGCGAGGCGCTGGCGATGGACCCGCAGCAGCGGATCATGCTGGAGATCTCGTGGGAG
23683 -------+---------+---------+---------+---------+---------+-- 23742

      P  R  E  A  L  A  M  D  P  Q  Q  R  I  M  L  E  I  S  W  E  -

      GCGCTGGAGCGCGCCGGCCACGATCCGGTGTCCCTGCGCGGCAGCGCGACCGGGGTGTTC
23743 -------+---------+---------+---------+---------+---------+-- 23802

      A  L  E  R  A  G  H  D  P  V  S  L  R  G  S  A  T  G  V  F  -

      ACCGGTGTCGGCACCGTGGACTACGGCCCGCGACCCGACGAGGCCCCGGACGAGGTCCTG
23803 -------+---------+---------+---------+---------+---------+-- 23862

      T  G  V  G  T  V  D  Y  G  P  R  P  D  E  A  P  D  E  V  L  -

      GGCTACGTCGGCACCGGCACCGCCTCCAGCGTCGCCTCCGGCCGGGTCGCCTACTGCCTG
23863 -------+---------+---------+---------+---------+---------+-- 23922

      G  Y  V  G  T  G  T  A  S  S  V  A  S  G  R  V  A  Y  C  L  -

      GGCCTGGAAGGCCCGGCGATGACCGTCGACACCGCCTGTTCCTCCGGGCTCACCGCCCTG
23923 -------+---------+---------+---------+---------+---------+-- 23982

      G  L  E  G  P  A  M  T  V  D  T  A  C  S  S  G  L  T  A  L  -
```

FIG.2-62

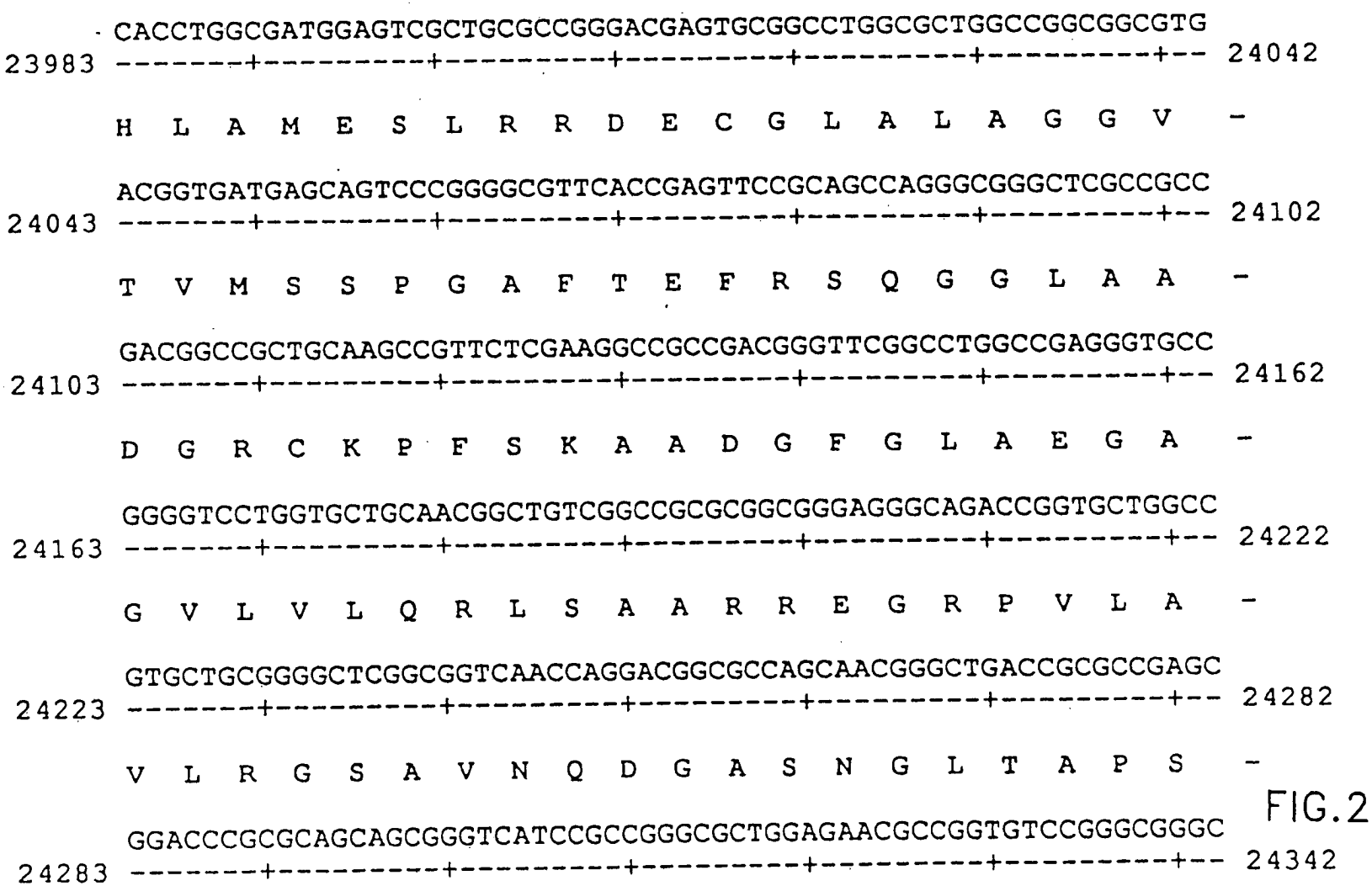

FIG.2-63

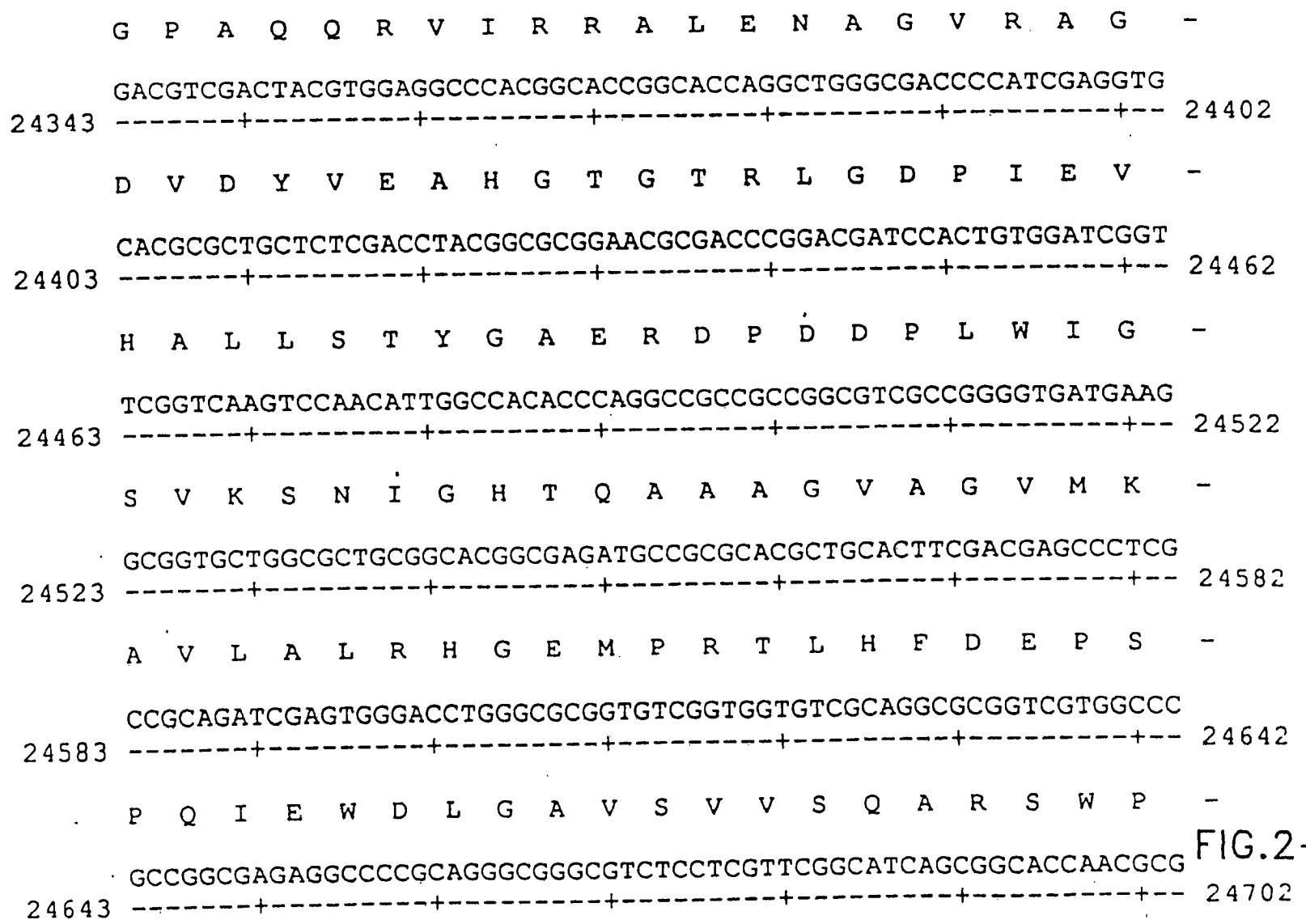
G P A Q Q R V I R R A L E N A G V R A G -
GACGTCGACTACGTGGAGGCCCACGGCACCGGCACCAGGCTGGGCGACCCCATCGAGGTG
24343 -------+---------+---------+---------+---------+---------+-- 24402
D V D Y V E A H G T G T R L G D P I E V -
CACGCGCTGCTCTCGACCTACGGCGCGGAACGCGACCCGGACGATCCACTGTGGATCGGT
24403 -------+---------+---------+---------+---------+---------+-- 24462
H A L L S T Y G A E R D P D D P L W I G -
TCGGTCAAGTCCAACATTGGCCACACCCAGGCCGCCGCCGGCGTCGCCGGGGTGATGAAG
24463 -------+---------+---------+---------+---------+---------+-- 24522
S V K S N I G H T Q A A A G V A G V M K -
GCGGTGCTGGCGCTGCGGCACGGCGAGATGCCGCGCACGCTGCACTTCGACGAGCCCTCG
24523 -------+---------+---------+---------+---------+---------+-- 24582
A V L A L R H G E M P R T L H F D E P S -
CCGCAGATCGAGTGGGACCTGGGCGCGGTGTCGGTGGTGTCGCAGGCGCGGTCGTGGCCC
24583 -------+---------+---------+---------+---------+---------+-- 24642
P Q I E W D L G A V S V V S Q A R S W P -
GCCGGCGAGAGGCCCCGCAGGGCGGGCGTCTCCTCGTTCGGCATCAGCGGCACCAACGCG
24643 -------+---------+---------+---------+---------+---------+-- 24702

FIG.2-64

```
      A  G  E  R  P  R  R  A  G  V  S  S  F  G  I  S  G  T  N  A  -

      CACGTCATCGTCGAAGAGGCGCCCGAGGCCGACGAGCCCGAGCCGGCACCCGACTCGGGT
24703 -------+---------+---------+---------+---------+---------+--- 24762

      H  V  I  V  E  E  A  P  E  A  D  E  P  E  P  A  P  D  S  G  -

      CCGGTCCCGCTGGTGTTGTCCGGCCGCGACGAGCAGGCGATGCGGGCGCAGGCGGGACGG
24763 -------+---------+---------+---------+---------+---------+--- 24822

      P  V  P  L  V  L  S  G  R  D  E  Q  A  M  R  A  Q  A  G  R  -

      CTGGCAGACCACCTCGCCCGCGAGCCGCGGAACTCGTTGCGCGACACCGGTtTCACGCTG
24823 -------+---------+---------+---------+---------+---------+--- 24882

      L  A  D  H  L  A  R  E  P  R  N  S  L  R  D  T  G  F  T  L  -

      GCCACCCGCCGCAGCGCGTGGGAGCACCGCGCGGTGGTGGTCGGCGACCGCGACGACGCC
24883 -------+---------+---------+---------+---------+---------+--- 24942

      A  T  R  R  S  A  W  E  H  R  A  V  V  V  G  D  R  D  D  A  -

      CTCGCCGGGCTGCGCGCGGTGGCCGACGGCCGCATCGCCGACCGGACGGCCACCGGGCAG
24943 -------+---------+---------+---------+---------+---------+--- 25002

      L  A  G  L  R  A  V  A  D  G  R  I  A  D  R  T  A  T  G  Q  -

      GCCCGAACTCGCCGCGGCGTCGCGATGGTGTTCCCCGGCCAGGGCGCGCAGTGGCAGGGG
25003 -------+---------+---------+---------+---------+---------+--- 25062
```

FIG.2-65

SUBSTITUTE SHEET

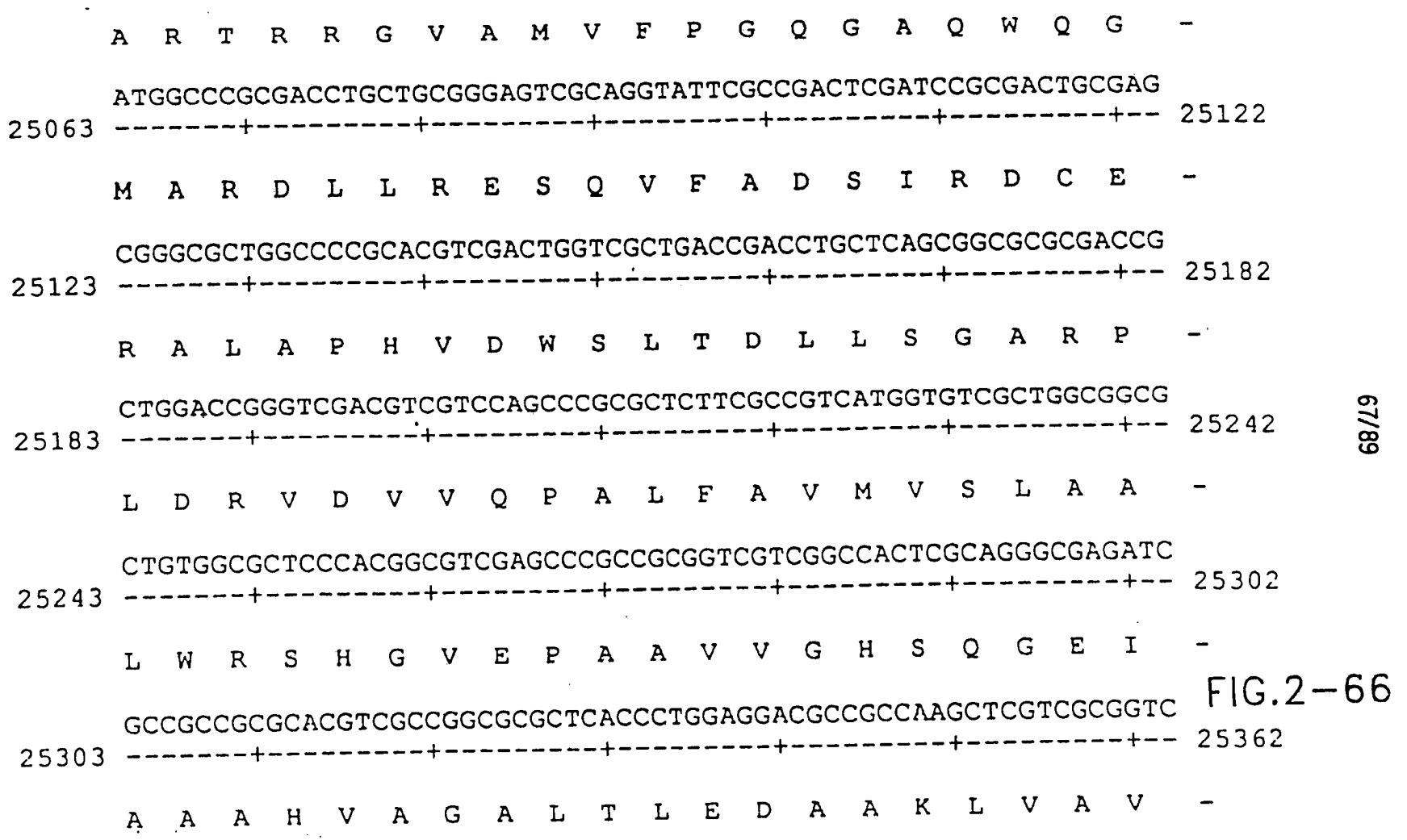

FIG.2-66

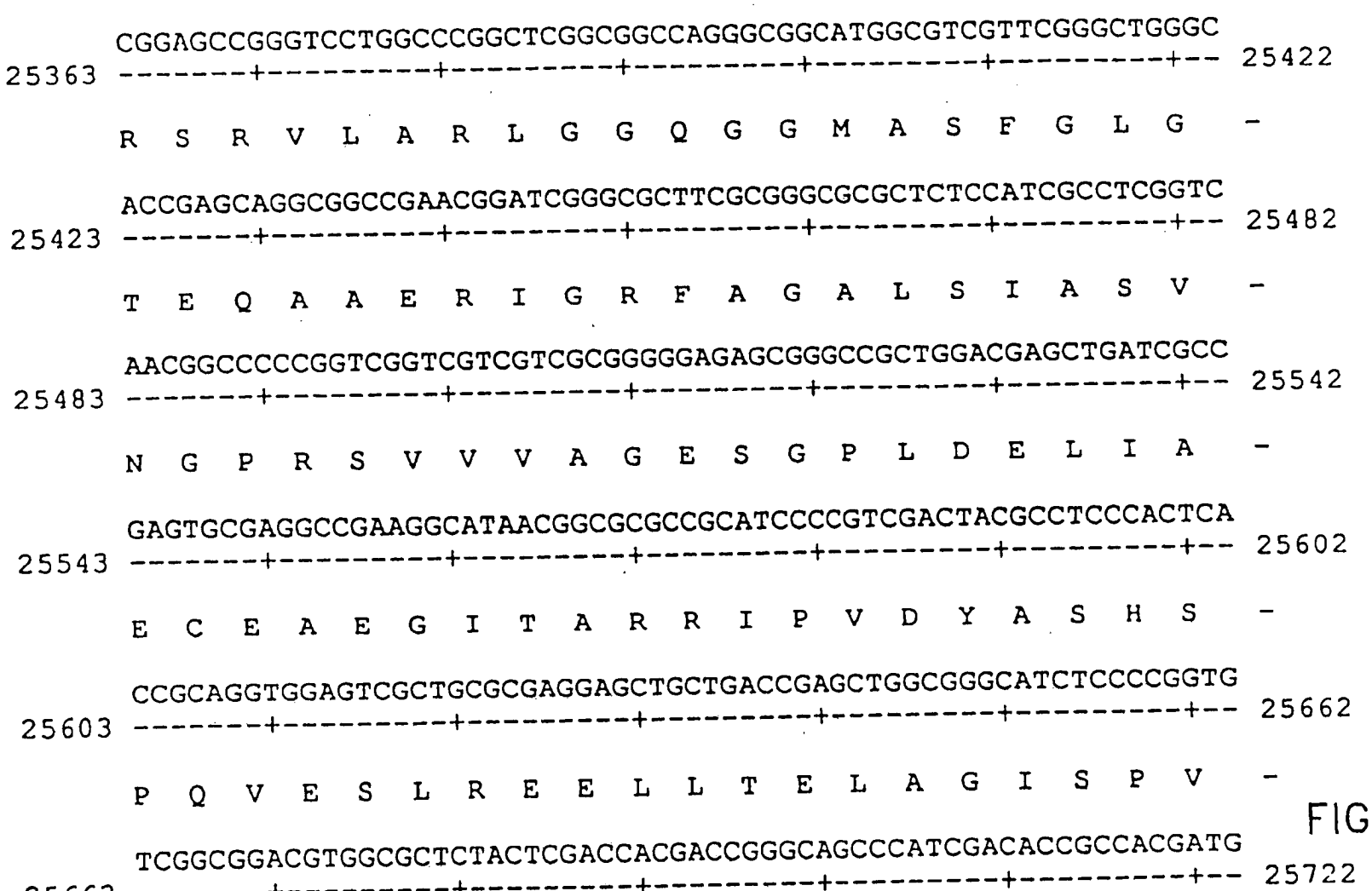

FIG.2-67

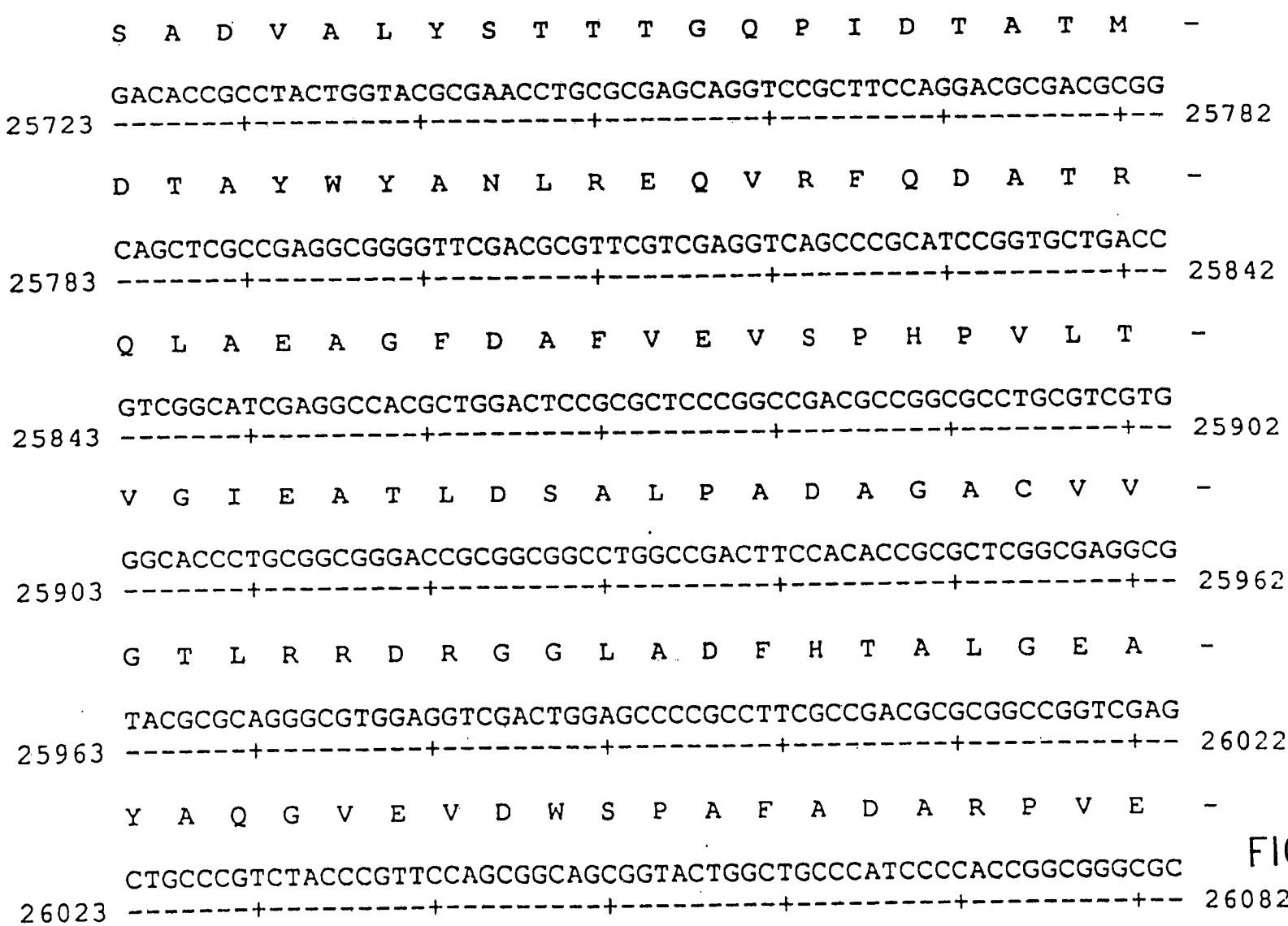
S A D V A L Y S T T T G Q P I D T A T M -
GACACCGCCTACTGGTACGCGAACCTGCGCGAGCAGGTCCGCTTCCAGGACGCGACGCGG
25723 -------+---------+---------+---------+---------+---------+-- 25782
D T A Y W Y A N L R E Q V R F Q D A T R -
CAGCTCGCCGAGGCGGGGTTCGACGCGTTCGTCGAGGTCAGCCCGCATCCGGTGCTGACC
25783 -------+---------+---------+---------+---------+---------+-- 25842
Q L A E A G F D A F V E V S P H P V L T -
GTCGGCATCGAGGCCACGCTGGACTCCGCGCTCCCGGCCGACGCCGGCGCCTGCGTCGTG
25843 -------+---------+---------+---------+---------+---------+-- 25902
V G I E A T L D S A L P A D A G A C V V -
GGCACCCTGCGGCGGGACCGCGGCGGCCTGGCCGACTTCCACACCGCGCTCGGCGAGGCG
25903 -------+---------+---------+---------+---------+---------+-- 25962
G T L R R D R G G L A D F H T A L G E A -
TACGCGCAGGGCGTGGAGGTCGACTGGAGCCCCGCCTTCGCCGACGCGCGGCCGGTCGAG
25963 -------+---------+---------+---------+---------+---------+-- 26022
Y A Q G V E V D W S P A F A D A R P V E -
CTGCCCGTCTACCCGTTCCAGCGGCAGCGGTACTGGCTGCCCATCCCCACCGGCGGGCGC
26023 -------+---------+---------+---------+---------+---------+-- 26082

FIG.2-68

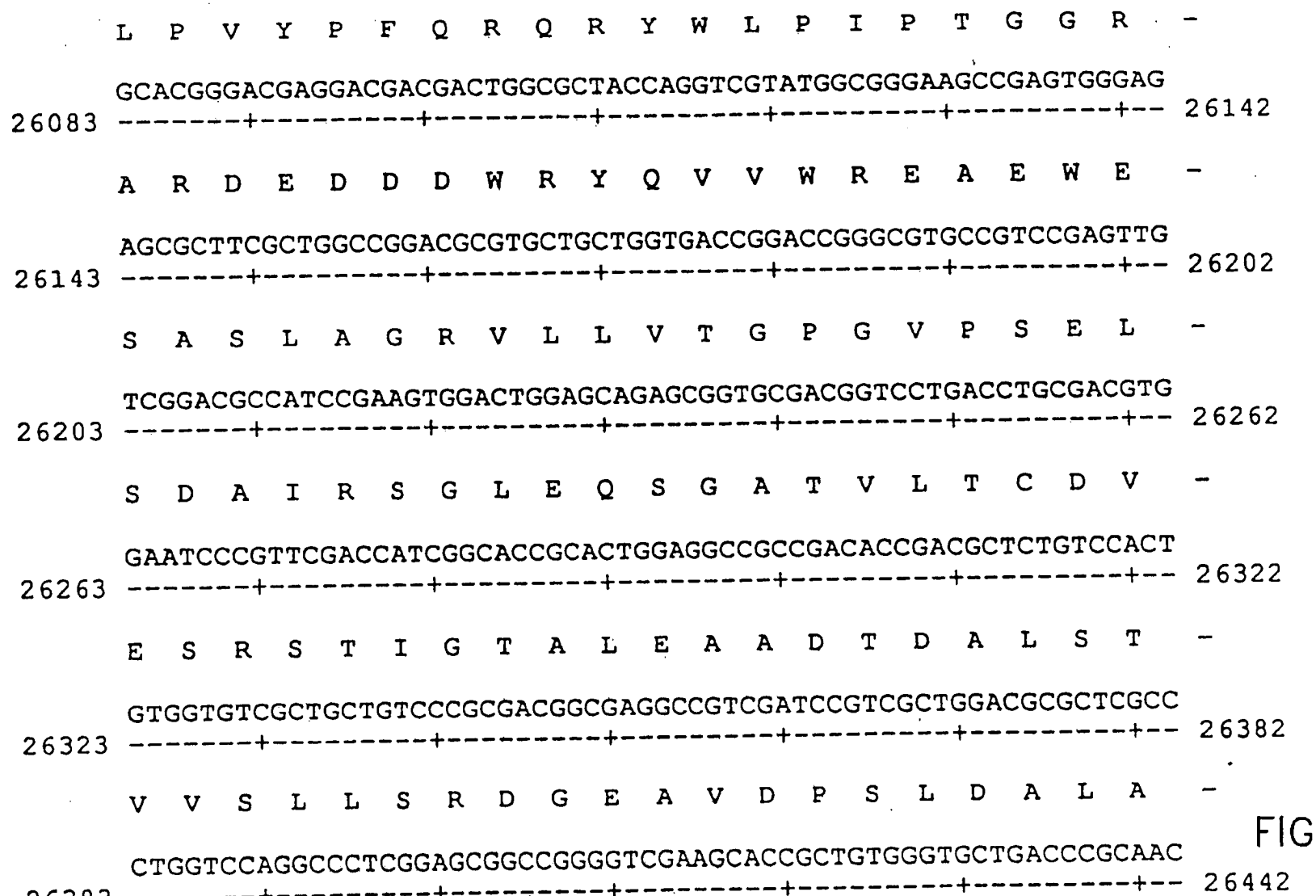

FIG.2-69

SUBSTITUTE SHEET

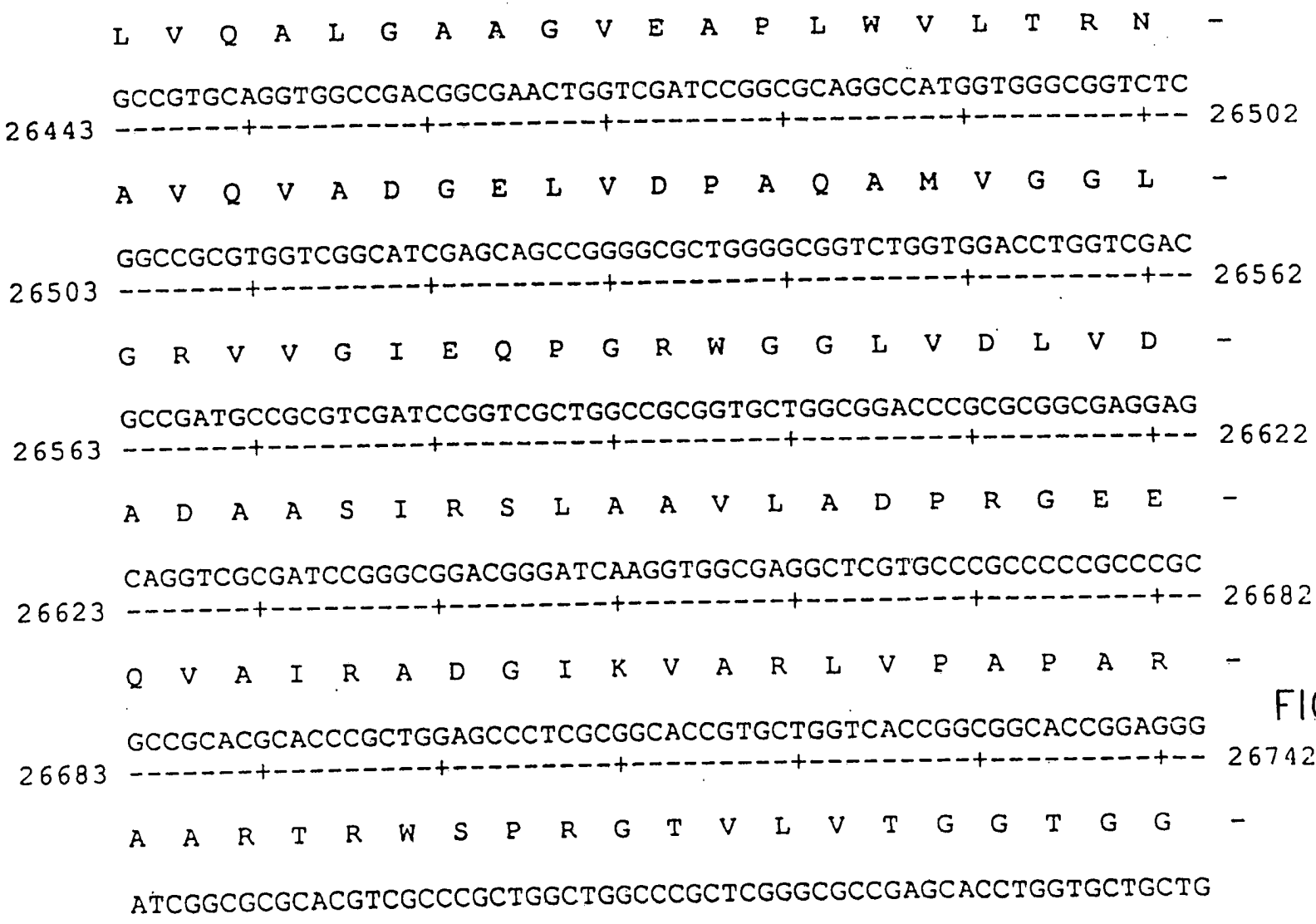

FIG.2-70

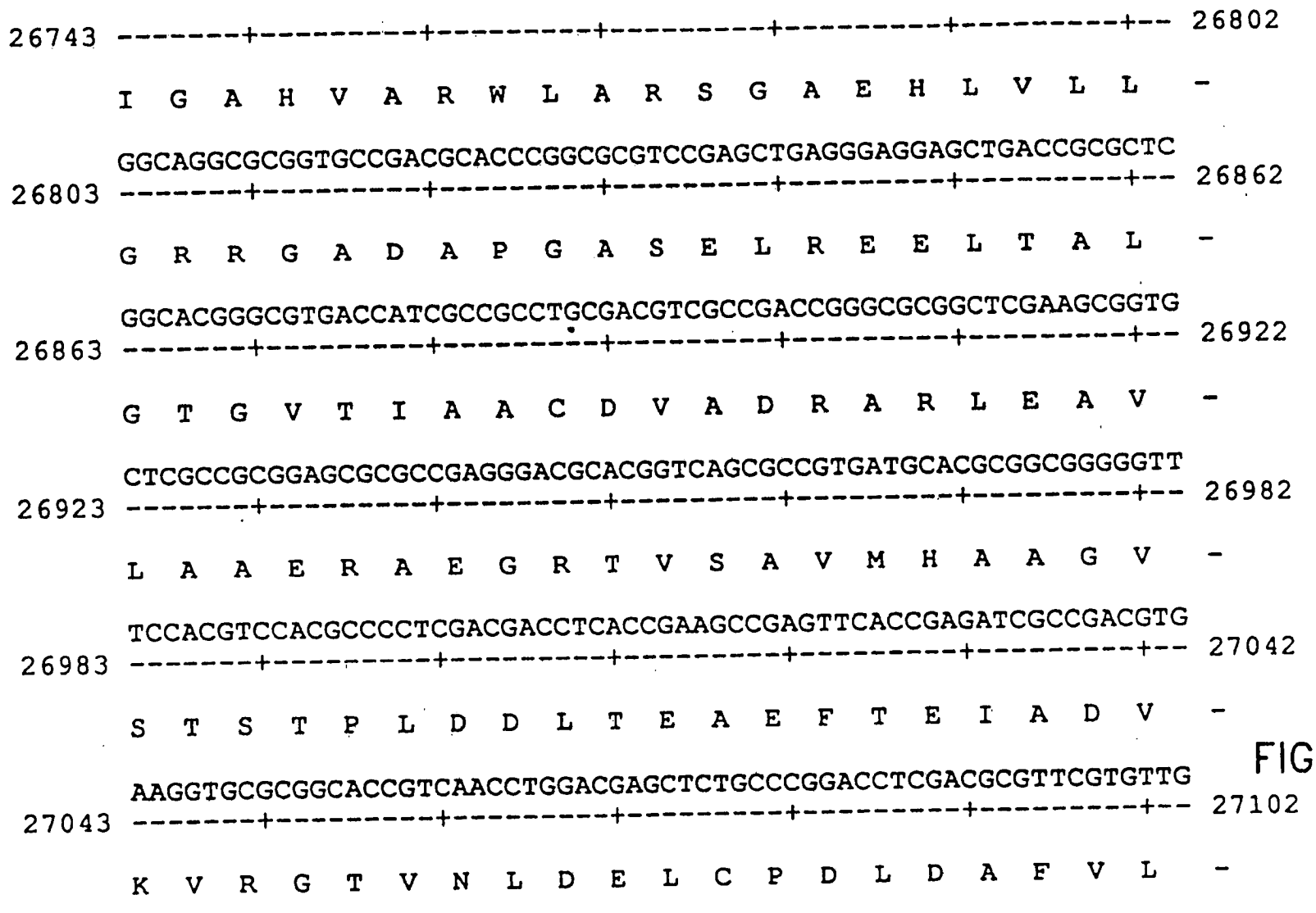

FIG.2-71

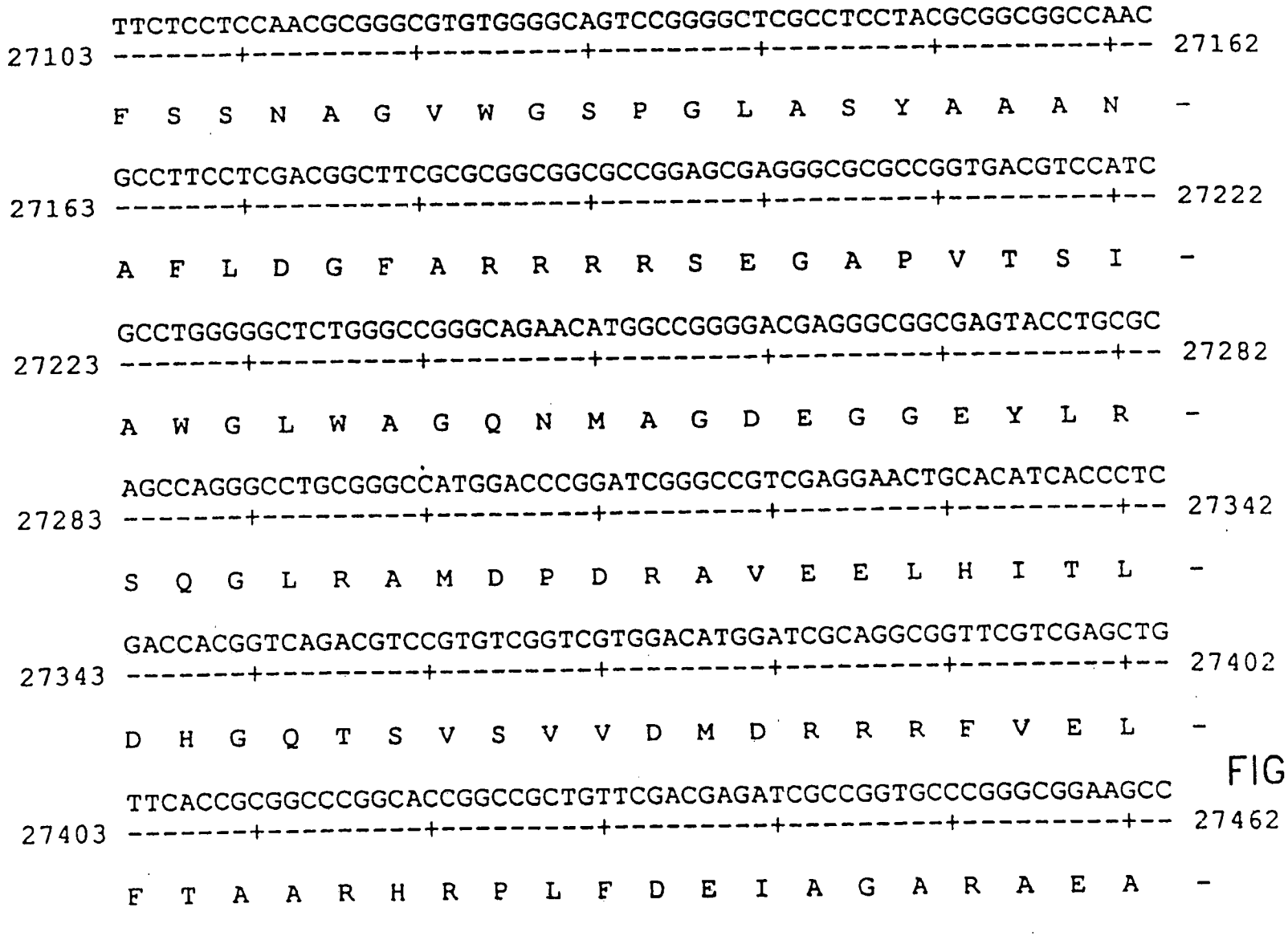

FIG.2-72

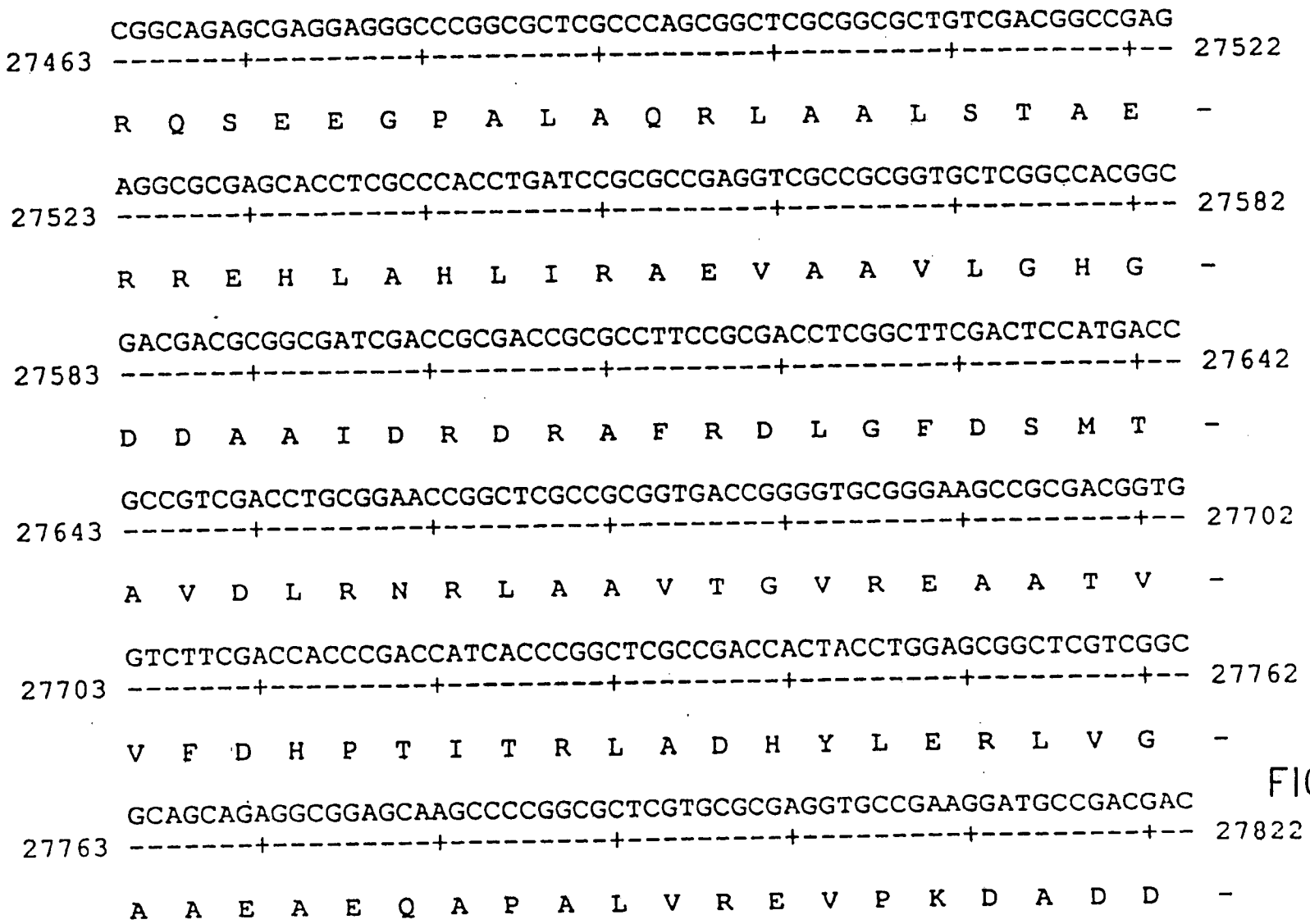

FIG.2-73

```
      CCGATCGCGATCGTCGGCATGGCCTGCCGCTTCCCCGGCGGCGTGCACAACCCCGGTGAG
27823 -------+---------+---------+---------+---------+---------+-- 27882
      P  I  A  I  V  G  M  A  C  R  F  P  G  G  V  H  N  P  G  E  -

      CTGTGGGAGTTCATCGTCGGCCGCGGAGACGCCGTGACGGAGATGCCCACCGACCGCGGC
27883 -------+---------+---------+---------+---------+---------+-- 27942
      L  W  E  F  I  V  G  R  G  D  A  V  T  E  M  P  T  D  R  G  -

      TGGGACCTCGACGCGCTGTTCGACCCCGACCCGCAGCGCCACGGAACCAGCTACTCGCGA
27943 -------+---------+---------+---------+---------+---------+-- 28002
      W  D  L  D  A  L  F  D  P  D  P  Q  R  H  G  T  S  Y  S  R  -

      CACGGCGCGTTCCTCGACGGGGCCGCCGACTTCGACGCGGCGTTCTTCGGGATCTCGCCG
28003 -------+---------+---------+---------+---------+---------+-- 28062
      H  G  A  F  L  D  G  A  A  D  F  D  A  A  F  F  G  I  S  P  -

      CGCGAGGCGCTGGCGATGGACCCGCAGCAGCGCCAGGTCCTGGAAACGACGTGGGAGCTG
28063 -------+---------+---------+---------+---------+---------+-- 28122
      R  E  A  L  A  M  D  P  Q  Q  R  Q  V  L  E  T  T  W  E  L  -

      TTCGAGAACGCCGGCATCGACCCGCACTCGCTGCGGGGCAGCGACACCGGCGTCTTCCTC
28123 -------+---------+---------+---------+---------+---------+-- 28182
```

FIG.2-74

```
      F  E  N  A  G  I  D  P  H  S  L  R  G  S  D  T  G  V  F  L  -

      GGCGCCGCGTACCAGGGCTACGGCCAGGACGCGGTGGTGCCCGAGGACAGCGAGGGCTAC
28183 -------+---------+---------+---------+---------+---------+-- 28242

      G  A  A  Y  Q  G  Y  G  Q  D  A  V  V  P  E  D  S  E  G  Y  -

      CTGCTCACCGGCAACTCCTCCGCCGTGGTGTCCGGCCGGGTCGCCTACGTGCTGGGGCTG
28243 -------+---------+---------+---------+---------+---------+-- 28302

      L  L  T  G  N  S  S  A  V  V  S  G  R  V  A  Y  V  L  G  L  -

      GAAGGCCCCGCGGTCACGGTGGACACGGCGTGTTCGTCGTCGTTGGTGGCCTTGCATTCG
28303 -------+---------+---------+---------+---------+---------+-- 28362

      E  G  P  A  V  T  V  D  T  A  C  S  S  S  L  V  A  L  H  S  -

      GCGTGTGGGTCGTTGCGTGACGGTGACTGCGGTCTTGCGGTGGCCGGTGGTGTGTCGGTG
28363 -------+---------+---------+---------+---------+---------+-- 28422

      A  C  G  S  L  R  D  G  D  C  G  L  A  V  A  G  G  V  S  V  -

      ATGGCGGGCCCGGAGGTGTTCACCGAGTTCTCCCGCCAGGGCGGCTTGGCCGTGGACGGG
28423 -------+---------+---------+---------+---------+---------+-- 28482

      M  A  G  P  E  V  F  T  E  F  S  R  Q  G  G  L  A  V  D  G  -

      CGCTGCAAGGCGTTCTCCGCGGAGGCCGACGGCTTCGGTTTCGCCGAGGGCGTCGCGGTG
28483 -------+---------+---------+---------+---------+---------+-- 28542
```

FIG.2-75

SUBSTITUTE SHEET

```
      R  C  K  A  F  S  A  E  A  D  G  F  G  F  A  E  G  V  A  V  -

      GTCCTGCTCCAGCGGTTGTCCGACGCCCGCAGGGCGGGTCGCCAGGTGCTCGGCGTGGTC
28543 -------+---------+---------+---------+---------+---------+-- 28602

      V  L  L  Q  R  L  S  D  A  R  R  A  G  R  Q  V  L  G  V  V  -

      GCGGGCTCGGCGATCAACCAGGACGGCGCGAGCAACGGTCTCGCGGCGCCGAGCGGCGTC
28603 -------+---------+---------+---------+---------+---------+-- 28662

      A  G  S  A  I  N  Q  D  G  A  S  N  G  L  A  A  P  S  G  V  -

      GCCCAGCAGCGCGTGATCCGCAAGGCGTGGGCGCGTGCGGGGATCACCGGCGCGGATGTG
28663 -------+---------+---------+---------+---------+---------+-- 28722

      A  Q  Q  R  V  I  R  K  A  W  A  R  A  G  I  T  G  A  D  V  -

      GCCGTGGTGGAGGCGCATGGGACCGGTACGCGGCTGGGCGATCCGGTGGAGGCGTCGGCG
28723 -------+---------+---------+---------+---------+---------+-- 28782

      A  V  V  E  A  H  G  T  G  T  R  L  G  D  P  V  E  A  S  A  -

      TTGCTGGCTACTTACGGCAAGTCGCGCGGGTCGTCGGGCCCGGTGCTGCTGGGTTCGGTG
28783 -------+---------+---------+---------+---------+---------+-- 28842

      L  L  A  T  Y  G  K  S  R  G  S  S  G  P  V  L  L  G  S  V  -
```

FIG.2-76

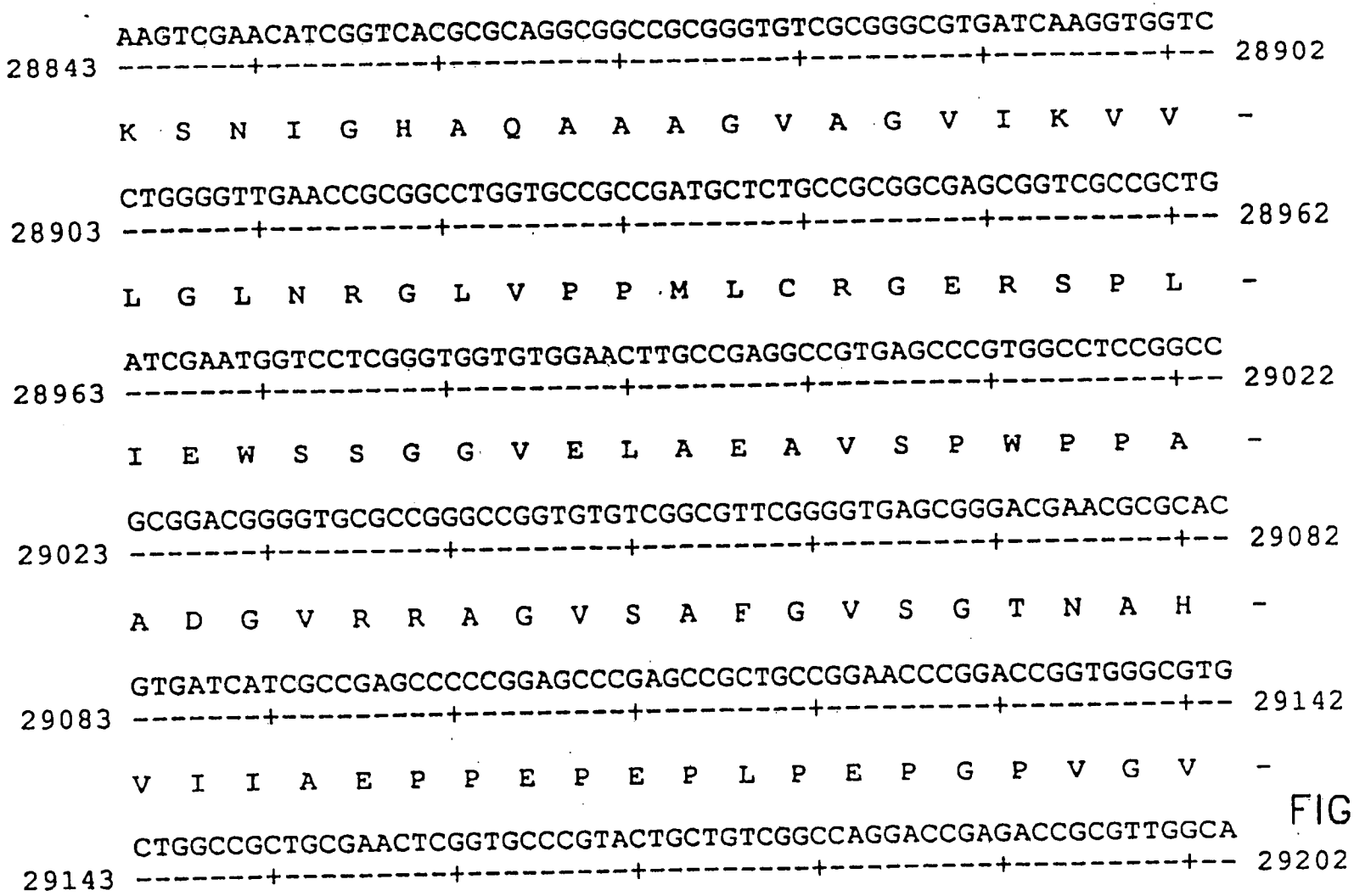

FIG.2-77

SUBSTITUTE SHEET

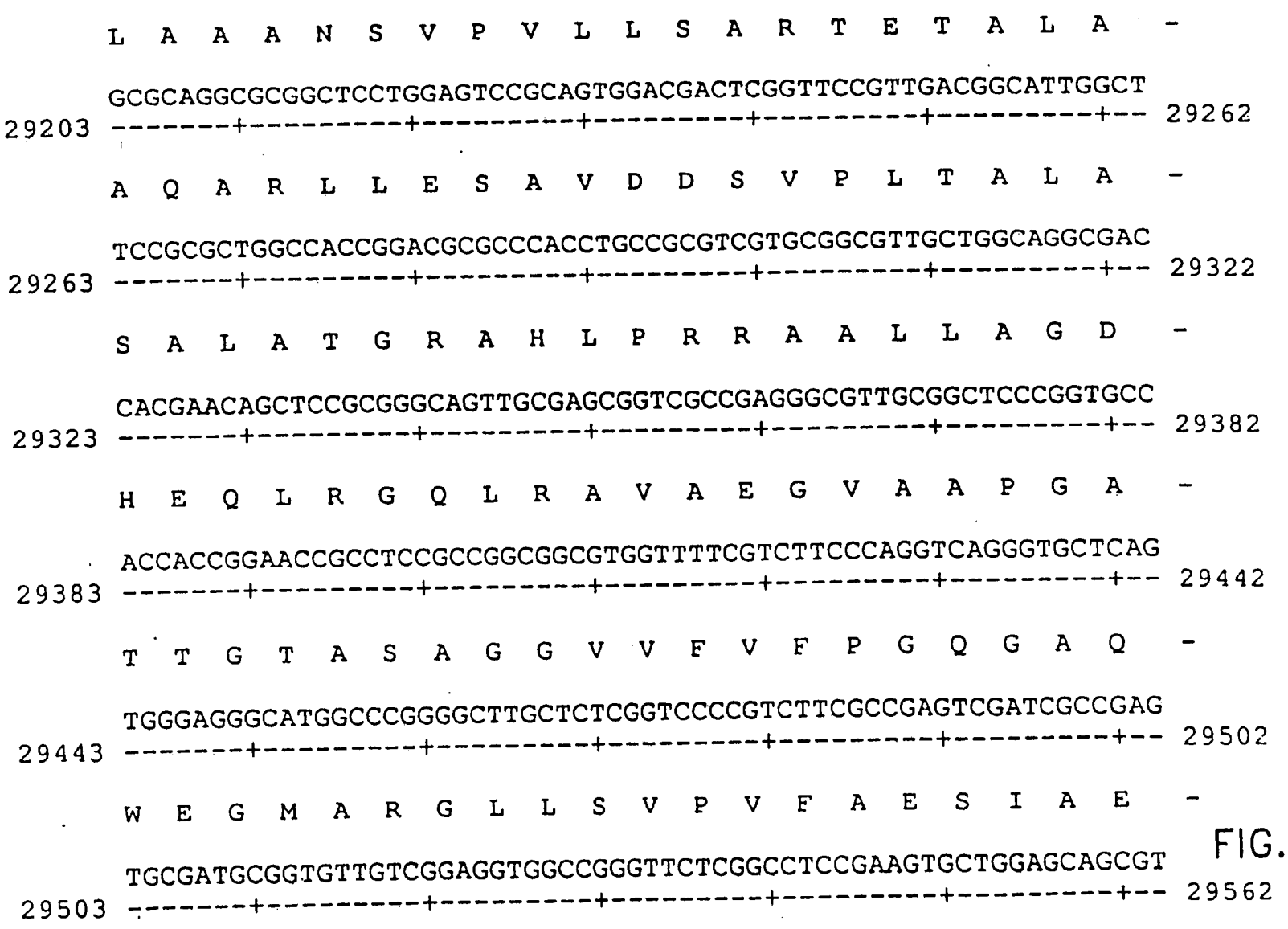
L A A A N S V P V L L S A R T E T A L A -
GCGCAGGCGCGGCTCCTGGAGTCCGCAGTGGACGACTCGGTTCCGTTGACGGCATTGGCT
29203 -------+---------+---------+---------+---------+---------+-- 29262
A Q A R L L E S A V D D S V P L T A L A -
TCCGCGCTGGCCACCGGACGCGCCCACCTGCCGCGTCGTGCGGCGTTGCTGGCAGGCGAC
29263 -------+---------+---------+---------+---------+---------+-- 29322
S A L A T G R A H L P R R A A L L A G D -
CACGAACAGCTCCGCGGGCAGTTGCGAGCGGTCGCCGAGGGCGTTGCGGCTCCCGGTGCC
29323 -------+---------+---------+---------+---------+---------+-- 29382
H E Q L R G Q L R A V A E G V A A P G A -
ACCACCGGAACCGCCTCCGCCGGCGGCGTGGTTTTCGTCTTCCCAGGTCAGGGTGCTCAG
29383 -------+---------+---------+---------+---------+---------+-- 29442
T T G T A S A G G V V F V F P G Q G A Q -
TGGGAGGGCATGGCCCGGGGCTTGCTCTCGGTCCCCGTCTTCGCCGAGTCGATCGCCGAG
29443 -------+---------+---------+---------+---------+---------+-- 29502
W E G M A R G L L S V P V F A E S I A E -
TGCGATGCGGTGTTGTCGGAGGTGGCCGGGTTCTCGGCCTCCGAAGTGCTGGAGCAGCGT
29503 -------+---------+---------+---------+---------+---------+-- 29562

FIG.2-78

```
      C  D  A  V  L  S  E  V  A  G  F  S  A  S  E  V  L  E  Q  R  -

      CCGGACGCGCCGTCGCTGGAGCGGGTCGACGTCGTACAGCCGGTGTTGTTCTCCGTGATG
29563 -------+---------+---------+---------+---------+---------+-- 29622

      P  D  A  P  S  L  E  R  V  D  V  V  Q  P  V  L  F  S  V  M  -

      GTGTCGCTGGCGCGGCTGTGGGGCGCTTGCGGAGTCAGCCCCTCGGCCGTCATCGGCCAT
29623 -------+---------+---------+---------+---------+---------+-- 29682

      V  S  L  A  R  L  W  G  A  C  G  V  S  P  S  A  V  I  G  H  -

      TCGCAGGGCGAGATCGCCGCCGCGGTGGTGGCCGGGGTGTTGTCGCTGGAGGACGGCGTG
29683 -------+---------+---------+---------+---------+---------+-- 29742

      S  Q  G  E  I  A  A  A  V  V  A  G  V  L  S  L  E  D  G  V  -

      CGCGTCGTGGCCCTGCGCGCGAAGGCGTTGCGTGCGCTGGCGGGCAAGGGCGGCATGGTC
29743 -------+---------+---------+---------+---------+---------+-- 29802

      R  V  V  A  L  R  A  K  A  L  R  A  L  A  G  K  G  G  M  V  -

      TCGTTGGCGGCTCCCGGTGAACGCGCCCGCGCGCTGATCGCACCGTGGGAGGACCGGATC
29803 -------+---------+---------+---------+---------+---------+-- 29862

      S  L  A  A  P  G  E  R  A  R  A  L  I  A  P  W  E  D  R  I  -

      TCCGTCGCGGCGGTCAACTCCCCGTCCTCGGTCGTGGTCTCCGGCGATCCGGAGGCGCTG
29863 -------+---------+---------+---------+---------+---------+-- 29922
```

FIG.2-79

```
     S  V  A  A  V  N  S  P  S  S  V  V  V  S  G  D  P  E  A  L  -

     GCCGAACTCGTCGCACGTTGCGAGGACGAGGGCGTGCGCGCCAAGACGCTCCCGGTGGAC
29923 -------+---------+---------+---------+---------+---------+-- 29982

     A  E  L  V  A  R  C  E  D  E  G  V  R  A  K  T  L  P  V  D  -

     TACGCCTCGCACTCCCGCCACGTCGAGGAGATCCGCGAGACGATCCTCGCCGACCTCGAC
29983 -------+---------+---------+---------+---------+---------+-- 30042

     Y  A  S  H  S  R  H  V  E  E  I  R  E  T  I  L  A  D  L  D  -

     GGCATCTCCGCGCGGCGTGCCGCCATCCCGCTCTACTCCACGCTGCACGGCGAACGGCGC
30043 -------+---------+---------+---------+---------+---------+-- 30102

     G  I  S  A  R  R  A  A  I  P  L  Y  S  T  L  H  G  E  R  R  -

     GACGGCGCCGACATGGGTCCGCGGTACTGGTACGACAACCTGCGCTCCCAGGTGCGCTTC
30103 -------+---------+---------+---------+---------+---------+-- 30162

     D  G  A  D  M  G  P  R  Y  W  Y  D  N  L  R  S  Q  V  R  F  -

     GACGAGGCGGTCTCGGCCGCCGTCGCCGACGGTCACGCCACCTTCGTCGAGATGAGCCCG
30163 -------+---------+---------+---------+---------+---------+-- 30222

     D  E  A  V  S  A  A  V  A  D  G  H  A  T  F  V  E  M  S  P  -

     CACCCGGTGCTCACCGCGGCGGTGCAGGAGATCGCCGCGGACGCCGTGGCCATCGGGTCG
```

FIG.2-80

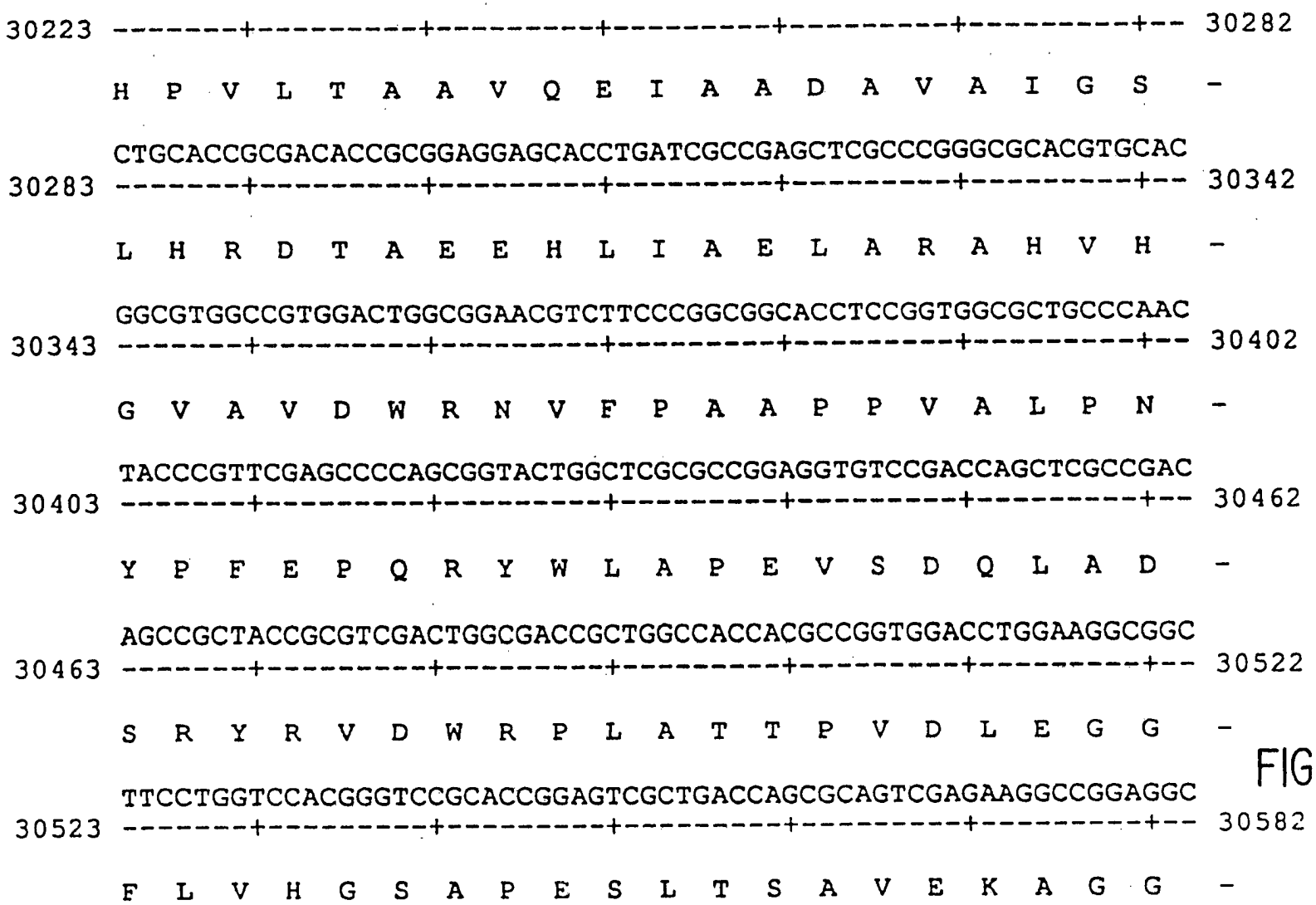

FIG.2-81

```
      CGCGTCGTGCCGGTCGCCTCGGCCGACCGCGAAGCCTCGGCGGCCCTGCGGGAGGTGCCG
30583 -------+---------+---------+---------+---------+---------+-- 30642

      R  V  V  P  V  A  S  A  D  R  E  A  S  A  A  L  R  E  V  P  -

      GGCGAGGTCGCCGGCGTGCTCTCGGTCCACACCGGCGCCGCAACGCACCTCGCCCTGCAC
30643 -------+---------+---------+---------+---------+---------+-- 30702

      G  E  V  A  G  V  L  S  V  H  T  G  A  A  T  H  L  A  L  H  -

      CAGTCGCTGGGTGAGGCCGGCGTGCGGGCCCCGCTCTGGCTGGTCACCAGCCGAGCGGTC
30703 -------+---------+---------+---------+---------+---------+-- 30762

      Q  S  L  G  E  A  G  V  R  A  P  L  W  L  V  T  S  R  A  V  -

      GCGCTCGGGGAGTCCGAGCCGGTCGATCCCGAGCAGGCGATGGTGTGGGGTCTCGGGCGC
30763 -------+---------+---------+---------+---------+---------+-- 30822

      A  L  G  E  S  E  P  V  D  P  E  Q  A  M  V  W  G  L  G  R  -

      GTCATGGGCCTGGAGACCCCGGAACGGTGGGGCGGTCTGGTGGACCTGCCCGCCGAACCC
30823 -------+---------+---------+---------+---------+---------+-- 30882

      V  M  G  L  E  T  P  E  R  W  G  G  L  V  D  L  P  A  E  P  -

      GCGCCGGGGGACGGCGAGGCGTTCGTCGCCTGCCTCGGCGCGGACGGCCACGAGGACCAG
30883 -------+---------+---------+---------+---------+---------+-- 30942

      A  P  G  D  G  E  A  F  V  A  C  L  G  A  D  G  H  E  D  Q  -
```

FIG.2-82

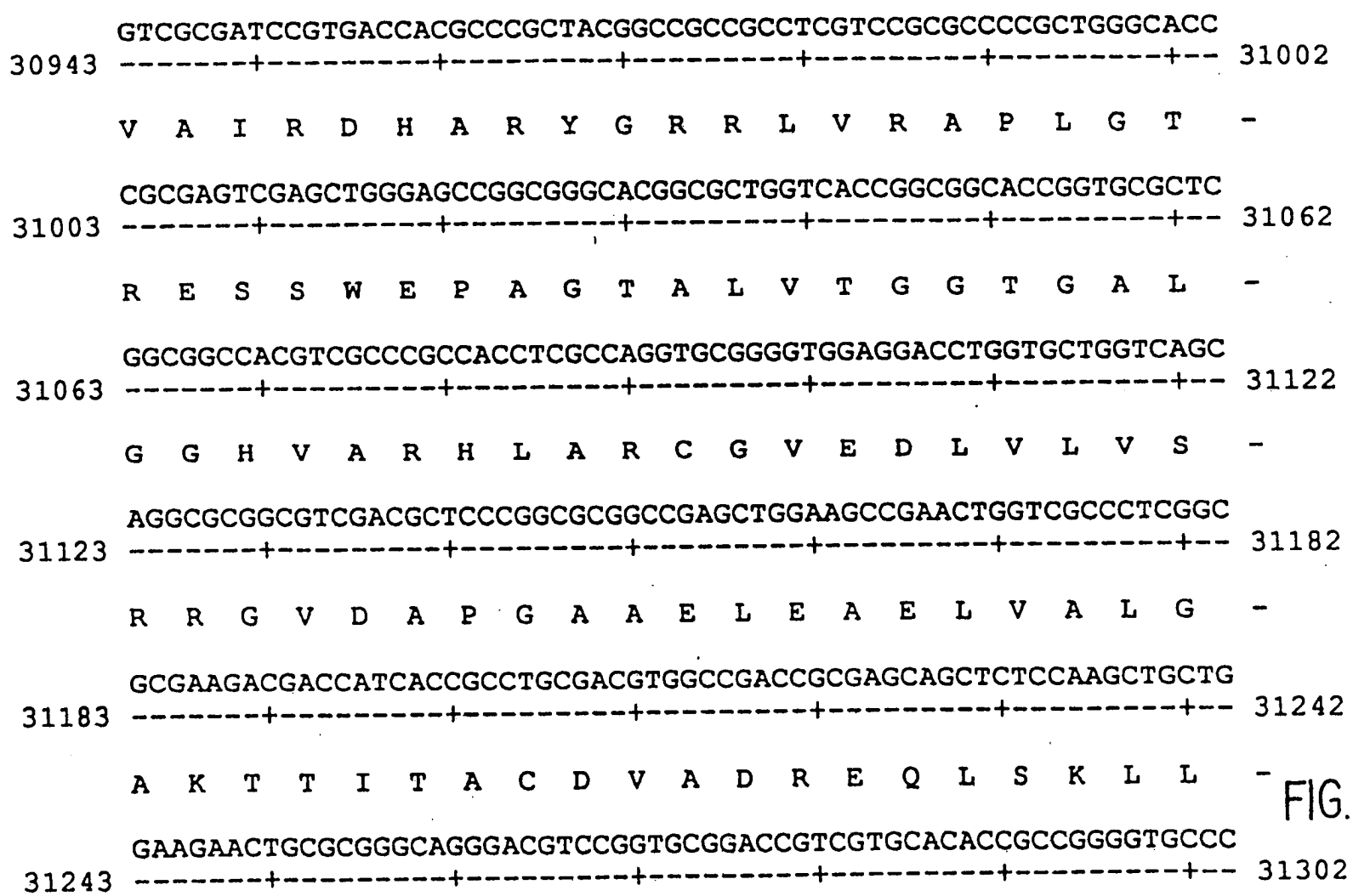

FIG.2-83

```
      E  E  L  R  G  Q  G  R  P  V  R  T  V  V  H  T  A  G  V  P  -

      GAATCGAGGCCGCTGCACGAGATCGGCGAGCTGGAGTCGGTCTGCGCGGCGAAGGTGACC
31303 -------+---------+---------+---------+---------+---------+-- 31362

      E  S  R  P  L  H  E  I  G  E  L  E  S  V  C  A  A  K  V  T  -

      GGGGCCCGGCTGCTCGACGAGCTGTGCCCGGACGCCGAGACCTTCGTCCTGTTCTCGTCC
31363 -------+---------+---------+---------+---------+---------+-- 31422

      G  A  R  L  L  D  E  L  C  P  D  A  E  T  F  V  L  F  S  S  -

      GGAGCGGGGGTGTGGGGCAGTGCGAACCTCGGCGCCTACTCCGCGGCCAACGCCTACCTC
31423 -------+---------+---------+---------+---------+---------+-- 31482

      G  A  G  V  W  G  S  A  N  L  G  A  Y  S  A  A  N  A  Y  L  -

      GACGCGCTGGCCCACCGCCGCCGTGCGGAAGGCCGTGCGGCGACGTCCGTCGCGTGGGGC
31483 -------+---------+---------+---------+---------+---------+-- 31542

      D  A  L  A  H  R  R  R  A  E  G  R  A  A  T  S  V  A  W  G  -

      GCCTGGGCGGGCGAGGGCATGGCCACCGGCGACCTCGAGGGGCTCACCCGGCGCGGCCTG
31543 -------+---------+---------+---------+---------+---------+-- 31602

      A  W  A  G  E  G  M  A  T  G  D  L  E  G  L  T  R  R  G  L  -

      CGCCCGATGGCGCCCGAGCGCGCGATCCGCGCGCTGCACCAGGCGCTGGACAACGGCGAC
31603 -------+---------+---------+---------+---------+---------+-- 31662
```

FIG.2-84

```
      R  P  M  A  P  E  R  A  I  R  A  L  H  Q  A  L  D  N  G  D  -

      ACGTGCGTTTCGATCGCCGACGTCGACTGGGAGGCCTTCGCGGTCGGCTTCACCGCCGCC
31663 -------+---------+---------+---------+---------+---------+-- 31722

      T  C  V  S  I  A  D  V  D  W  E  A  F  A  V  G  F  T  A  A  -

      CGGCCGCGTCCGCTGCTGGACGAGCTCGTCACGCCGGCGGTGGGGGCCGTCCCCGCGGTG
31723 -------+---------+---------+---------+---------+---------+-- 31782

      R  P  R  P  L  L  D  E  L  V  T  P  A  V  G  A  V  P  A  V  -

      CAGGCGGCCCCGGCGCGGGAGATGACGTCGCAGGAGTTGCTGGAGTTCACGCACTCGCAC
31783 -------+---------+---------+---------+---------+---------+-- 31842

      Q  A  A  P  A  R  E  M  T  S  Q  E  L  L  E  F  T  H  S  H  -

      GTCGCGGCGATCCTCGGGCATTCCAGCCCGGACGCGGTCGGGCAGGACCAGCCGTTCACC
31843 -------+---------+---------+---------+---------+---------+-- 31902

      V  A  A  I  L  G  H  S  S  P  D  A  V  G  Q  D  Q  P  F  T  -

      GAGCTCGGCTTCGACTCGCTGACCGCGGTCGGGCTGCGCAACCAGCTCCAGCAGGCCACC
31903 -------+---------+---------+---------+---------+---------+-- 31962

      E  L  G  F  D  S  L  T  A  V  G  L  R  N  Q  L  Q  Q  A  T  -

      GGGCTCGCGCTGCCCGCGACCCTGGTGTTCGAGCACCCCACGGTCCGCAGGTTGGCCGAC
31963 -------+---------+---------+---------+---------+---------+-- 32022
```

FIG. 2-85

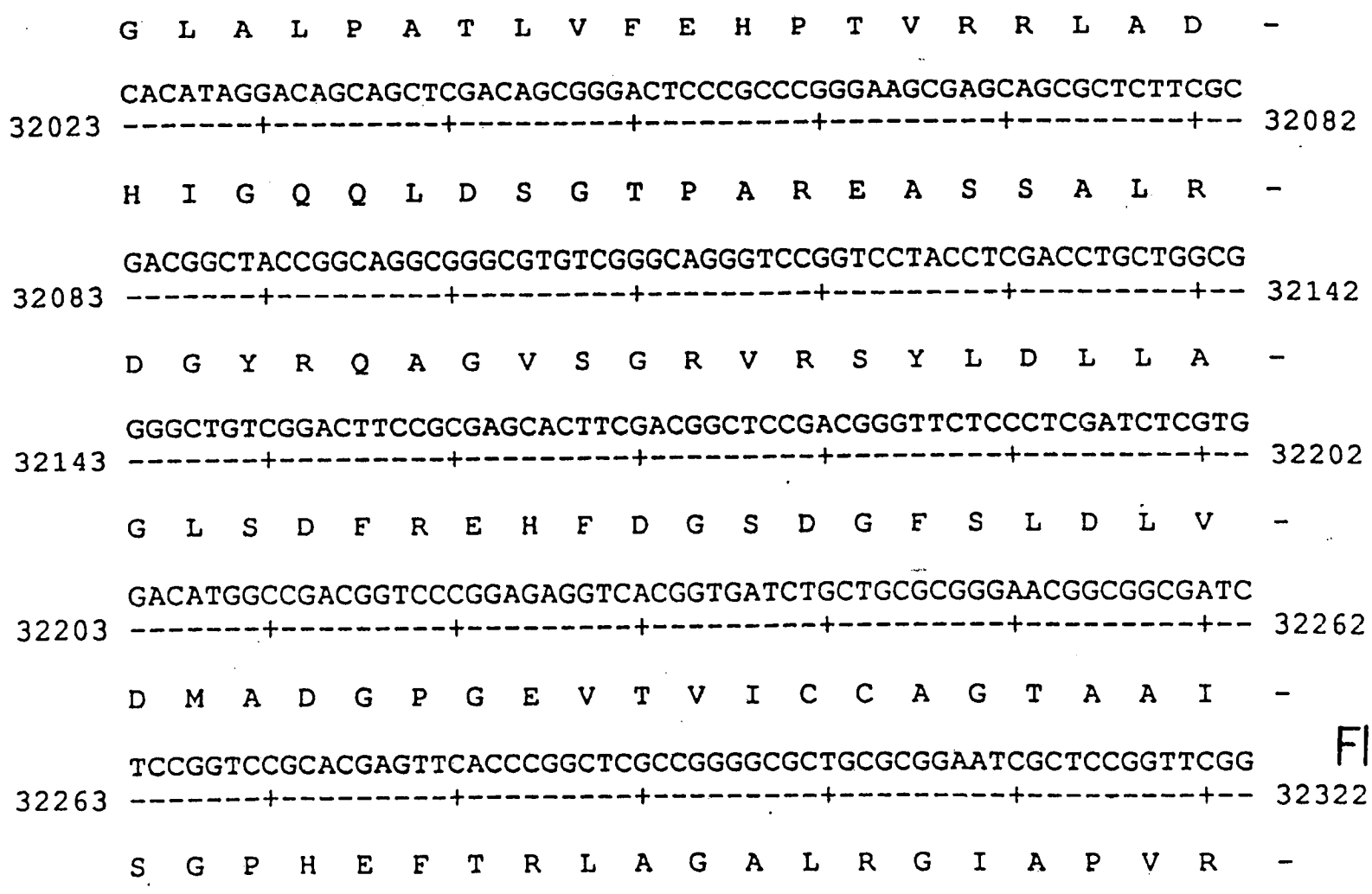

FIG.2-86

SUBSTITUTE SHEET

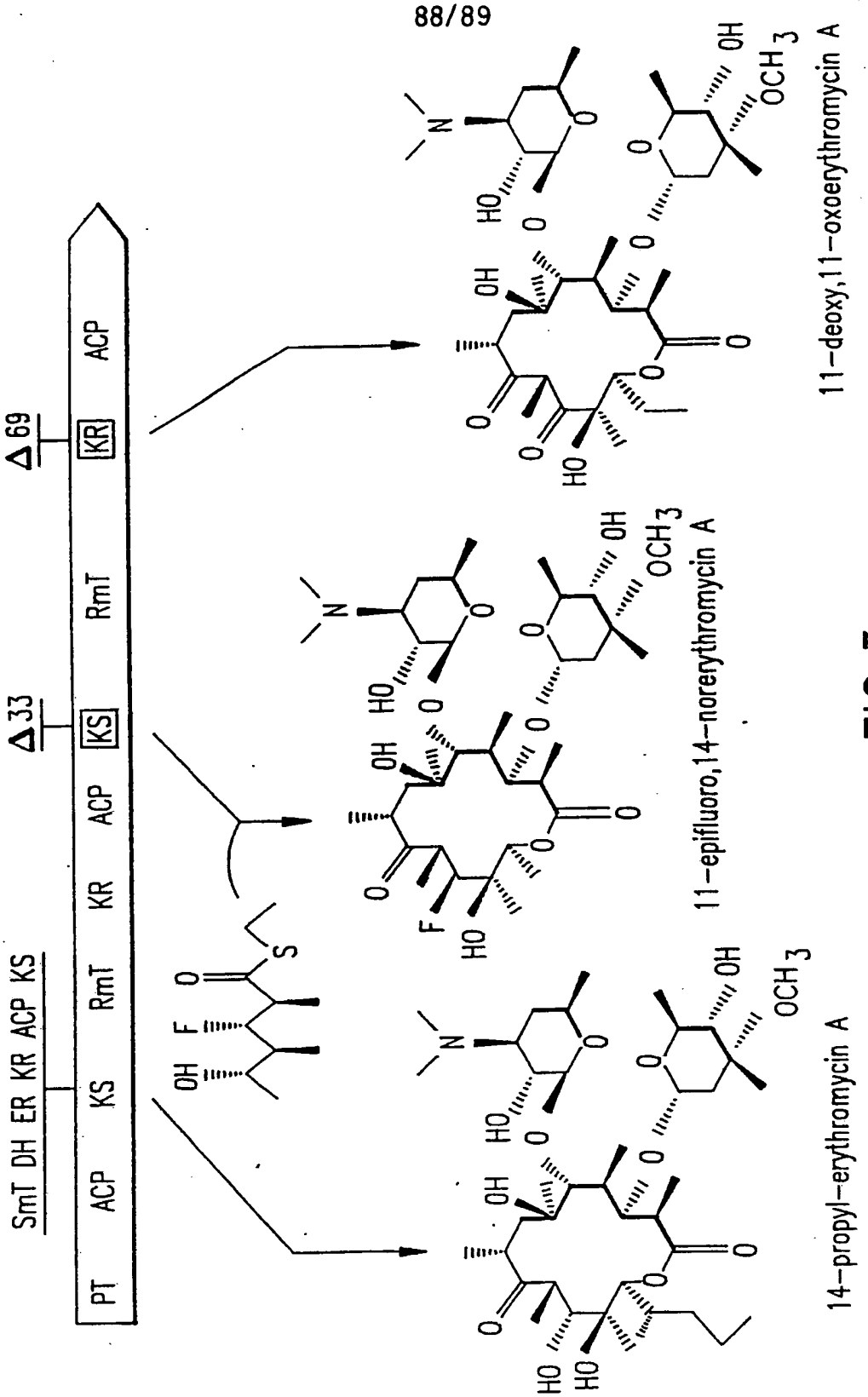
SmT DH ER KR ACP KS
PT ACP KS RmT KR ACP KS RmT KR ACP
Δ33
Δ69
11-deoxy,11-oxoerythromycin A
11-epifluoro,14-norerythromycin A
14-propyl-erythromycin A

FIG.3

NUMBER	SITE	DISTANCE (Kb)[a]
1	BamHI	−3.60
2	PvuII	−3.50
3	PvuII	−3.40
4	PstI	−3.05
5	BamHI	−2.95
6	XhoI	−2.80
7	PstI	−2.00
8	HindII	−1.60
9	SphI	−1.55
10	EcoRI	−1.50
11	KpnI	−1.35
12	EcoRI	−1.05
13	SmaI[b]	−0.90
14	SphI	−0.75
15	KpnI	−0.65
16	SmaI	−0.20

FIG.4

INTERNATIONAL SEARCH REPORT

International Application No. PCT/US92/00427

I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all)[3]

According to International Patent Classification (IPC) or to both National Classification and IPC

IPC (5): A01N 43/22; A61K 31/71; C07H 17/08; C12N 1/21; C12P 19/62
US CL : 435/76, 252.35, 886; 514/29; 536/7.2

II. FIELDS SEARCHED

Minimum Documentation Searched[4]

Classification System	Classification Symbols
U.S.	435/76, 183, 252.35, 886; 514/29; 536/7.2

Documentation Searched other than Minimum Documentation to the extent that such Documents are included in the Fields Searched[6]

APS, BIOSIS, MEDLINE, BIOTECH. ABSTRACTS

III. DOCUMENTS CONSIDERED TO BE RELEVANT[14]

Category*	Citation of Document,[16] with indication, where appropriate, of the relevant passages[17]	Relevant to Claim No.[18]
y	J. of Bacteriology, Volume 164, No. 1, issued October 1985, J.M. Weber et al, "Genetic Analysis of Erythromycin Production in Streptomyces erythreus", pages 425-433, See the entire document.	1-30
y	J. of Bacteriology, Volume 172, No. 5, issued May 1990, J.M. Weber et al, "Organization of a Cluster of Erythromycin Genes in Saccharomyces erythraea", pages 2372-2383, See the entire document.	1-30
y	US, A, 4,874,748 (Katz et al) 17 October 1989, see column 1, lines 41-59; column 3, lines 47-60.	1-30
y	US, A, 4,935,340 (Baltz et al) 19 June 1990, see entire document.	1-29

* Special categories of cited documents:[15]

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

IV. CERTIFICATION

Date of the Actual Completion of the International Search[2]	Date of Mailing of this International Search Report[2]
20 MARCH 1992	30 MAR 1992
International Searching Authority[1]	Signature of Authorized Officer[20]
ISA/US	Dian Cook